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(54) **OBSERVATION METHOD USING BINDING AND DISSOCIATION PROBE**

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(57) **ABSTRACT**

(21) Appl. No.: **15/696,089**

An observation method of a sample containing a target substance, the observation method including an imaging step in which a step of obtaining a speckle image including, as a speckle, light emitted from a luminescent substance in which a medium is brought into contact with the sample is performed a plurality of times so as to obtain a plurality of speckle images, the medium containing a probe that contains the luminescent substance emitting light and that repeatedly binds to and dissociates from the target substance directly and specifically, and an observation image generation step of generating an observation image of the target substance in the sample from the plurality of speckle images, wherein a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.

(22) Filed: **Sep. 5, 2017**

Related U.S. Application Data

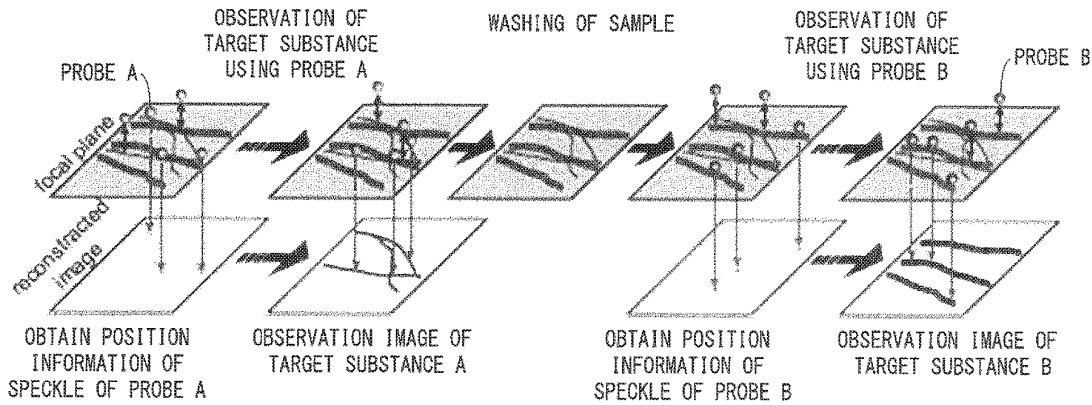
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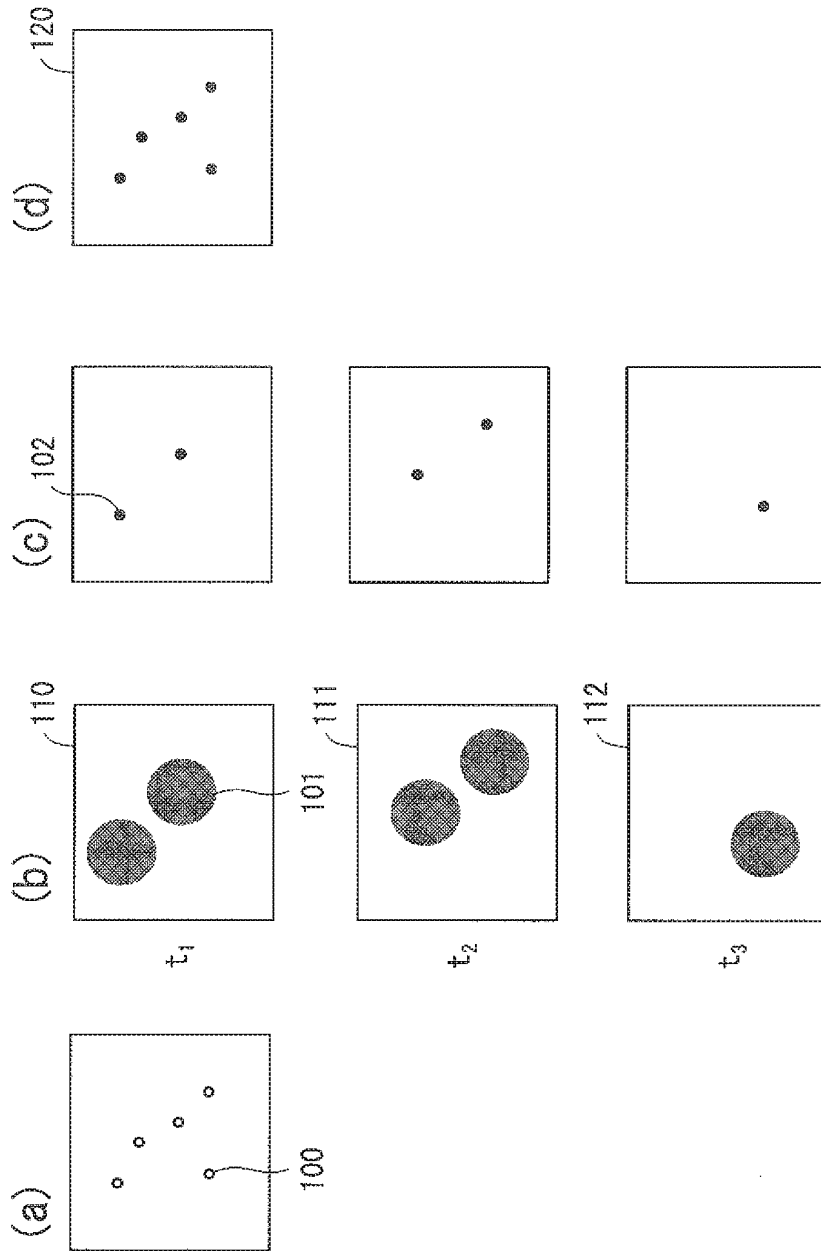


FIG. 1

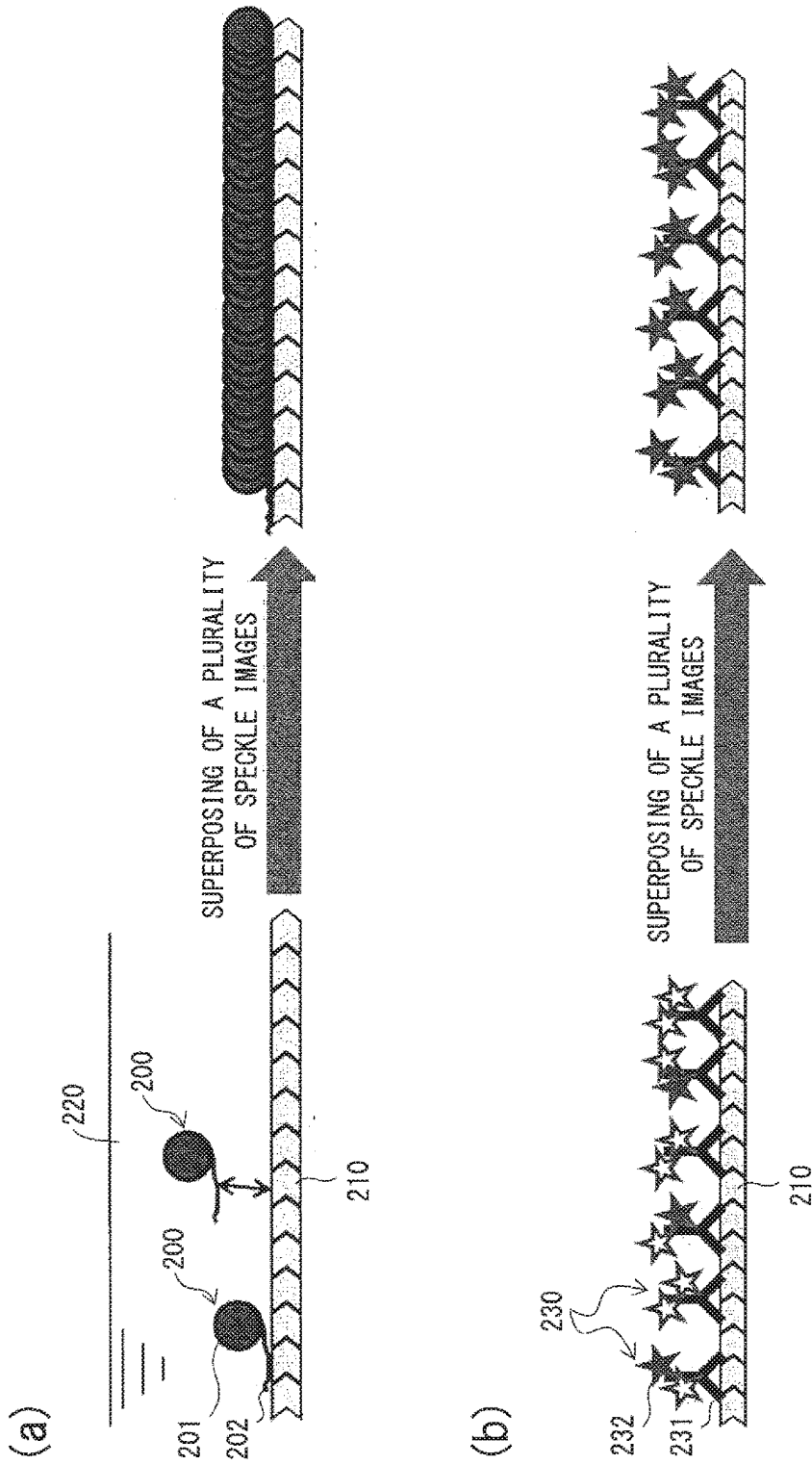


FIG. 2

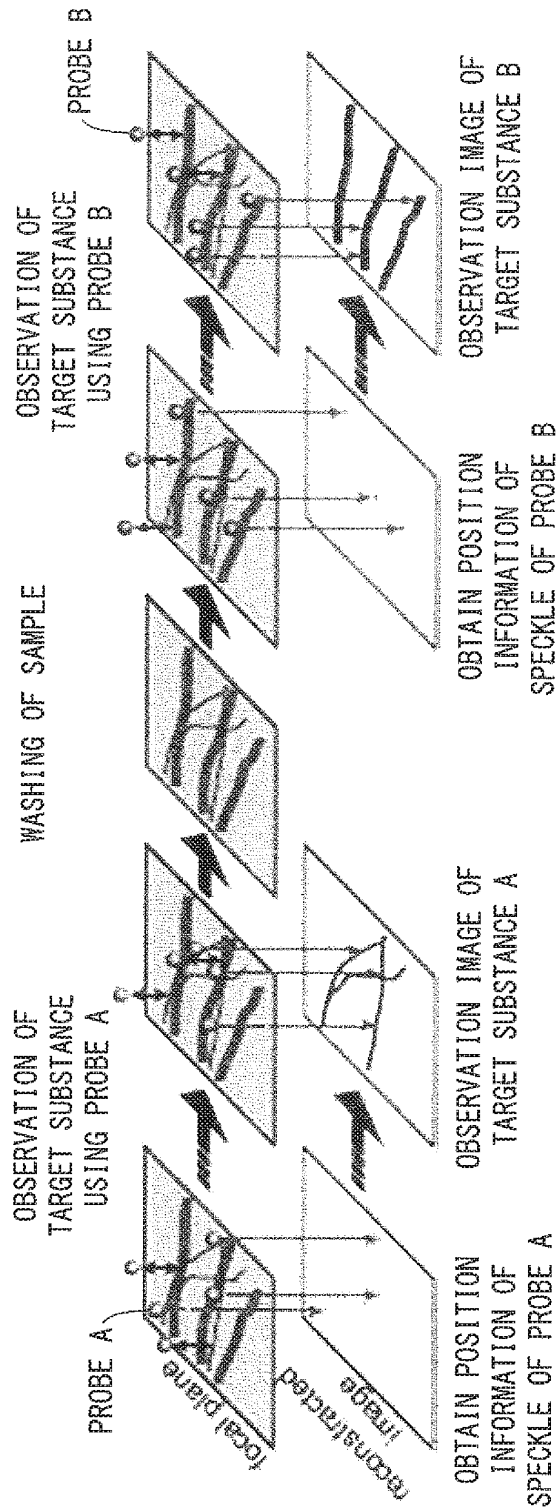


FIG. 3

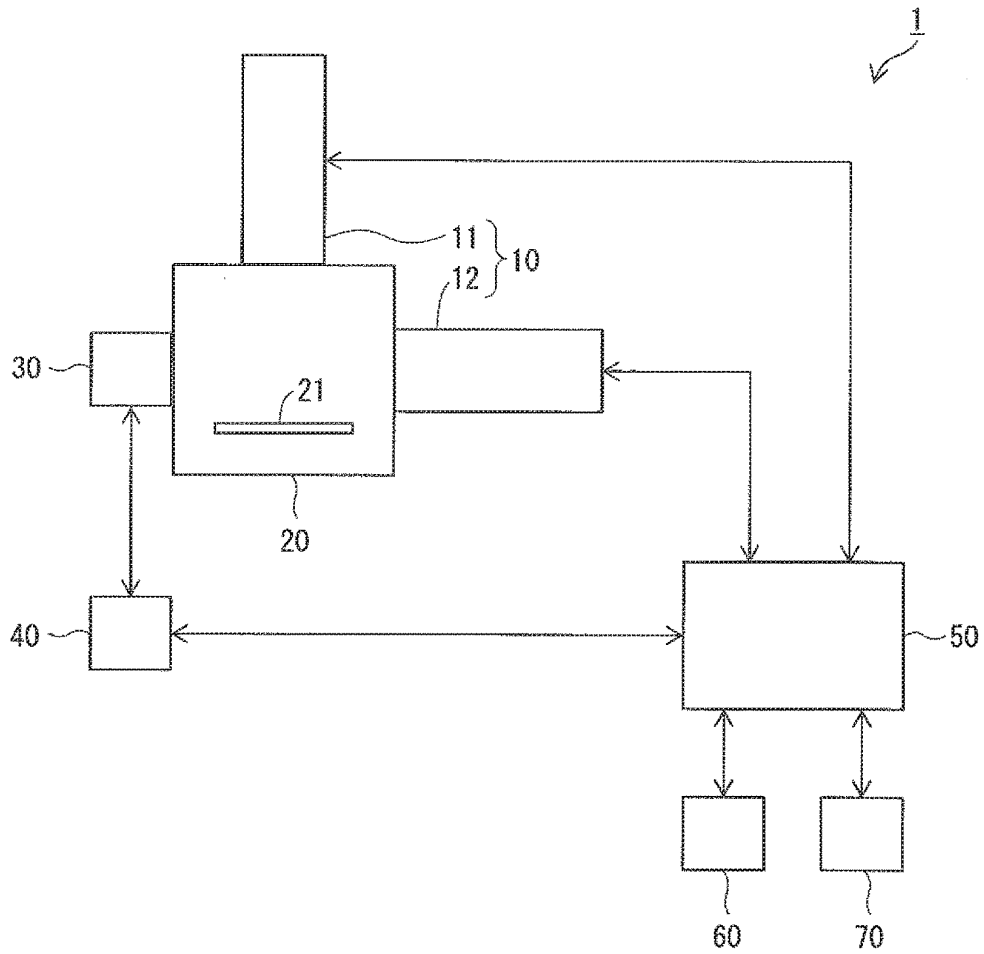


FIG. 4

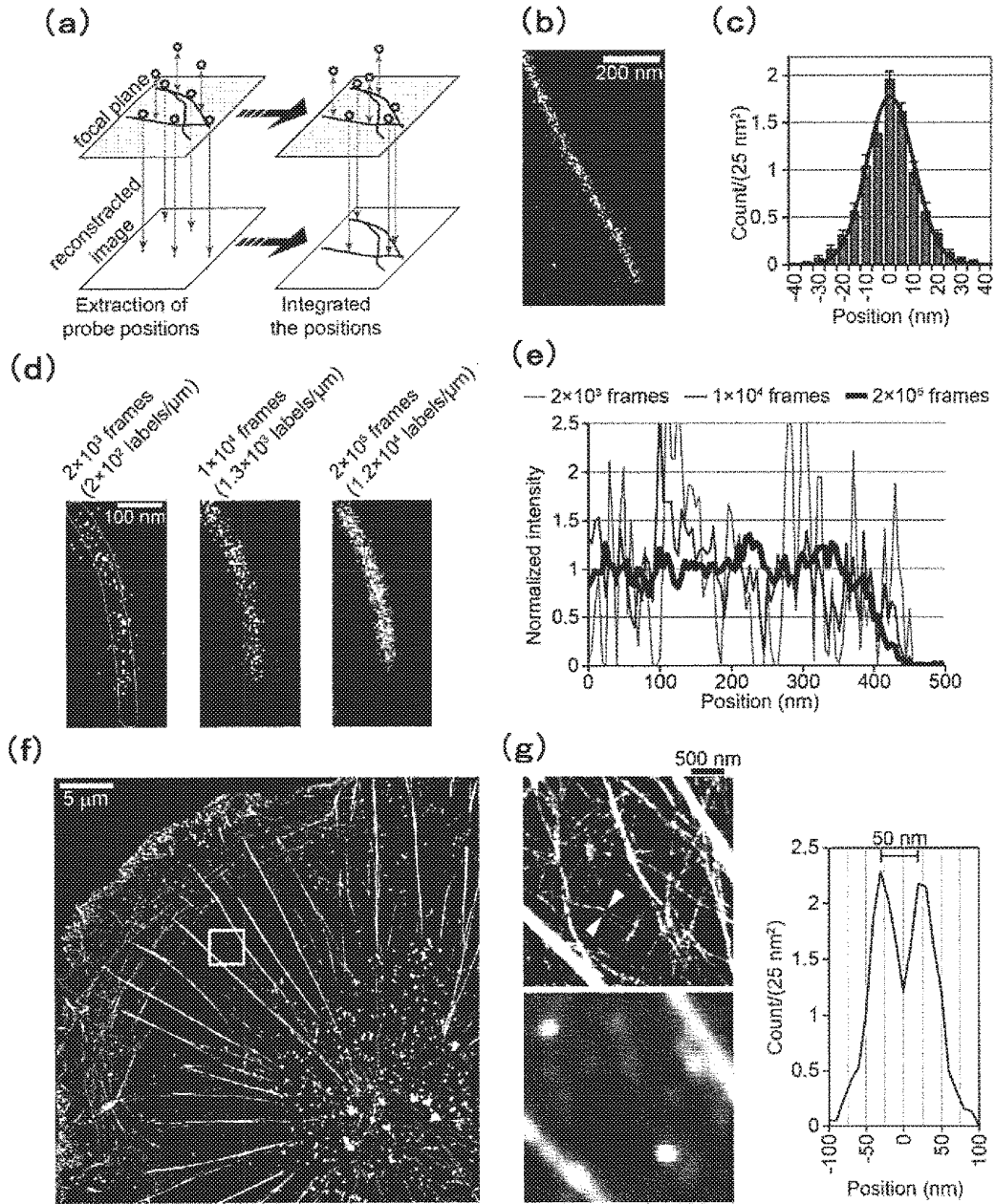


FIG. 5

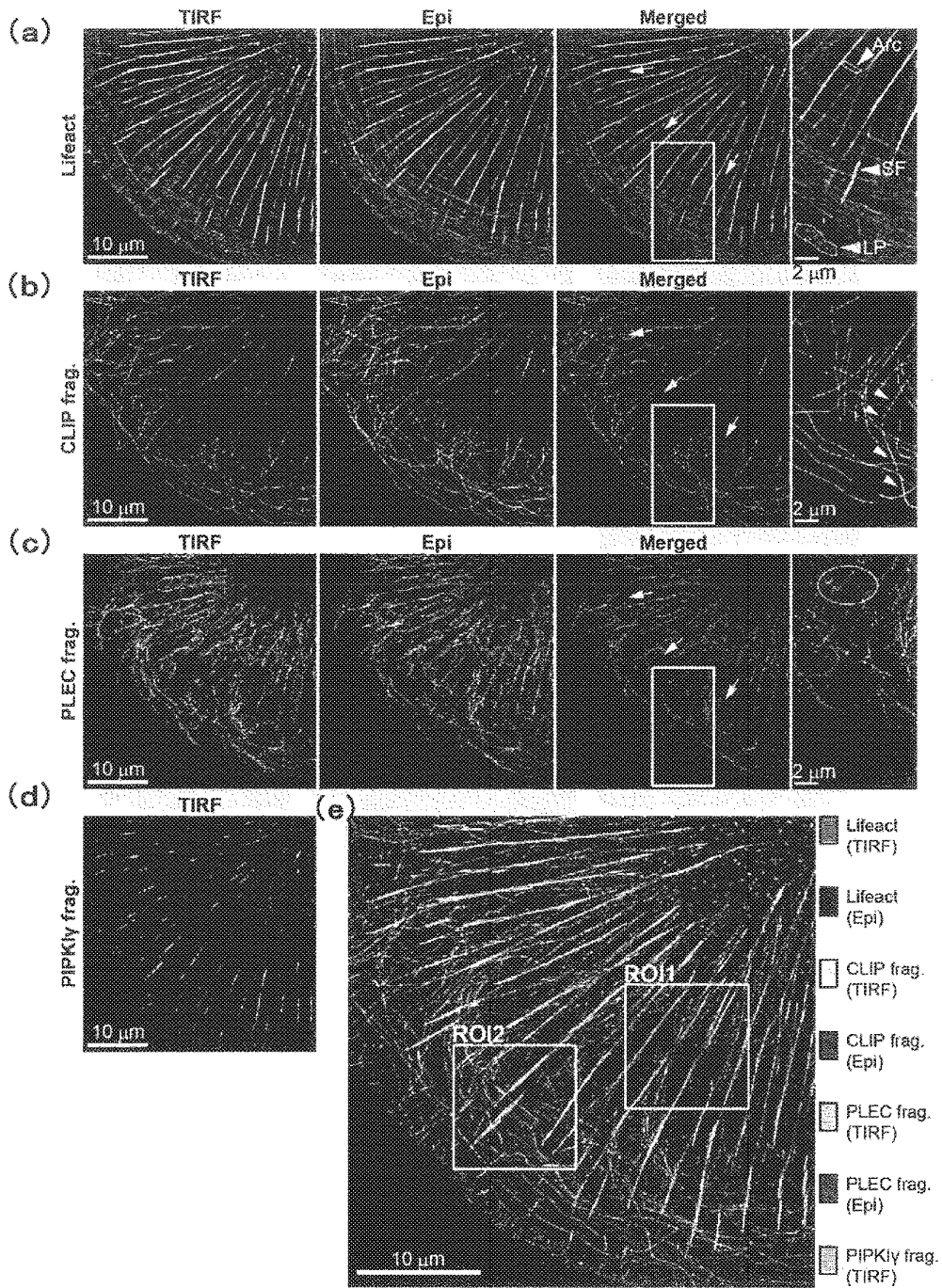


FIG. 6

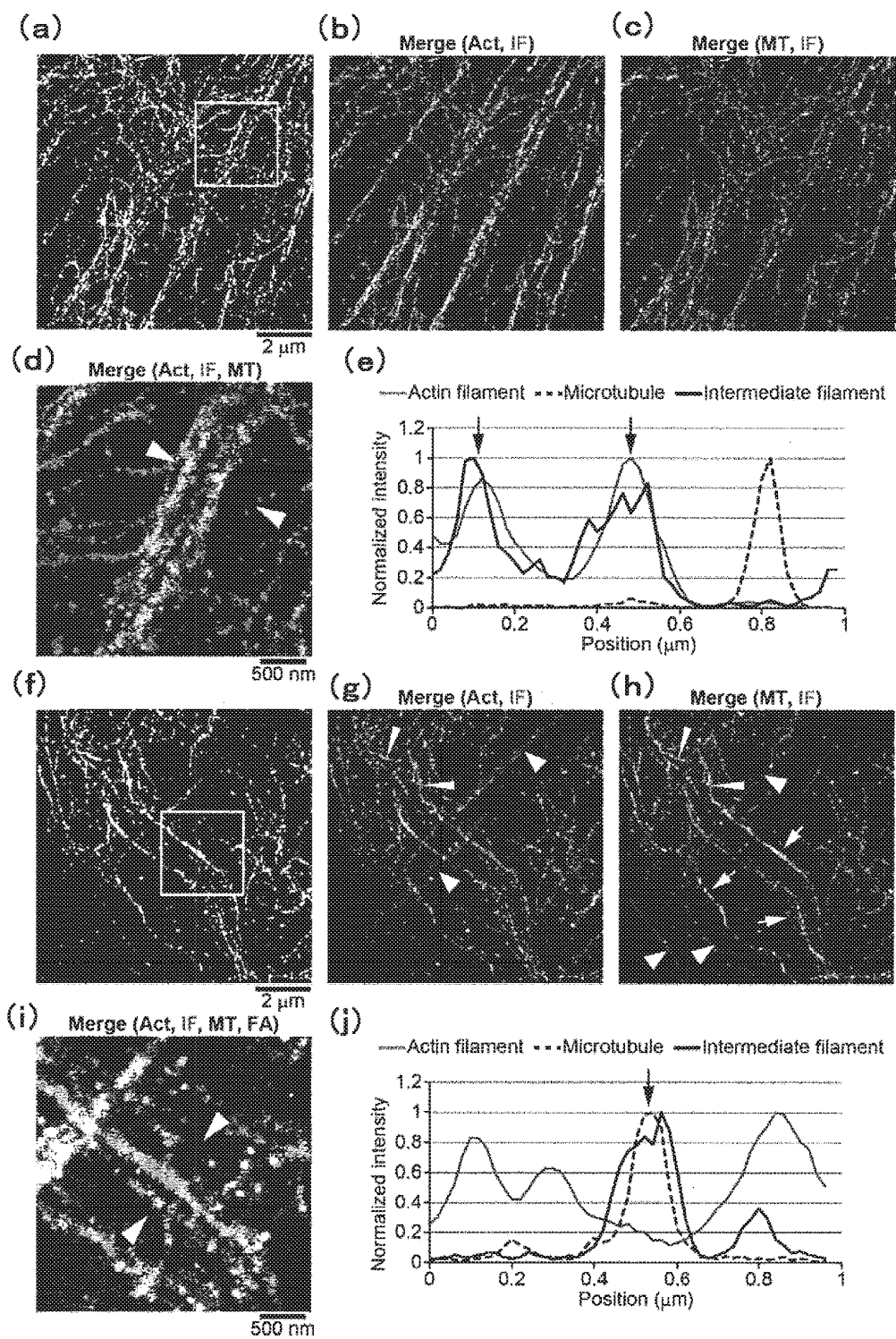


FIG. 7

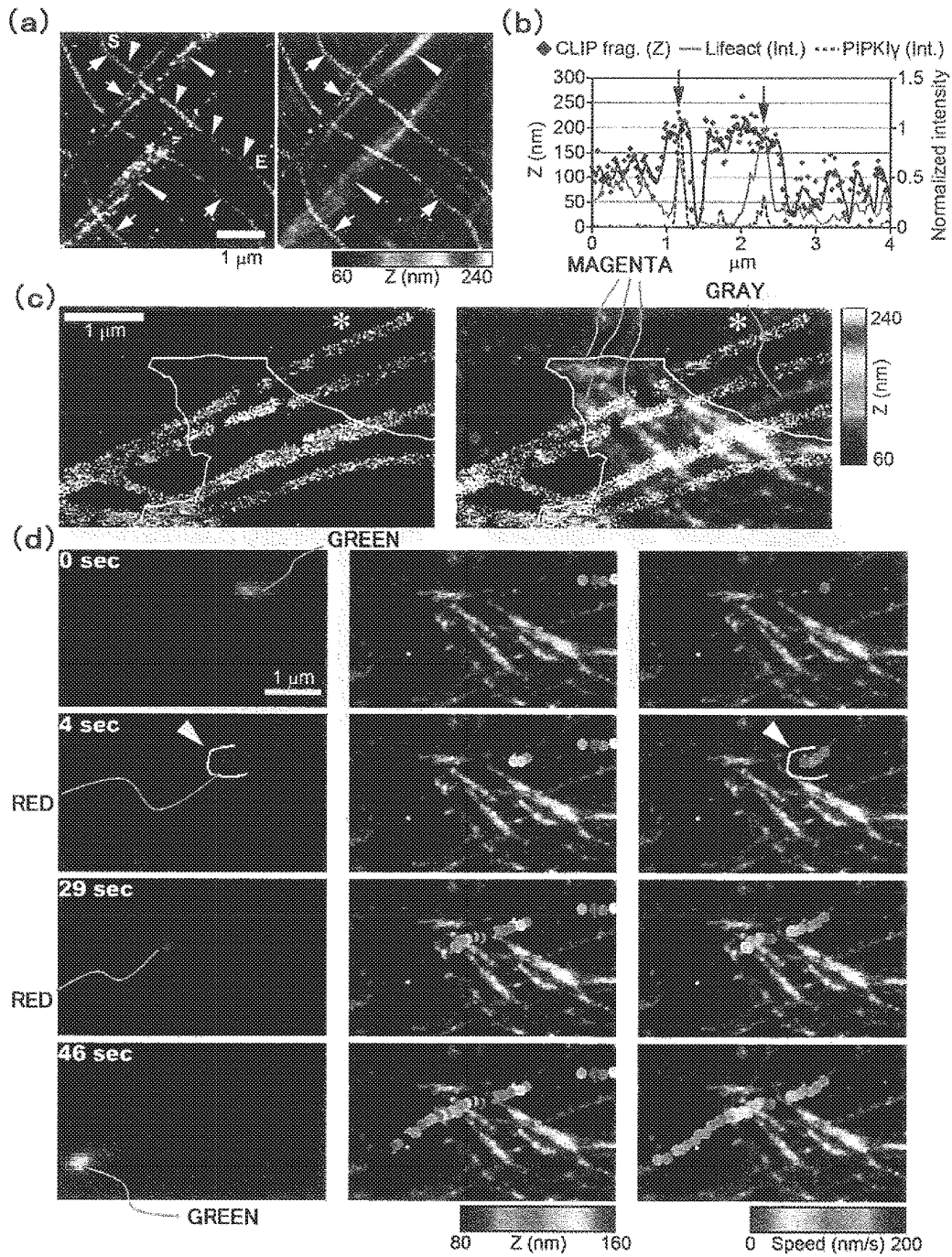


FIG. 8

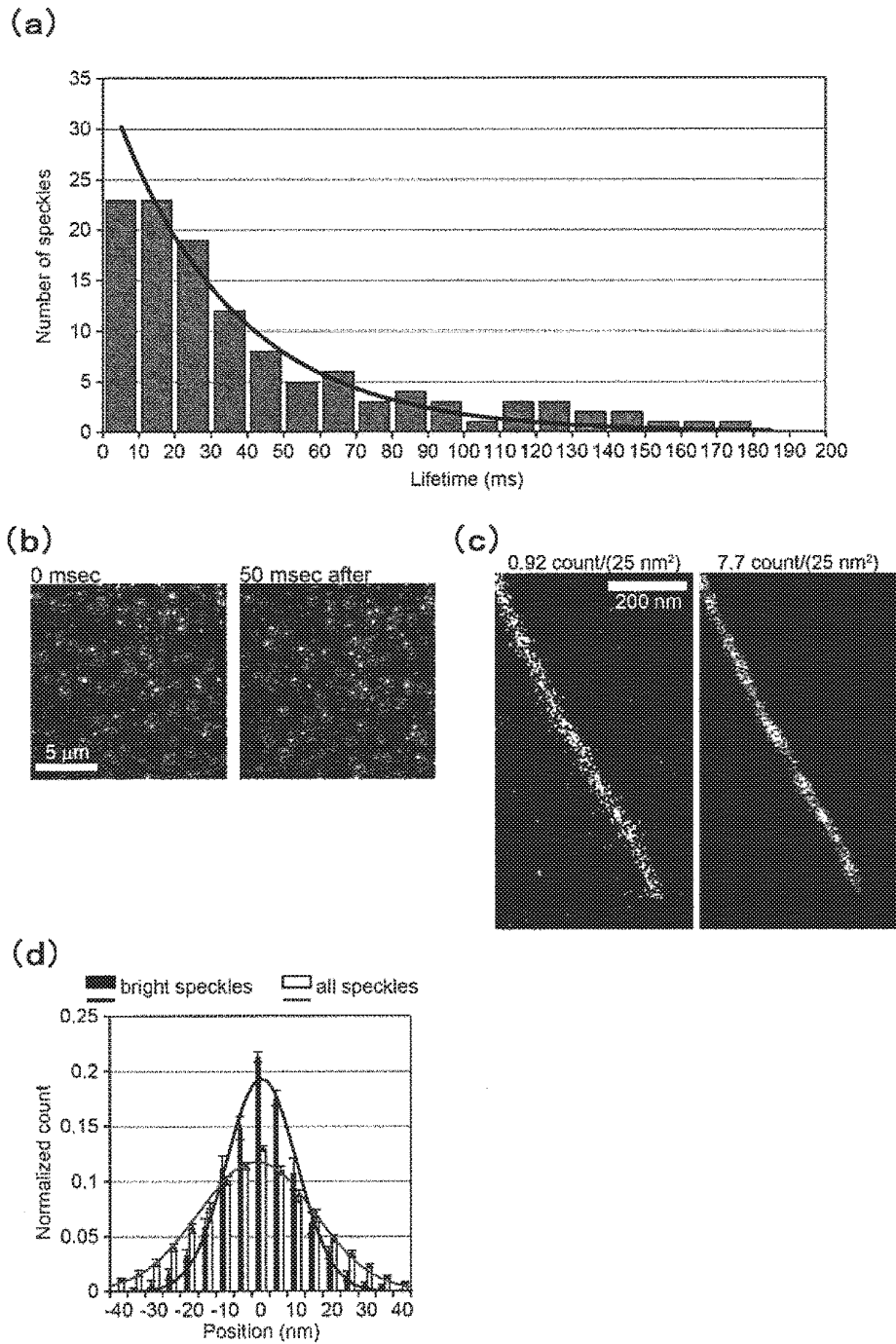


FIG. 9

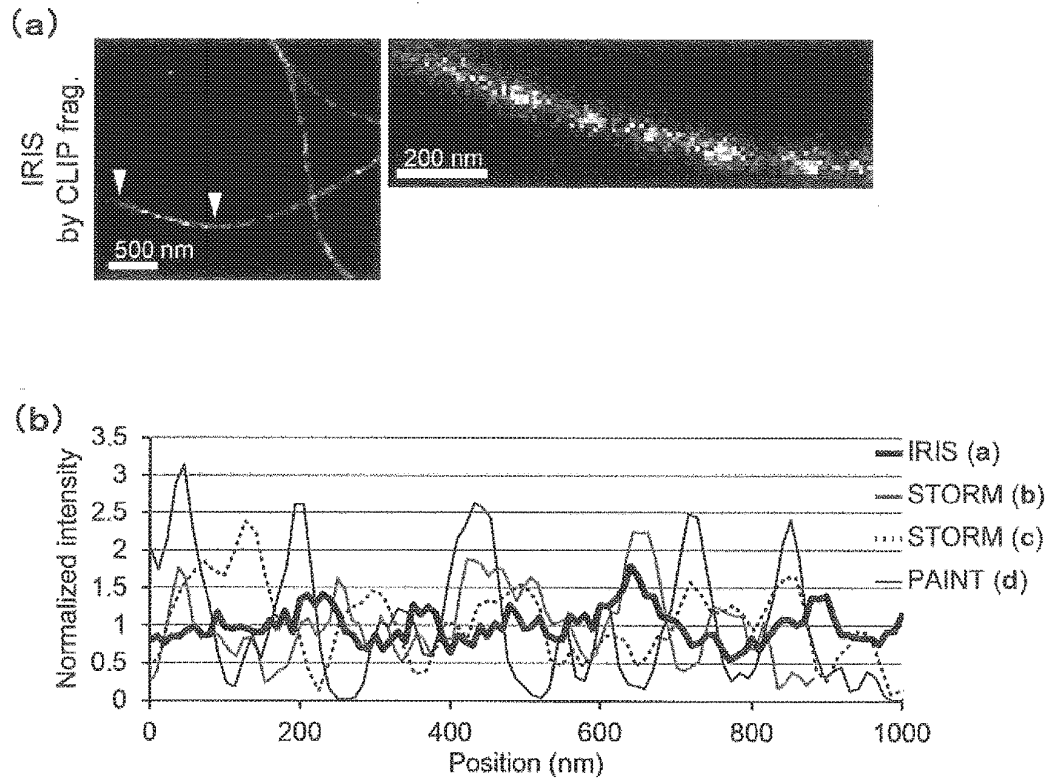
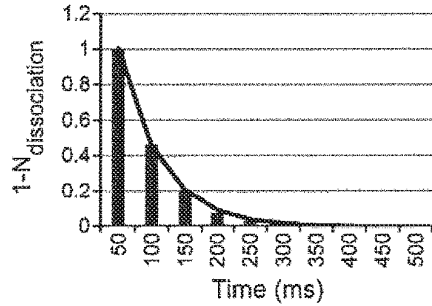
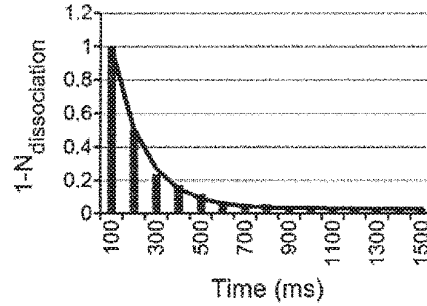


FIG. 10

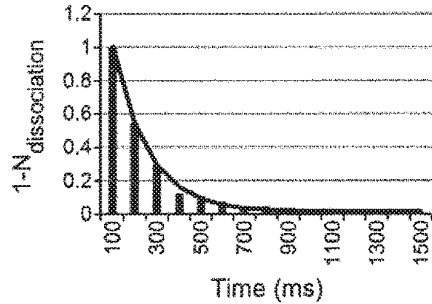
CLIP-170 frag. (3-309), half-life 44 ms



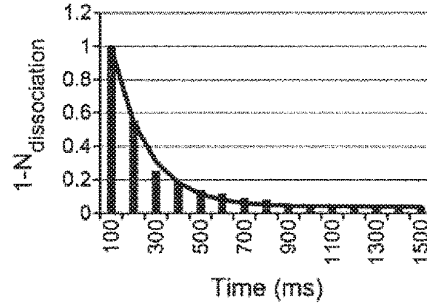
APC frag. (2536-2843), half-life 100 ms



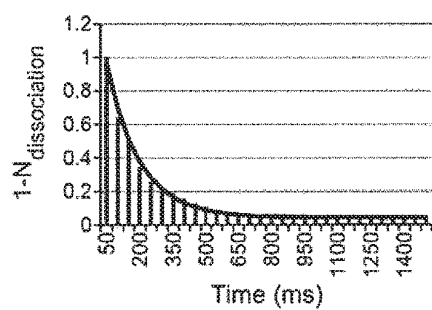
MAP4 frag. (1-908), half-life 109 ms



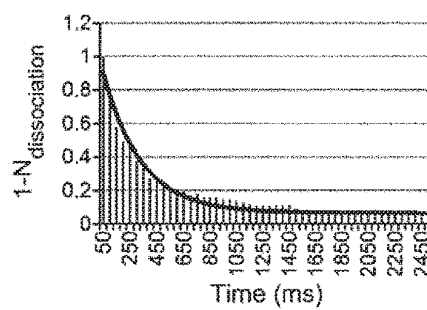
Tau isoform3, half-life 110 ms



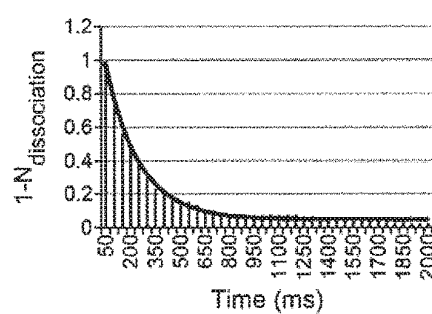
PLEC frag. (4022-4364), half-life 103 ms



Paxillin full length, half-life 196 ms



Src frag. (3-251), half-life 141 ms



PIPKly frag. (641-668), half-life 496 ms

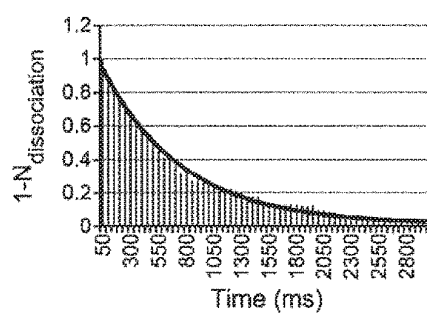


FIG. 11

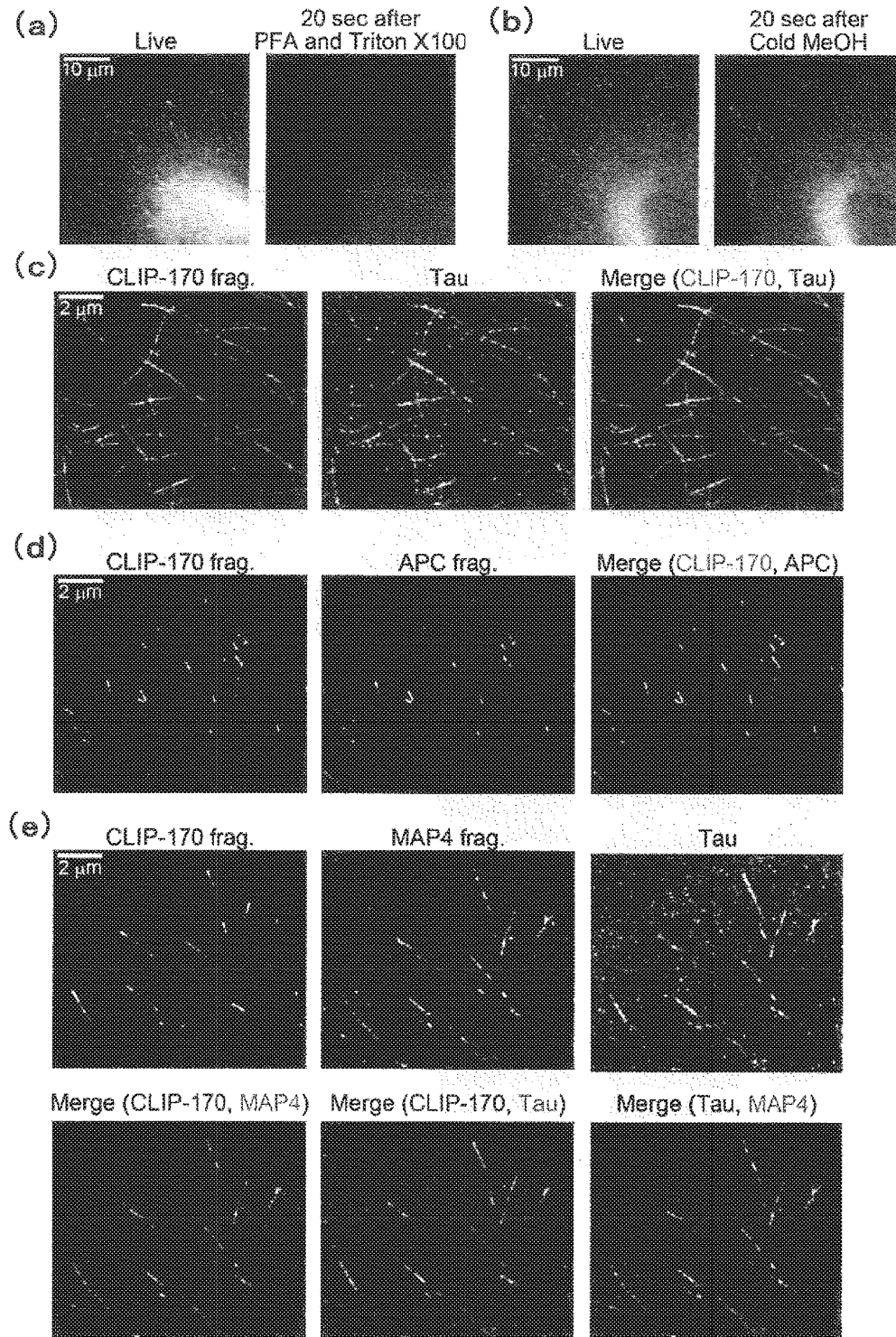


FIG. 12

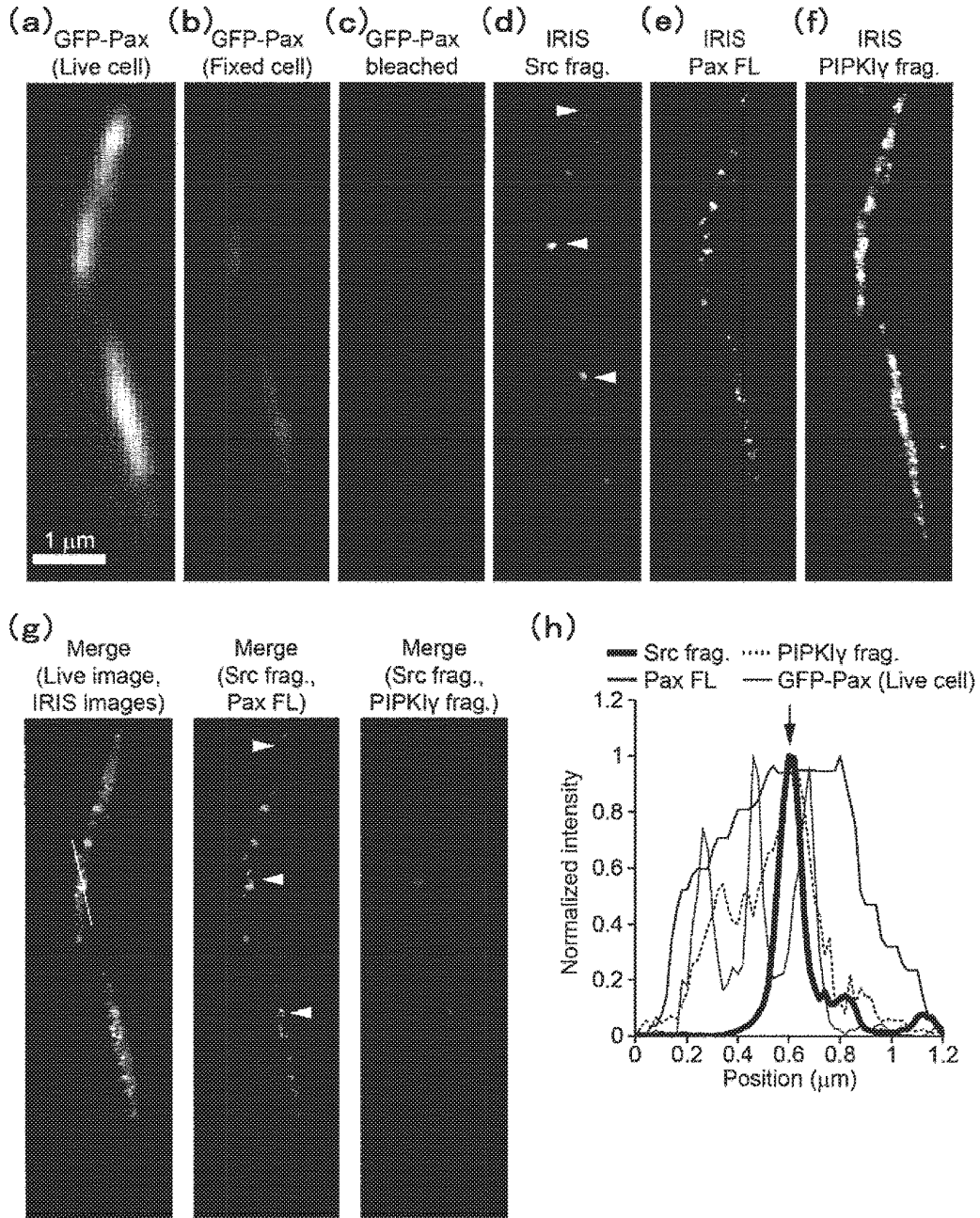


FIG. 13

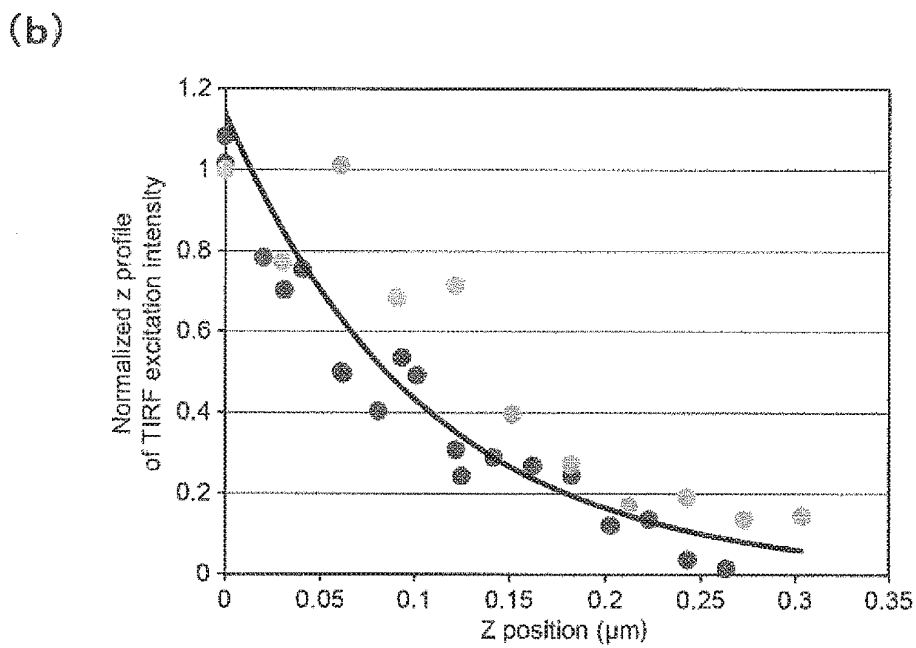
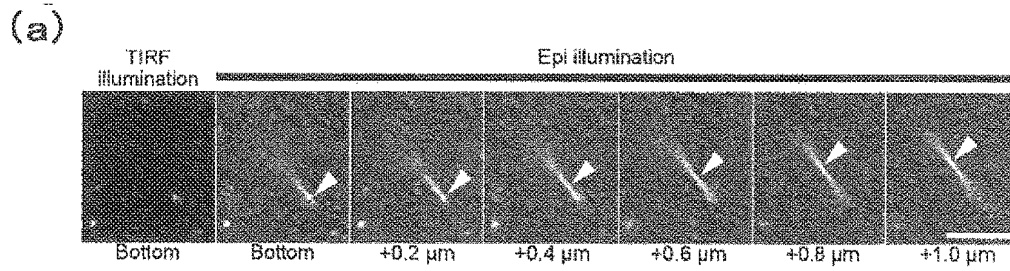


FIG. 14

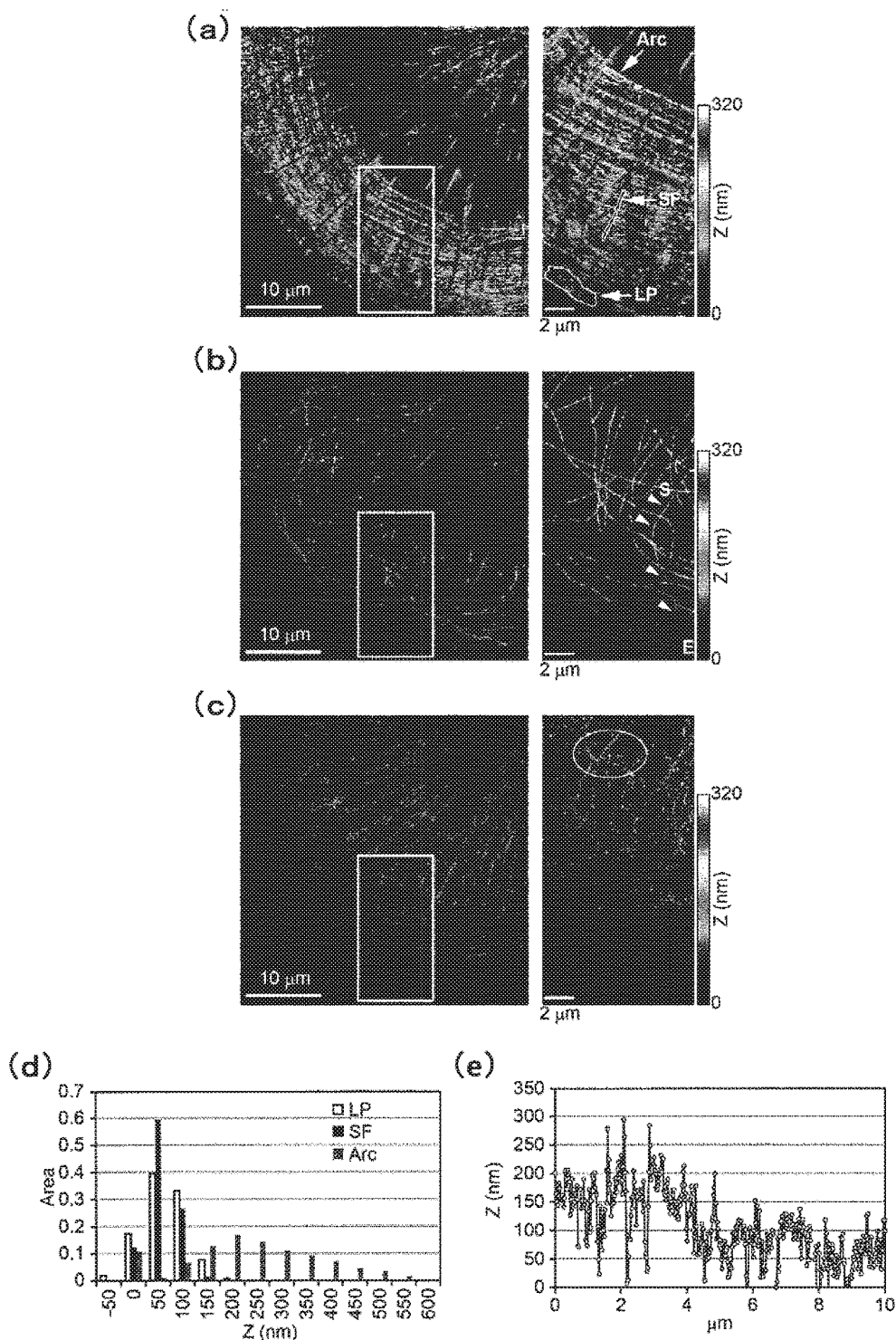


FIG. 15

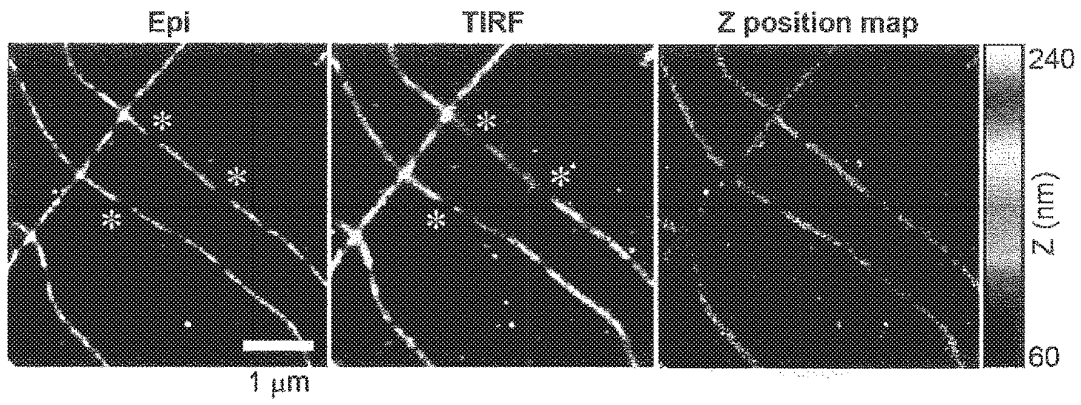


FIG. 16

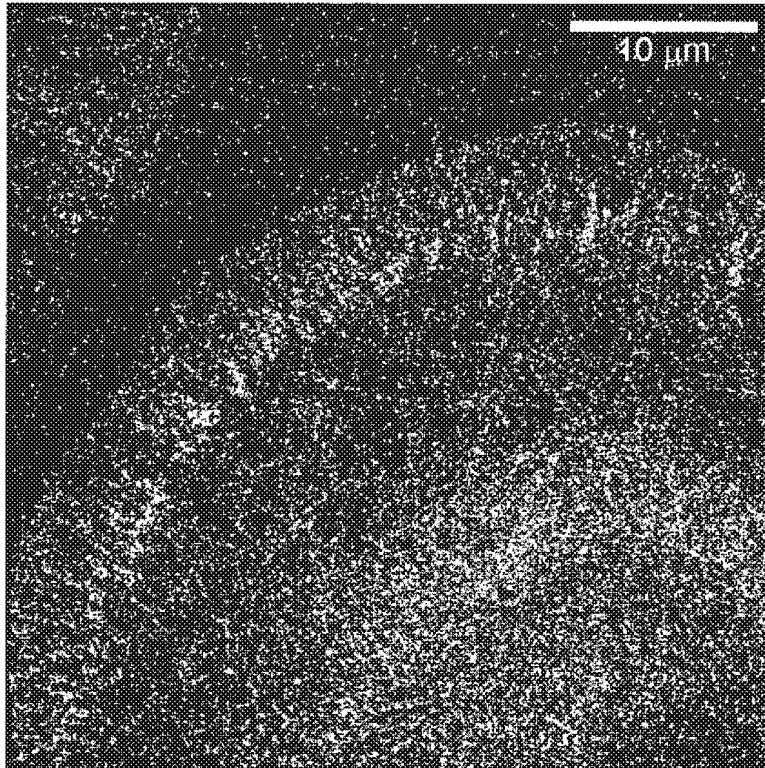


FIG. 17

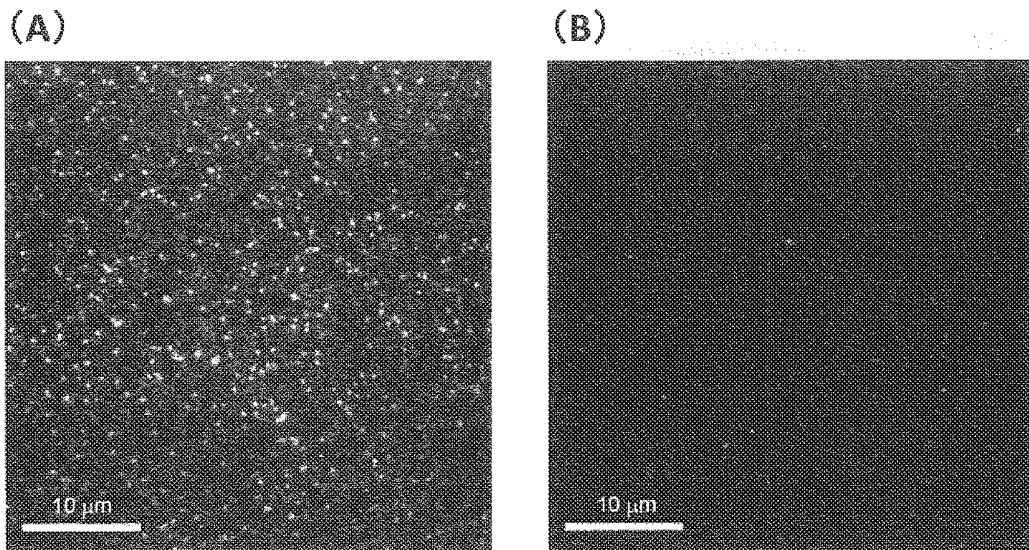


FIG. 18

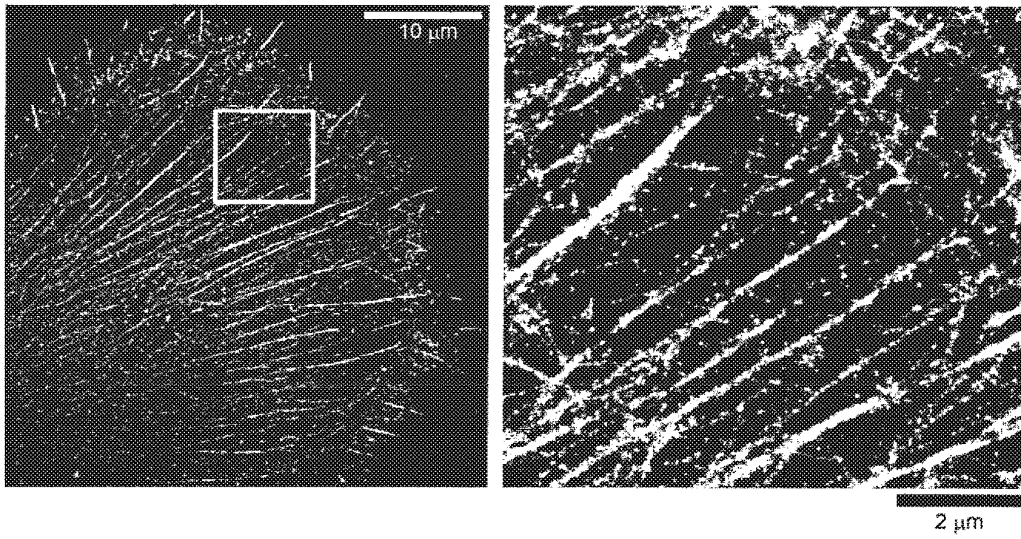


FIG. 19

OBSERVATION METHOD USING BINDING AND DISSOCIATION PROBE

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application is based upon and claims the benefit of priority from prior Japanese Patent Application No. 2015-048692, filed Mar. 11, 2015, the entire contents of which are incorporated herein by this reference. This is a Continuation Application of PCT Application No. PCT/JP2016/057817, filed Mar. 11, 2016, which was not published under PCT Article 21 (2) in English.

BACKGROUND OF THE INVENTION

Field of the Invention

[0002] The present invention is related to an observation method using a binding and dissociation probe by which a super-resolution image of a sample can be obtained.

Description of the Related Art

[0003] In recent years, super-resolution microscopy (STED, SIM, PALM/STORM; resolution capability: 10 nm through 100 nm) with a resolution capability exceeding that of optical microscopes (>200 nm) has been developed (Nat. Rev. Mol. Cell Biol., 9, 929-943, 2008), and major microscope manufacturing companies (Nikon, Carl Zeiss, Leica) are selling their super-resolution microscope apparatuses. In the field of biological science particularly, important discoveries are expected from applications of PALM (photo-activated localization microscopy, Japanese National Publication of International Patent Application No. 2008-542826, Science, 313, 1642-1645, 2006)/STORM (stochastic optical reconstruction microscopy, Nature Methods, 3, 793-795 and United States Patent Application No. US2008/0032414). In each super-resolution microscopy, fluorescent dye is made to bind to a target so as to label the target, and the spatial distribution of the fluorescent dye is observed with a high resolution capability in Stimulated Emission Depletion (STED), structured illumination microscopy (SIM), and localization microscopy (PALM/STORM).

[0004] The outline of localization microscopy is as follows. First, a target substance to be observed is labeled with a luminescent substance such as a fluorescent substance. Then, the luminescent substance with which the target substance is labeled is made to emit light at a low density so that speckle images are obtained in which the speckles of emissions of light are separated individually, i.e., in which luminescent substances of single molecules are separated. The central positions of the individual speckles can be obtained in the respective speckle images that have been obtained. The above step of obtaining speckle images including separated speckles so as to obtain information of the positions of speckles is repeated a plurality of times, e.g., hundreds of times through hundreds of thousands of times, and the central positions of roughly all luminescent substances on the target substance are obtained, and thereby an observation image of the target substance is constructed. Explanations will be schematically given for the localization microscopy by referring to FIG. 1. For example, it is assumed as shown in FIG. 1 (a) that five molecules **100** of target substances to be observed are distributed. The five molecules **100** of the target substance are respectively

labeled with luminescent substances and are made to emit light separately in such a manner that the speckles of light emission do not overlap each other. For example, as shown in FIG. 1 (b), three speckle images **110**, **111** and **112** having separate speckles **101** are obtained at different times **t1**, **t2** and **t3**. A gauss function is fit to the image of each speckle **101** in each of the speckle images **110**, **111** and **112** so as to obtain the central positions of the speckles, and position information **102** which is information of the central positions is recorded (FIG. 1 (c)). Pieces of position information **102** of individual recorded speckles are synthesized so as to draw an observation image **120** as shown in FIG. 1 (d). The above method can be implemented by using for example DAOSTORM, a computer program (Nature methods, 8 279-280, 2011). In localization microscopy, as schematically shown in FIG. (1b), it is necessary that images of the molecules **100**, which are labeled target substances, be picked up over time while the molecules **100** are sequentially made to emit light at a relatively low density so as to obtain a plurality of speckle images so that the speckles of luminescent substances of single molecules can be separated.

[0005] PALM/STORM use a fluorescent substance that can perform activation or switching through irradiation with light for making a fluorescent substance emit light at a low density so as to adjust a light irradiation condition in order to stochastically perform activation or switching on the state of the fluorescent substance, and thereby obtain speckle images in which speckles are separated individually.

[0006] Proceedings of the national Academy of Sciences of the United States of America 103, 18911-18916 (2006) reports a method referred to as PAINT (point accumulation for imaging in nanoscale topography). The document discloses that super-resolution observation of a form of a lipid bilayer was conducted through PAINT by using the fluorescent dye Nile-red that swiftly goes back and forth between an aqueous solution and the lipid bilayer.

[0007] Recently, as a multicolor super-resolution microscopy to which PAINT above is applied, a method has been reported in which a target substance is labeled with an antibody that was made to fuse with a DNA oligomer and a fluorescent DNA oligomer which is a complementary base sequence that temporarily binds to and dissociates from the DNA oligomer is used (Exchange-PAINT, Nature methods, 11, 313-318, 2014).

SUMMARY OF THE INVENTION

[0008] The present inventors have found that a luminescent probe can be used for labeling different positions on a target substance in respective speckle images so that the labeling density with respect to the target substance can be substantially increased by increasing the number of the speckle images that are picked up and a high resolution observation image exceeding a diffraction limit can be generated by the localization microscopy from respective speckle images when a plurality of images of light emission speckles (speckle images) are picked up at different times and while keeping a contact state between the luminescent probe and a sample, by using the luminescent probe, as a binding and dissociation luminescent probe, that is a luminescent probe which repeatedly binds to and dissociates from a target substance directly and specifically and for which the half-life of a probe-target complex formed by binding between the luminescent probe and the target sub-

stance is in a prescribed scope, and the present inventors have completed the present invention. The present invention incorporates the following inventions.

(1) An observation method of a sample containing a target substance, the observation method comprising:

[0009] an imaging step in which a step of obtaining a speckle image including, as a speckle, light emitted from a luminescent substance under a prescribed condition in a state in which a medium is brought into contact with the sample is performed a plurality of times at different times respectively so as to obtain a plurality of speckle images, the medium containing a probe that contains the luminescent substance emitting light under the prescribed condition and that repeatedly binds to and dissociates from the target substance directly and specifically; and

[0010] an observation image generation step of generating an observation image of the target substance in the sample from the plurality of speckle images, wherein

[0011] a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.

(2) The method according to (1), wherein

[0012] the observation image generation step is a step in which information of a position of a speckle included in each of the plurality of speckle images is obtained for each of the plurality of speckle images and the observation image is generated on the basis of the information from the plurality of speckle images.

(3) The method according to (1), wherein

[0013] the sample includes two or more target substances, the imaging step is sequentially performed on the sample by using the probe that is specific to each of the target substances, and

[0014] the observation image generation step is a step in which observation images of the respective target substances in the sample are respectively generated from the plurality of speckle images obtained from the respective imaging steps.

(4) The method according to (3), further comprising

[0015] a multiple-observation image generation step in which observation images of the respective target substances in the sample generated in the observation image generation step are superposed so as to generate a multiple-observation image, which is an observation image of the two or more target substances in the sample.

(5) The method according to (1), wherein

[0016] the luminescent substance is a fluorescent substance, and the prescribed condition is irradiation with excitation light.

(6) The method according to (1), wherein

[0017] a combination between the probe and the target substance is selected from a group of:

[0018] a combination wherein the probe is (a1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 19, (a2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (a1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of

a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (a3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (a1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is an actin polymer;

[0019] a combination wherein the probe is (b1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 12, that at least partially contains an amino acid sequence of 3-309 and that has 407 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2536-2843 and that has 408 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2781-2819 and that has 138 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 1-908 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 659-908 and that has 394 or fewer amino acids, an amino acid sequence of sequence number 5, or an amino acid sequence of sequence number 6, (b2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (b1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (b3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (b1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is a microtubule;

[0020] a combination wherein the probe is (c1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4684 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of

3777-4364 and that has 688 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4313 and that has 637 or fewer amino acids, or an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 4022-4364 and that has 443 or fewer amino acids, (c2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (c1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (c3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (c1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is an intermediate filament; and

[0021] a combination wherein the probe is (d1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 15, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-557 and that has 556 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-498 and that has 545 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 167-557 and that has 491 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino acid sequence of 1-251 and that has 351 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino acid sequence of 3-251 and that has 349 or fewer amino acids or an amino acid sequence of sequence number 18, (d2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (d1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (d3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (d1) and for which a half-life of a probe-target complex

formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is a focal adhesion.

(7) The method according to (1), wherein

[0022] the probe contains an antibody or a fragment of an antibody, the antibody or the fragment being to the target substance and the antibody or the fragment being linked to the luminescent substance.

(8) The method according to (7), wherein the fragment of the antibody is a Fab fragment.

(9) A probe used for labeling a target substance, wherein

[0023] the probe contains a luminescent substance that emits light under a prescribed condition,

[0024] the probe can repeatedly bind to and dissociate from the target substance directly and specifically, and

[0025] a half-life of a probe-target complex formed by binding to the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.

(10) The probe according to (9), wherein

[0026] the target substance is an actin polymer and the probe is (a1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 19, (a2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (a1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (a3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (a1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds,

[0027] the target substance is a microtubule and the probe is (b1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 12, that at least partially contains an amino acid sequence of 3-309 and that has 407 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2536-2843 and that has 408 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2781-2819 and that has 138 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 1-908 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 659-908 and that has 394 or fewer amino acids, an amino acid sequence of sequence number 5 or an amino acid sequence of sequence number 6, (b2) a polypeptide, linked to the luminescent substance, which consists of the amino

acid sequence described in (b1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (b3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (b1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds,

[0028] the target substance is an intermediate filament and the probe is (c1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4684 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4364 and that has 688 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4313 and that has 637 or fewer amino acids or an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 4022-4364 and that has 443 or fewer amino acids, (c2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (c1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (c3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (c1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or

[0029] the target substance is a focal adhesion and the probe is (d1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 15, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-557 and that has 556 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-498 and that has 545 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 167-557 and that has 491 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence

of sequence number 16, that at least partially contains an amino acid sequence of 1-251 and that has 351 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino acid sequence of 3-251 and that has 349 or fewer amino acids, or an amino acid sequence of sequence number 18, (d2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (d1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds or (d3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (d1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.

(11) The probe according to (9), wherein

[0030] the probe contains an antibody or a fragment of an antibody, the antibody or the fragment being to the target substance and the antibody or the fragment being linked to the luminescent substance.

(12) The probe according to (11), wherein

[0031] the fragment of the antibody is a Fab fragment.

(13) A reagent kit for labeling a target substance, wherein

[0032] the reagent kit at least includes the probe according to (9).

(14) A screening method of a site in which identifies a target substance in the probe according to (9), the screening method comprising:

[0033] an immobilization step in which a candidate substance of the site or a substance partially containing the candidate substance is fixed to a solid support;

[0034] an observation step in which a target substance linked to a luminescent substance and a solid support obtained in the immobilization step are observed in a medium while the target substance linked to a luminescent substance and the solid support obtained in the immobilization step are kept in contact, in a condition that allows observation, in units of 1 molecule, of light emission from the luminescent substance in a probe-target complex formed by binding between the target substance and the candidate substance, and

[0035] a screening step in which the candidate substance resulting in a half-life of the probe-target complex that is equal to or more than 10 milliseconds and equal to or less than 3 seconds is selected as the site on the basis of observation in the observation step.

(15) The method according to (14) wherein

[0036] the candidate substance is an antibody or a fragment of an antibody from a library of hybridoma that produces an antibody to the target substance, and

[0037] the antibody is fixed to a solid support in the immobilization step.

[0038] The present document incorporates the contents of the disclosure of Japanese patent application No. 2015-048692, on the basis of which the priority is claimed for the present application.

[0039] According to the present invention, operations and effects that are remarkably more advantageous than those of existing super-resolution microscopy can be provided because (i) the problem of a labeling density, which has been the cause of reduced reliability of existing super-resolution microscopy, can be resolved and (ii) it is easy to visualize a plurality of target substances by protein-based exchangeable probes and there are no limitations on the number of target substances.

BRIEF DESCRIPTION OF THE DRAWINGS

[0040] The present invention will be more apparent from the following detailed description when the accompanying drawings are referenced.

[0041] FIG. 1 explains the outline of the localization microscopy. FIG. 1 (a) shows the distribution of the actual probe molecules. FIG. 1 (b) shows speckle images containing fluorescent speckles picked up at different times. FIG. 1 (c) shows position information of speckles in the respective speckle images. FIG. 1 (d) shows an observation image reconstructed on the basis of the position information.

[0042] FIG. 2 (a) is a schematic view for explaining the principle by which the labeling density can be increased by using the method of the present invention. FIG. 2 (b) is a schematic view for explaining the principle of STORM.

[0043] FIG. 3 is a schematic view for explaining procedures for observing a plurality of target substances in one sample by using the method of the present invention.

[0044] FIG. 4 is a functional block diagram of a microscope apparatus used for implementing the method of the present invention.

[0045] FIG. 5 are related to super-resolution microscopy according to the present invention. Super-resolution microscopy according to the present invention will be referred to as IRIS (image reconstruction by integrating exchangeable single-molecule localization). FIG. 5 (a) shows the outline of IRIS. Transient associations of single-molecule fluorescent probes with their targets are visualized, and central positions of the probes are identified with nanometer accuracy. Integrating the position information from many frames produces a super-resolution image of the target substance. FIG. 5 (b) shows a super-resolution image of a single actin filament *in vitro* by TIRF (total internal reflection fluorescence) microscopy using Atto 488-Lifeact. An observation image was reconstructed by using high-brightness speckles in order to improve the localization accuracy (see FIG. 9c). FIG. 5 (c) shows the cross-sectional profile of single actin filaments ($n=10$ filaments, gray bars). The black curve shows the Gaussian fit to the mean profile, with a FWHM of 23 nm. Error bars show S.E.M. FIG. 5 (d) shows dependency of image quality on the labeling density *in vitro*. The labeling density of Atto 488-Lifeact per unit length is indicated at the top of each view. FIG. 5 (e) shows line profiles of the labeling density of Atto 488-Lifeact along the frame in FIG. 5 (d). The labeling density is shown after being normalized by the mean labeling density of an entire actin filament. FIG. 5 (f) shows an IRIS image of actin filaments in a cell using Atto 488-Lifeact and total internal reflection illumination. The image was reconstructed from 5×10^5 frames. FIG. 5 (g) shows a comparison between the IRIS super-resolution image (upper left) and the added SIMS image (lower left) of the region in the frame in FIG. 5 (f). The right graph shows cross-sectional profiles of two adjacent filaments between the two arrowheads in the upper

left image. Atto 488-Lifeact was excited by the simultaneous irradiation with 473-nm and 488-nm laser beams in the TIRF mode to obtain a strong signal from each speckle. The stage drift of the microscope was corrected with a bright-field image of nonfluorescent beads in observation of an actin filament *in vitro* and was corrected with a bright-field image of the cell itself in observation of an actin filament in the cell (see "Methods" in the examples).

[0046] FIG. 6 (a) shows super-resolution images of actin filaments using Lifeact. FIG. 6 (b) shows super-resolution images of microtubules using a CLIP-170 fragment (amino acid residues 3-309 of sequence number 12). FIG. 6 (c) shows super-resolution images of intermediate filaments using a Plecin-1 (PLEC) fragment (amino acid residue 4022-4364 of sequence number 8). FIG. 6 (d) shows a super-resolution image of a focal adhesion using a phosphatidylinositol-(4)-phosphate 5-kinase type 1 γ -90 (PIP1 γ) fragment (641-668) (sequence number 18). Except for a case when a PIP1 γ fragment was used as a probe, SIMS images (speckle images) of individual probes were obtained by alternately conducting total internal reflection illumination and epi-illumination. Specifically, imaging (exposure time was 50 ms/frame, 20 Hz) for 250 consecutive frames of speckle images based on the total internal reflection fluorescence in accordance with the condition explained in "procedures for imaging of multicolor super-resolution by IRIS" in "methods" of examples and imaging (exposure time was 50 ms/frame, 20 Hz) for 250 consecutive frames of speckle images based on the epi-fluorescence were repeated for observation of actin filament, microtubule and intermediate filament. For imaging of focal adhesions, speckle images based on epi-illumination were not obtained and imaging (exposure time was 50 ms/frame, 20 Hz) of 500 consecutive frames based on total internal reflection illumination was repeated. Then, the super-resolution images in the cell bottom (TIRF=total internal reflection fluorescence) and the entire cell peripheral regions (Epi=epi-fluorescence) were merged so as to reconstruct a super-resolution image. In FIG. 6 (a) through FIG. 6 (c), images obtained by merging TIRF images and epi-fluorescence images are shown, and the enlarged images of the regions enclosed by the frames are shown as the rightmost images. FIG. 6 (e) shows an image resulting from merging seven super-resolution images. The images were respectively reconstructed from 2×10^7 frames (Lifeact (TIRF), Lifeact (Epi)), 4×10^4 frames (CLIP frag. (TIRF), CLIP frag. (Epi)), 1.2×10^5 frames (PLEC frag. (TIRF), PLEC frag. (Epi)) and 4×10^4 frames (PIP1 γ frag. (TIRF)). The number of frames of speckle images used for constructing the TIRF image of FIG. 6 (a) is 2×10^5 , the number of frames of speckle images used for constructing the Epi image of FIG. 6 (a) is 2×10^5 , the number of frames of speckle images used for constructing the TIRF image of FIG. 6 (b) is 4×10^4 , the number of frames of speckle images used for constructing the Epi of FIG. 6 (b) is 4×10^4 , the number of frames of speckle images used for constructing the TIRF image of FIG. 6 (c) is 1.2×10^5 , the number of frames of speckle images used for constructing the Epi of FIG. 6 (c) is 1.2×10^5 , and the number of frames of speckle images used for constructing the TIRF image of FIG. 6 (d) is 4×10^4 . Also, the probe concentration in the imaging solution for speckle imaging was 2.4 nM for the imaging in FIG. 6 (a), 8 nM for the imaging in FIG. 6 (b), 9.2 nM for the imaging in FIG. 6 (c) and 84 nM for the imaging in FIG. 6 (d). Note that the above probe concen-

trations are the actual probe concentration in the case when Atto488-Lifeact was used as the probe and are conversion values based on the fluorescence intensity of labeling fluorescent proteins in the case when other probes were used.

[0047] FIG. 7 show region-specific proximity between cytoskeletons and focal adhesions. FIG. 7(a) shows an epi-fluorescence super-resolution image of intermediate filaments in the lamella region in ROI1 in FIG. 6 (e). FIG. 7 (b) shows a merged super-resolution image of the intermediate filaments (IF) and actin filaments (Act). FIG. 7 (c) shows a merged super-resolution image of intermediate filaments (IF) and microtubules (MT). FIG. 7 (d) shows a merged super-resolution of intermediate filaments (IF), actin filaments (Act) and microtubules (MT) in super-resolution image (a). FIG. 7 (e) shows cross-sectional profiles of three types of cytoskeletons between the arrowheads in FIG. 7 (d). Intermediate filaments are tangled with actin stress fibers (at the positions of arrows) in the lamellar region but are not tangled with microtubules. FIG. 7 (f) shows a TIRF super-resolution image of intermediate filaments in the peripheral region of ROI2 in FIG. 6 (e). FIG. 7 (g) shows a merged TIRF super-resolution image of intermediate filaments (IF) and actin filaments (Act). FIG. 7 (h) shows a merged TIRF super-resolution image of intermediate filaments (IF) and microtubules (MT). FIG. 7 (i) shows a merged TIRF super-resolution image of intermediate filaments (IF), actin filaments (Act), microtubules (MT) and focal adhesions (FA) in the region in the frame in super-resolution image (f). In FIG. 7 (g), which shows a TIRF super-resolution image, the intermediate filaments (IF) are denoted by narrow arrowheads and the actin filaments (Act) are denoted by thick arrowheads. In FIG. 7 (h), which shows a TIRF super-resolution image, the intermediate filaments (IF) are denoted by narrow arrowheads and the microtubules (MT) are denoted by thick arrowheads. FIG. 7 (j) shows cross-sectional profiles of three types of cytoskeletons between the arrowheads in FIG. 7 (i). In this peripheral region, the intermediate filaments overlap a microtubule at the position of the arrow but do not overlap actin filaments.

[0048] FIG. 8 show movements of microtubule tips in the vicinity of actin stress fibers and focal adhesions. The left image of FIG. 8 (a) is an image resulting from merging a z position map (FIG. 16) of microtubules (MT) and a TIRF super-resolution image of a focal adhesion (FA) in the central region in FIG. 7 (i). The right image of FIG. 8 (a) is an image resulting from merging a z position map (FIG. 16) of microtubules (MT) and a TIRF super-resolution image of actin filaments (Act) in the central region in FIG. 7 (i). The z positions of microtubules were calculated from the signal intensity ratio between an epi-fluorescence super-resolution image and a TIRF super-resolution image (see "Methods" in the examples). In the left image of FIG. 8 (a), the microtubules (MT) are denoted by arrows and the focal adhesions (FA) are denoted by narrow arrowheads. In the right image of FIG. 8 (a), the microtubules (MT) are denoted by arrows and the actin filaments (Act) are denoted by narrow arrowheads. FIG. 8 (b) shows the line profiles (CLIP frag) of the z position of a microtubule, the intensity (Lifeact) of an actin filament and the intensity (PIP2K1 γ) of a focal adhesion along the interval between S and E of the microtubules denoted by the thick arrowheads in FIG. 8 (a). The curve of the thick solid line shows a movement average of four data points for the z position of the microtubule. The left image of FIG. 8 (c) is a map of the z positions of microtubules, and the right

image of FIG. 8 (c) is an image resulting from merging the TIRF super-resolution images of actin filaments (grey) and focal adhesions (magenta). The focal adhesion was visualized by combining a super-resolution image obtained by using the Src fragment (residues 3-251 of sequence number 15) as a probe and a super-resolution image obtained by using the Paxillin full length as a probe. In FIG. 8 (d), the left panels are live-cell epi-fluorescence observation images (red) and TIRF observation images (green) of EB1-EGFP obtained before IRIS imaging. The middle panels are results of causing overlap between the z position of the tip of the EB1-labeled microtubule, the actin filament and focal adhesion shown in the right image of FIG. 8 (c), and the right panels are results of causing overlap between the speed of the portion of the tip of the EB1-labeled microtubule, the actin filament and focal adhesion shown in the right image of FIG. 8 (c). The microtubule analyzed with the trajectory of EB1 is indicated by an asterisk in FIG. 8 (c).

[0049] FIG. 9 show characteristic evaluation of Atto 488-Lifeact. FIG. 9 (a) shows speckle lifetime distribution of Atto488-Lifeact in fixed XTC cells. The SiMS images (speckle images) were picked up consecutively with an exposure time of 10 ms/frame and at a frame rate of 100 Hz by using a 488 nm laser (with the main body output of 50 mW but reaching the sample after being attenuated by AOTF etc.). The measurement of the speckle lifetime (having obtained 100 pixels \times 100 pixels) of a narrow scope of the sample was performed using the Speckle TrackerJ plug-in so that 100 Hz is achieved, i.e. so that 1 frame was able to be obtained per 10 ms. The black line shows a half-life of 23 ms fit to the single exponential curve of the lifetime distribution of speckles between 20 ms and 110 ms. The photobleaching rate of Atto488-Lifeact was negligible in the term of 200 ms. FIG. 9b shows analysis of Atto488-Lifeact speckles using DAOSTORM (circles in the image). The SiMS images of Atto488-Lifeact in a cell were obtained with an exposure time of 50 ms. The central position of each speckle was determined with nanometer accuracy. The distribution of speckles changed greatly in the frame next to a frame in which an image was picked up 50 milliseconds later. FIG. 9 (c) shows super-resolved images of a single actin filament in vitro by TIRF microscopy using Atto488-Lifeact. The left image is an image obtained by the reconstruction from only high-brightness speckles (top 12% of all measured speckles) and is the same as that shown in FIG. 5 (b). The right image was reconstructed from all speckles. The labeling density on the actin filament is indicated at the top of each image. FIG. 9 (d) shows the cross-sectional profiles of single actin filaments in vitro (n=10 filaments) in the image reconstructed from high-brightness speckles (black) or in the image reconstructed from all speckles (white). Normalization was conducted so that all counts became 1. The black curve shows a Gaussian fit curve with a FWHM of 23 nm (high-brightness speckles) and the white curve shows a Gaussian fit curve with a FWHM of 38 nm (all speckles). The error bar represents \pm S.E.M.

[0050] FIG. 10 show a comparison of labeling patterns along the longitudinal direction of microtubules between IRIS and other super-resolution techniques. FIG. 10 (a) shows super-resolution images of microtubules by IRIS using CLIP-170 as probes. These images are part of the epi-fluorescence IRIS observation image of FIG. 6 (b). FIG. 10 (b) shows line profiles of labeling intensity along microtubules in the right image of FIG. 10 (a) (assumed to be IRIS

(a)), the super-resolution of a microtubule by STORM using the anti- β tubulin described in document 21 (assumed to be STORM (b)), the super-resolution image of a microtubule by STORM using the anti- β tubulin described in document 22 (assumed to be STORM (c)), and the super-resolution image of a microtubule by Exchange-PAINT using the anti- β tubulin described in document 23 (assumed to be PAINT (d)). The labeling density is shown after being normalized by the mean labeling density of an entire actin filament.

[0051] FIG. 11 shows the lifetime distribution data of the selected IRIS probes on the targets. 100 to 112 lifetimes were plotted in accordance with a complementary cumulative distribution function (1-Ndissociation) for each probe. Ndissociation is the cumulative relative frequency of dissociated probes. The half-life of probes on the targets was determined by fitting the life time distribution data by a single exponential function. Part of the measurement results shown on table 1 is shown here.

[0052] FIG. 12 show labeling patterns of microtubules by a CLIP-170 fragment (amino acid residues 3-309 of sequence number 12), an APC fragment (amino acid residues 2536-2843 of sequence number 14), a MAP4 fragment (amino acid residues 1-908 of sequence number 4) and Tau isoform 3 (full length of sequence number 5). The patterns are different depending upon procedures for fixing cells. FIG. 12 (a) and FIG. 12 (b) show live-cell imaging during the fixation operation. In FIG. 12 (a), the accumulation of EB1-EGFP at the tips of microtubules disappeared in a process by 3.7% PFA and 0.5% Triton-X 100. In FIG. 12 (b), EB1-EGFP was fixed in a state in which it was accumulated at the tips of the microtubules in a process by cold methanol. FIG. 12 (c) shows TIRF IRIS images (observation images) of microtubules using CLIP-170 fragment and Tau isoform 3 in cells processed by PFA and Triton-X 100. With these probes, IRIS images of the entire microtubules were obtained. The image resulting from merging these IRIS images indicates that the entire microtubules visualized by these probes correspond to each other. FIG. 12 (d) shows the TIRF IRIS images of microtubules using a CLIP-170 fragment and an APC fragment in cells preprocessed with cold methanol for 5 seconds and next with PFA and Triton X-100 as a preprocess. These probes resulted in IRIS images of microtubules where the microtubule tips were visualized with much stronger signals than were the entire microtubules. The image resulting from merging these images shows a coincidence of the microtubule tips visualized by these probes. FIG. 12 (e) shows TIRF IRIS images of microtubules using a CLIP-170 fragment, a MAP4 fragment and Tau isoform 3 in cold methanol-preprocessed cells. The microtubule tips were labeled strongly with a CLIP-170 fragment while they were weakly labeled with a MAP4 fragment and Tau isoform 3 (see merged images in the lower tier).

[0053] FIG. 13 show labeling patterns of focal adhesions by Paxillin full length (FL) (full length of sequence number 15), the Src fragment (amino acid residues 3-251 of sequence number 16) and a PIPKI γ fragment (residues 641-668) (sequence number 18). FIG. 13 (a) shows live-cell images in which GFP-Paxillin was used for visualizing focal adhesions for comparison. FIG. 13 (b) shows images of cells obtained by fixing and providing a permeabilization process to the living cells after the imaging of FIG. 13 (a). FIG. 13 (c) shows a state in which a remaining fluorescence of GFP-Paxillin was completely bleached by irradiation with a

strong excitation laser. FIG. 13 (d) through FIG. 13 (f) are IRIS images in the order of the Src fragment, Paxillin FL, and a PIPKI γ fragment after the photobleaching of FIG. 13 (c). In an IRIS image view of FIG. 13 (d), generated by using the Src fragment, some dot structures appeared (arrowheads in FIG. 13 (d)) in the focal adhesion. These dot structures were not reconstructed by probes that did not dissolve, but were reconstructed by probes that repeatedly bound. In Paxillin FL, a focal adhesion was labeled partially. The left image of FIG. 13 (g) is a result of merging FIGS. 13 (a) (d) (e) and (f), the center image is a result of merging FIGS. 13 (d) and (e), and the right image is a result of merging FIGS. 13 (d) and (f). In the center image, the dot structures visualized by Src fragments (portions denoted by the arrowheads) have many portions that do not correspond to the focal adhesions visualized by Paxillin FL. FIG. 13 (h) shows the cross-sectional profiles of the focal adhesions in three IRIS images and the living-cell image of GFP-Paxillin. The measured scope is denoted by the line in the left image of FIG. 13 (g). The dot structures visualized by Src fragments are not completely visualized by Paxillin FL (arrow in FIG. 13 (h)). These proteins have different binding partners in focal adhesions (documents 17, 38 and 39). These differences between IRIS images may indicate a distribution of partners that respective proteins can associate with in focal adhesions.

[0054] FIG. 14 show the z profile of the TIRF excitation intensity. FIG. 14 (a) shows fluorescence images of HyLight488-labeled microtubules obtained by total internal reflection fluorescence (TIRF) and epi fluorescence. The epi fluorescence images were obtained as z-stack images (0.2 μ m step size). The z-stack epi-fluorescence images were used to determine the z-directional distance of each point along the tilted microtubule (see "Methods"). The bar represents 5 μ m. FIG. 14 (b) shows the Z profile of the TIRF excitation intensity.

[0055] FIG. 15 (a) shows z-position mapping of actin bundles, FIG. 15 (b) shows z-position mapping of microtubules, and FIG. 15 (c) shows z-position mapping of intermediate filaments. The respective images of FIGS. 15 (a), (b) and (c) were calculated and generated from the total internal reflection fluorescence (TIRF) IRIS image and the epi-fluorescence IRIS image shown in FIGS. 6 (a), (b) and (c) (see "Methods"). The right image is an enlarged image of the region in the frame shown in the left image. FIG. 15 (d) shows a distribution of z positions of actin bundles in lamellipodia (LP), stress fibers (SF) and actin arcs (Arc). In these layered actin structures, a calculated z position represents the height position of the center of gravity in the z axial direction of an actin filament. FIG. 15 (e) shows the z-position profile along the longitudinal direction of a microtubule. The microtubule submerges toward the periphery of a cell (arrowheads in FIG. 15 (b)). S and E in FIG. 15 (b) represent the starting and ending points of the line profile.

[0056] FIG. 16 shows an epi-fluorescence super-resolution image of microtubules (left), a TIRF image (center) and the z-position mapping shown in FIG. 8 (a) (right). In CLIP-170 fragment-visualized microtubules, signals were partially lost (asterisks in the left and center images).

[0057] FIG. 17 shows a result of IRIS super-resolution imaging using a probe containing a Fab fragment (Fab probe). It is a result of imaging a distribution of p40, which is a subunit of an Arp2/3 complex, by using a Fab probe that is produced from anti-p40 polyclonal antibody.

[0058] FIG. 18 show binding of FLAG-EGFP to a solid-phased antibody. An antibody contained in hybridoma culture supernatant was fixed to a glass surface and a FLAG-EGFP solution was added so as to observe it with a TIRF microscope. Each speckle is FLAG-EGFP that has bound to a solid-phased antibody. FIG. 18 (A) shows a reaction positive example. Speckles representing binding between FLAG-EGFP and antibodies are observed at a high density. An observation result of a great number of speckles indicates that solid-phased antibodies are anti-FLAG antibodies having a binding capacity to FLAG-EGFP. FIG. 18 (B) shows a reaction negative example. Few speckles are observed. This is a similar level to a case where FLAG-EGFP was added to a glass surface that was not solid phased.

[0059] FIG. 19 shows super-resolution images of FLAG fused actins obtained by using a Fab probe derived from an anti-FLAG monoclonal antibody selected from a hybridoma library (203 milliseconds as the half-life of a probe-target complex). This cell has forcibly expressed a FLAG tag fused actin. The right image is an enlarged image of the region in the white frame in the left image.

DESCRIPTION OF THE EMBODIMENTS

[0060] In super-resolution microscopy, an increased resolution capability has brought the new problems of a low labeling density and unevenness thereof. According to a sampling theorem for example, it has been shown that one labeling substance has to be included at 10 nm in a target in order to obtain a spatial resolution capability of 20 nm (J. Cell Sci., 126, 3505-3513, 2013). In conventional super-resolution imaging of proteins in cells, target proteins are labeled by using the expression of target proteins with which fluorescent proteins have been made to fuse or by using a fluorescent antibody. This imposes limitations on a labeling density, depending upon the expression amount ratio to endogenous target proteins and upon the size (about 10 nm) of an antibody itself. Also, because fluorescent dyes that have bound to a target substance are used, the maximum number of types of target substances that can be visualized in one sample is two or three. The limitations on the number of proteins that can be stained have been a long-standing issue in the study of a cell consisting of various types of proteins.

[0061] In Exchange-PAINT above as well, target substances have to be labeled with antibodies that were made to fuse with DNA oligomers, and thus it is expected that a plurality of antibodies will interfere with each other spatially in a region at or below the diffraction limit. In principle, the greater the number of types of antibodies there are, the more difficult it is to perform labeling evenly.

[0062] Uneven labeling leads to reconstruction of false super-resolution, which is problematic.

[0063] Proceedings of the national Academy of Sciences of the United States of America 103, 18911-18916 (2006), which discloses PAINT, does not at all describe increasing of the labeling density of target substances by labeling substances.

[0064] Thus, it is an object of the present invention to provide a super-resolution microscope observation method that can obtain position information of luminescent substances at a high density, the luminescent substances being used for labeling.

[0065] According to the method of the present invention, it is possible to label target substances at a high density in

a sample by using luminescent probes and to generate a highly accurate observation image of a target substance.

1. Principle of IRIS

[0066] The present inventors have named the observation method of the present invention IRIS (image reconstruction by integrating exchangeable single-molecule localization).

[0067] FIG. 2 (a) explains functions of a luminescent probe in the observation method of the present invention. A probe **200** contains a luminescent substance **201** that emits light under a prescribed condition (for example a fluorescent substance that emits fluorescence when irradiated with excitation light), and usually further contains a binding substance **202** that is involved in binding to and dissociation from a target substance that is linked to the luminescent substance **201**. The probe **200** can repeatedly bind to and dissociate from a target substance **210** directly and specifically. Characteristics of binding of the probe **200** are as below. When the probe **200** is in a state in which it has bound to the target substance **210**, the light emitted by the luminescent substance **201** can be picked up as a speckle in a speckle image, and when the probe **200** has dissociated from the target substance **210**, the light emitted by the luminescent substance **201** is not picked up as a speckle in an image because the probe is in a disordered thermal motion in a medium **220**. In an imaging step of the present invention, a step is performed a plurality of times (e.g. hundreds of times through hundreds of thousands of times respectively at different times) in which a speckle image including, as a speckle, light emitted from the luminescent substance **201** under a prescribed condition is obtained in a state in which the medium **220** containing the probe **200** is brought into contact with a sample containing the target substance **210**. As is shown when superposing the obtained plurality speckle images, an imaging step of the present invention can attain the same effect as that attained by labeling the target substances **210** with probes **200** at a high density and picking up images of them. Theoretically, there is no upper limit on the density of the labeling in the above. Also, when the concentration of the probes **200** in the medium **220** is adjusted appropriately, it is possible to identify a plurality of individual speckles in a separate manner in one speckle image.

[0068] Meanwhile, in PALM, STORM and Exchange-PAINT, which have conventionally been known as localization microscopy, an antibody to which a luminescent substance is linked is used for labeling a target substance. In an example of STORM for example, as shown in FIG. 2 (b), luminescent antibodies **230** resulting from linking luminescent substances **232** and antibodies **231** for the target substance **210** are made to bind to the target substance **210** and images are picked up a plurality of times (e.g. hundreds of times through hundreds of thousands of times) at different times while making the luminescent substances **232** emit light discretely at a density that is low enough to prevent the speckles from overlapping. By superposing the obtained plurality of speckle images, an image with all the luminescent substances **232** emitting light can be obtained. However, the labeling density at which the target substance is labeled with the luminescent antibodies **230** is limited by the size (about 10 nm) of the antibodies themselves. In order to achieve a resolution capability of Xnm, it is necessary to label a target substance at intervals of X/2 nm (Nyquist Sampling Theorem). The luminescent antibody **230** has a

size of at least about 10 nm, so making labeling at a high density is impossible. Also, in Exchange-PAINT, a plurality of target substances in one sample are labeled with a plurality of antibodies that are specific to the respective target substances, which causes interference between the plurality of antibodies, making it further difficult to perform labeling at a high density. The observation method according to the present invention can solve the above problem of labeling density in conventional localization microscopy.

2. Observation of a Plurality of Target Substances

[0069] The observation method according to the present invention can preferably be implemented even when a plurality of target substances exist in one sample.

[0070] When a plurality of target substances exist in one sample, the observation method according to the present invention are implemented through the following procedures. A medium containing a probe that repeatedly binds to and dissociates from one type from among a plurality of target substances directly and specifically is brought into contact with a sample so that an imaging step of the present invention is conducted and thereby a plurality of speckle images including speckles of light emitted from the probes that have bound to that one type of the target substance are obtained. Next, probes are removed by washing the sample and a medium containing a probe that repeatedly binds to and dissociates from one different type from among a plurality of target substances directly and specifically is brought into contact with the sample so that an imaging step of the present invention is conducted and thereby a plurality of speckle images including speckles of light emitted from the probes that have bound to that one different type of the target substance are obtained. These procedures are performed until a plurality of speckle images are obtained for each of all the target substances that are to be observed. The wash of the sample between the respective imaging steps can be implemented by for example performing at least one time an operation in which an appropriate washing medium such as the above medium etc. not containing the probe is brought into contact with the sample the sample is washed. Observation images of respective target substances can be generated from a plurality of speckle images for respective target substances obtained by using respective probes. Also, by superposing observation images of respective target substances in the same sample, a multiple-observation image including observation images of a plurality of target substances in the sample can also be generated.

[0071] A probe used in the present invention is a probe that can repeatedly bind to and dissociate from a target substance directly and specifically. "Bind to a target substance directly" used herein refers to binding between a probe and a target substance that is not through another binding substance such as an antibody etc. When a probe and a target substance are binding through at least one type of binding selected from a group consisting of for example hydrogen binding, hydrophilic and/or hydrophobic binding, electrostatic binding and van der Waals binding, the probe and the target substance can be treated as binding directly.

[0072] In the present invention, because a probe and a target substance bind to and dissociate from each other directly, the original state of the sample can be recovered when the sample is washed so as to remove the probe after performing an imaging step in which the sample is processed with the probe so as to make the sample emit light.

Even in a case when an operation is repeatedly performed sequentially for each target substance, in which a medium containing a probe is brought into contact with a sample, an imaging step is performed and washing is performed in order to obtain observation images of a plurality of target substances included in one sample, and each imaging step can be performed in a state where only a probe for one type of a target substance that is to be observed exists and probes for other target substances do not exist. This prevents interference from occurring between probes and makes it possible to observe each target substance in a sample that is in a more natural state.

[0073] FIG. 3 schematically shows procedures for obtaining each observation image by using one sample containing target substances A and target substances B. First, an imaging step is performed in a state where a medium containing probes A that are specific to target substances A is brought into contact with a sample so as to obtain a plurality of speckle images. Next, the sample is washed so as to remove the probes A. Thereafter, an imaging step is performed in a state where a medium containing probes B that are specific to target substances B is brought into contact with the sample so as to obtain a plurality of speckle images. An observation image is generated for each of target substance A and target substance B from a plurality of speckle images for the respective target substances. It is also possible to generate a multiple-observation image by superposing an observation image of target substance A and an observation image of target substance B.

3. Explanations for Sample and Probe

[0074] A sample used for observation is typically a biological sample such as a cell, a tissue etc. consisting of a plurality of cells. When a sample is a biological sample, it is preferable to use a sample in a fixed state, and it is particularly preferable to use a sample that has further received a permeabilization process as necessary.

[0075] A sample contains at least one target substance. "Target substance" used herein refers to an object, as an observation target, that is contained in a sample. Examples of a target substance may include a structure body that constitutes a cytoskeleton preferably such as an actin polymer, a microtubule, an intermediate filament, a focal adhesion, etc. A target substance is preferably a structure that is, like the above structure, formed as a result of many constituents assembling, the constituents having the same structure or having a structure having common characteristics. Origin organisms for a structure body that constitutes a biological sample such as a cell, a tissue, etc. that can be an observation sample and a cytoskeleton that can be a target substance are not particularly limited. For example, a structure originating from vertebrates such as mammals (humans, rabbits, rodents, etc.), amphibians (frogs etc. *xenopus* for example), fish including bony fish and cartilaginous fish, reptiles, birds, etc., invertebrates such as mollusks, protochordates, echinoderm, cnidarians, arthropods, etc., and unicellular organisms such as eukaryotic unicellular organisms (yeast etc.) can be used. Among them, structure bodies constituting cytoskeletons are held by a wide variety of species, and a method of observing a structure body constituting a cytoskeleton whose effectiveness has been confirmed in the examples can be applied regardless of the type of origin organism. It is preferable that a target substance be an object having a shape that does not substantially change

during an imaging step of picking up a plurality of speckle images, and a target substance may receive a fixation process as necessary.

[0076] A target substance is preferably a target substance containing a protein, and is more preferably a protein target substance that exits together with one or more different proteins, which are not the target substance, and that are in an observation target sample. In the present embodiment of the present invention, a probe specifically binds to a protein that is a target substance from among a plurality of types of proteins, making it possible to selectively visualize a target substance.

[0077] A probe has a property of repeatedly binding to and dissociating from one specific type of target substances in a sample directly and specifically. The meaning of "binding to a target substance directly" is as mentioned previously.

[0078] While a probe may be a luminescent substance that by itself has a property of binding to and dissociating from a target substance, it usually contains a luminescent substance and a binding substance that is linked to that luminescent substance and that has a property of repeatedly binding to and dissociating from a target substance directly and specifically.

[0079] A probe and a target substance have the following binding characteristics. Specifically, the half-life of a probe-target complex formed by binding between a probe and a target substance is preferably equal to or more than 10 milliseconds and equal to or less than 3 seconds, more preferably equal to or more than 10 milliseconds and equal to or less than 2 seconds, more preferably equal to or more than 10 milliseconds and equal to or less than 1 second, more preferably equal to or more than 10 milliseconds and equal to or less than 900 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 800 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 700 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 600 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 500 milliseconds, more preferably equal to or more than 20 milliseconds, and more preferably equal to or less than 300 milliseconds, particularly preferably equal to or less than 250 milliseconds. The above half-life herein is preferably a half-life of a probe-target complex formed by binding between a probe and a target substance in a case when a medium containing a probe and a sample containing a target substance that is to be observed are brought into contact under a condition for performing an imaging step. The half-life is defined by a period of time before the number of probes that have bound to target substances at a given moment is reduced to half through dissociation. Measurement procedures for the half-life are as below. A medium containing a probe is brought into contact with a sample containing a target substance that is to be observed and a period of time between when light emission speckle based on the probe appears and when it disappears is measured for each light emission speckle while performing observation under a condition that is used for observation. Then, periods of time between the appearance and disappearance of the speckles are plotted in accordance with a complementary cumulative relative frequency function (1-Ndissociation). Ndissociation is a cumulative relative frequency of a probe that dissociated. A cumulative relative frequency is a ratio of the speckles that disappeared within a given period of time

to the total number of measured speckles (i.e. probes) (i.e. probes that dissociated within a given period of time), and represents the total number as 1. Then, by fitting the above complementary cumulative relative frequency function with an exponent function, that half-life is calculated.

[0080] The period of time between the appearance and disappearance of a light emission speckle based on a probe can be measured by the following procedures. Specifically, while keeping a medium containing a probe in contact with a target substance and providing a prescribed condition necessary for emitting light (for example irradiation with excitation light), speckle images including speckles based on the light emission of luminescent substances of probes are consecutively picked up with an exposure time of X seconds (for example 0.050 seconds, 0.100 seconds, etc.) and at a frame rate of 1/XHz. Then, the period of time is measured between when a probe that has bound to a target appears in a speckle image and when it disappears through dissociation. In the above, when observation is performed for Y consecutive frames, and for a speckle not observed in frames before and after them, the period of time between the appearance and disappearance of that speckle can be treated as YX seconds.

[0081] A probe with the above half-life equal to or more than 10 milliseconds provides a period of time between binding to and dissociation from a target substance that is sufficiently long to allow an ordinary highly sensitive imaging device such as an EM-CCD camera etc. to pick up an image of a speckle based on the luminescent substance of the probe that has been bound. When by contrast the half-life is too long, a region including a probe having a long binding time to a target substance represents a signal that is particularly strong in a reconstructed observation image, which can cause uneven labeling. This makes it difficult to obtain an accurate distribution of target substances. From the results obtained thus far, it is possible to obtain a speckle image and an observation image that are relatively even when a probe whose half-life is equal to or less than 3 seconds is used and the shorter the half-life of a probe that is used is, the easier it is to obtain a speckle image and an observation image that are even. This point of view has made obvious that a half-life that is equal to or less than 500 milliseconds is particularly preferable. Also, as a general rule, the longer a half-life is, the longer an exposure time for one frame and imaging intervals have to be, which sometimes requires a long period of time for obtaining speckle images that are needed for generating a reconstruction image. From this point of view, it is preferable that the half-life be equal to or less than 3 seconds or that the half-life be further shorter.

[0082] In the present invention, a step of obtaining each speckle image will be referred to as a "frame imaging step". In each frame imaging step, a speckle image including a speckle of light emitted from a luminescent substance under a prescribed condition is picked up by using an imaging device. A term in which one speckle image is picked up will be referred to as a "frame", a term between the starting time of a frame and the starting time of the next frame will be referred to as an "interval", and a term between the ending time of a frame and the starting time of the next frame will be referred to as an "inter-frame term". A period of time for one frame (exposure time) can be determined in accordance with the binding half-life between a probe and a target substance in an appropriate manner. It is preferable that a period of time for one frame (exposure time) be longer than

the binding half-life between a probe and a target substance, but the scope of the present embodiment is not limited to this.

[0083] Target substances and probes that meet the above requirements can be selected appropriately while a combination between a target substance and a binding substance of a probe is preferably selected from a group of:

[0084] a combination wherein the probe is (a1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 19, (a2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (a1) where one or a plurality of amino acids have been substituted, deleted, inserted or added, and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (a3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (a1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is an actin polymer;

[0085] a combination wherein the probe is (b1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 12, which at least partially contains an amino acid sequence of 3-309 and which has 407 or fewer, preferably 357 or fewer, more preferably 327 or fewer and most preferably 307 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 14, which at least partially contains an amino acid sequence of 2536-2843 and which has 408 or fewer, preferably 358 or fewer, more preferably 328 or fewer and most preferably 308 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 14, which at least partially contains an amino acid sequence of 2781-2819 and which has 138 or fewer, preferably 88 or fewer, more preferably 58 or fewer and most preferably 38 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 4, which at least partially contains an amino acid sequence of 1-908 and which has 1008 or fewer, preferably 958 or fewer, more preferably 928 or fewer and most preferably 908 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 4, which at least partially contains an amino acid sequence of 659-908 and which has 394 or fewer, preferably 344 or fewer, more preferably 314 or fewer and most preferably 294 amino acids, an amino acid sequence of sequence number 5 or an amino acid sequence of sequence number 6, (b2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (b1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the

target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds or (b3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (b1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is a microtubule;

[0086] a combination wherein the probe is (c1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 8, which at least partially contains an amino acid sequence of 3777-4684 and which has 1008 or fewer, preferably 958 or fewer, more preferably 928 or fewer and most preferably 908 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 8, which at least partially contains an amino acid sequence of 3777-4364 and which has 688 or fewer, preferably 638 or fewer, more preferably 608 or fewer and most preferably 588 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 8, which at least partially contains an amino acid sequence of 3777-4313 and which has 637 or fewer, preferably 587 or fewer, more preferably 557 or fewer and most preferably 537 amino acids, or an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 8, which at least partially contains an amino acid sequence of 4022-4364 and which has 443 or fewer, preferably 393 or fewer, more preferably 363 or fewer and most preferably 343 amino acids, (c2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (c1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (c3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (c1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is an intermediate filament; and (d1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 15, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 15, which at least partially contains an amino acid sequence of 54-557 and which has 556 or fewer, more preferably 524 or fewer and most preferably 504 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 15, which at least partially contains an amino acid sequence of 54-498 and which has 545 or fewer, preferably 495 or fewer, more preferably 465 or fewer and most preferably 445 amino acids, an amino acid

sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 15, which at least partially contains an amino acid sequence of 167-557 and which has 491 or fewer, preferably 441 or fewer, more preferably 411 or fewer and most preferably 391 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 16, which at least partially contains an amino acid sequence of 1-251 and which has 351 or fewer, preferably 301 or fewer, more preferably 271 or fewer and most preferably 251 amino acids, an amino acid sequence which is a partial amino acid sequence of an amino acid sequence of sequence number 16, which at least partially contains an amino acid sequence of 3-251 and which has 349 or fewer, preferably 299 or fewer, more preferably 269 or fewer and most preferably 249 amino acids, or an amino acid sequence of sequence number 18, (d2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (d1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds or (d3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (d1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is a focal adhesion. It is also preferable that a more preferable scope of half-lives of probe-target complexes in the above respective combinations be in accordance with the above and particularly be a period of time that is equal to or more than 10 milliseconds and equal to or less than 500 milliseconds or shorter. In the above combinations, more preferable scopes for a half-life of a probe-target complex are as described above and it is particularly preferable that a half-life of a probe-target complex be equal to or more than 10 milliseconds and equal to or less than 500 milliseconds or that a half-life of a probe-target complex be further shorter.

[0087] Origin organisms for an actin polymer, a microtubule, an intermediate filament and/or focal adhesion in the above combinations are not particularly limited, however a substance originating from vertebrates such as mammals (humans, rabbits, rodents, etc.), amphibians (frogs etc. *xenopus* for example), fish including bony fishes and cartilaginous fishes, reptiles, birds, etc., invertebrates such as mollusks, protochordates, echinoderm, cnidarians, arthropods, etc., a unicellular organism such as eukaryotic unicellular organisms (yeast etc.), and a substance resulting from artificially introducing variation to them can be used.

[0088] "Actin polymer" used herein refers to a structure body formed through polymerization of an actin molecule, and typically to an actin filament.

[0089] In this document, a "microtubule" refers to for example a structure in which 13 protofilaments where heterodimers of tubulin and (tubulin are connected in a fibrous manner are collected to form a tubular structure body having a diameter of 25 nm.

[0090] In this document, intermediate filaments of type I, type II, type III and type IV are known as an "intermediate filament", and all of them may be observation targets of the present invention. Plectin can bind to all types of intermediate filaments.

[0091] In this document, "focal adhesion" refers to a structure made by for example a plurality of proteins (integrin, paxillin, vinculin, talin, etc.) being provided at adhesion points between a cell and an extracellular matrix.

[0092] "One or a plurality" for substitution, deletion, insertion or addition of an amino acid in the above (a2), (b2), (c2) and (d2) refers to for example 1 to 50, preferably 1 to 25, more preferably 1 to 20, more preferably 1 to 15, more preferably 1 to 10, more preferably 1 to 7, more preferably 1 to 5, more preferably 1 to 4, more preferably 1 to 3, and most preferably 1 or 2. Further preferably, the above expression of "one or a plurality" refers to preferably equal to or less than 20%, more preferably equal to or less than 10% and most preferably equal to or less than 5% of the number of amino acids of the polypeptides in the above (a1), (b1), (c1) and (d1).

[0093] Substitution of an amino acid is preferably conservative amino acid substitution. "conservative amino acid substitution" refers to substitution between amino acids having similar physicochemical functions such as an electric charge, a side chain, a polarity, an aromatic property, etc. Amino acids having similar physicochemical functions can be categorized into for example a basic amino acid (arginine, lysine, histidine), an acidic amino acid (aspartic acid, glutamic acid), a non-charged polar amino acid (asparagine, glutamine, serine, threonine, cysteine, tyrosine), a non-polar amino acid (glycine, leucine, isoleucine, alanine, valine, proline, phenylalanine, tryptophan, methionine), a branched-chain amino acid (leucine, valine, isoleucine), an aromatic amino acid (phenylalanine, tyrosine, tryptophan, histidine), etc. Addition of one or a plurality of amino acids in each amino acid sequence is preferably addition of a total of one or a plurality of amino acids to at least one of the N-terminus and the C-terminus of that amino acid sequence. Also, deletion of one or a plurality of amino acids in each amino acid sequence is preferably deletion of a total of one or a plurality of amino acids from at least one of the N-terminus and the C-terminus of that amino acid sequence.

[0094] Identity with the amino acid sequences respectively described in the above (a1), (b1), (c1) and (d1) in the above (a3), (b3), (c3), and (d3) is preferably equal to or more than 80%, more preferably equal to or more than 85%, more preferably equal to or more than 90%, more preferably equal to or more than 95%, more preferably equal to or more than 97%, more preferably equal to or more than 98%, and most preferably equal to or more than 99%. In the present invention, the value of identity of amino acid sequences is calculated in a default setting by using software that computes identity between a plurality of amino acid sequences (for example FASTA, DANASY and BLAST). The value of identity of amino acid sequences is calculated by calculating the number of amino acid residues that match when a pair of amino acid sequences is aligned in such a manner that the matching degree becomes the maximum and is calculated as a ratio of the number of the matching amino acid residues to the total number of the amino acid residues of the amino acid sequences that were compared. In this example, when there are gaps, the above total number of the amino acid residues is the number of amino acid residues obtained

by counting one gap as one amino acid residue. When thus calculated, the total numbers of all the amino acid residues are different between the amino acid sequences that are compared, and the identity is calculated on the basis of the greater total number of the amino acid residues. For a detailed method of determining identity, reference is to be made to for example Altschul et al, *Nuc. Acids. Res.* 25, 3389-3402, 1977 and Altschul et al, *J. Mol. Biol.* 215, 403-410, 1990.

[0095] Also, more preferably, polypeptides in the above (a3), (b3), (c3) and (d3) are polypeptides having preferably equal to or more than 75%, more preferably equal to or more than 80%, more preferably equal to or more than 85%, more preferably equal to or more than 90%, more preferably equal to or more than 95%, more preferably equal to or more than 97%, more preferably equal to or more than 98%, and most preferably equal to or more than 99%, of similarly to the amino acid sequences respectively described in the above (a1), (b1), (c1) and (d1). The value of similarity of amino acid sequences is calculated by calculating the total of the number of amino acid residues that match when a pair of amino acid sequences is aligned in such a manner that the matching degree becomes the maximum and the amino acid residues have similar physicochemical functions, and is calculated as a ratio of the total number to the total number of the amino acid residues of the amino acid sequences that were compared. In this example, similarity of amino acid sequences can be calculated by a computer by using software similar to that described for the identity of amino acid sequences. A method of calculating the total number of amino acid residues is as described above for amino acid identity. The meaning of amino acid residues having similar physicochemical functions is as described above.

[0096] In a preferable embodiment of the present invention, a binding substance contained in a probe (a site for identification of a target substance) is an antibody or a fragment of an antibody, to a target substance, and particularly preferably a fragment of an antibody.

[0097] A technique for producing an antibody to an arbitrary target substance has already been established. Thus, using an antibody or a fragment of an antibody produced in accordance with a target substance as a binding substance of a probe makes it possible to use the technique of the present invention for observation of a target substance ranging in a wide scope.

[0098] It is sufficient if a target substance in this embodiment is a target substance presenting antigenicity, and it is typically a protein.

[0099] An antibody that an antibody or a fragment of an antibody contained in a probe is from is typically immunoglobulin G (IgG), however a different isotype is possible, and it may be for example immunoglobulin M (IgM), immunoglobulin D (IgD), immunoglobulin A (IgA), immunoglobulin E (IgE), etc. When these types of immunoglobulin contain a plurality of subclasses, one belonging to an arbitrary subclass is possible. Examples of another type of antibody may include a single domain antibody. These antibodies may be a variation or may be in a form in which it is made to fuse with a different polypeptide.

[0100] Although origins of an antibody are not particularly limited, for example, an antibody originating from for example nonhuman animals such as mice, rats, llamas, camels, etc. and from humans etc. can be used. Also, an

antibody may be a chimeric antibody formed by causing a fusion of domains of antibodies of a plurality of origins.

[0101] An antibody may be a polyclonal antibody or may be a monoclonal antibody.

[0102] "Fragment" of an antibody in the present embodiment refers to a "functional fragment" having avidity to a target substance (antigen). A fragment of an antibody may be a variant or may be in a form in which it is made to fuse with a different polypeptide.

[0103] Examples of a fragment of an antibody may include a Fab fragment, a Fab' fragment, a F(ab')₂ fragment, an scFv (single-strand Fv) fragment, a VHH fragment of a single domain antibody (for example a commercial name of a nanobody), etc. A Fab fragment can be obtained by cutting an antibody with papain, a protein breakdown enzyme. A F(ab')₂ fragment can be obtained by cutting an antibody with pepsin. A Fab' fragment can be obtained by further processing a F(ab')₂ fragment under a reduction condition. An scFv fragment is a result of linking a heavy-chain variable region and a light-chain variable region of an antibody with a linker of a polypeptide so as to make them single-stranded and can be produced by a gene engineering method that utilizes a polynucleotide having a base sequence that encodes the heavy-chain variable region, the linker and a light-chain variable region.

[0104] In the present embodiment, the half-life of a probe-target complex formed by binding of a probe containing an antibody or a fragment of an antibody to a target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds. Generally, because an antibody has a bivalent binding capacity to an antigen (i.e., an antibody of 1 molecule has two antigen binding sites), the binding tendency to an antigen is strong and the half-life of a probe-target complex between an antibody and an antigen far exceeds the upper limit of the above scope. Thus, it is preferable that a fragment of an antibody that can repeatedly bind to and dissociate from a target substance be used and that a fragment of an antibody have a univalent binding capacity to a target substance, which is an antigen (i.e., 1 molecule has 1 antigen binding site). Examples of an antibody fragment having a univalent binding capacity to a target substance may include a Fab fragment, a Fab' fragment, an scFv fragment, a VHH fragment of a single domain antibody (for example a commercial name of nanobody), etc.

[0105] Methods of producing the above antibody and fragment of an antibody are not particularly limited. A preferable embodiment for a method of screening for an antibody presenting the above prescribed half-life from candidates for antibodies adjusted by an arbitrary method such as a hybridoma method etc. will be explained later.

[0106] Luminescent substances contained in probes are not particularly limited as long as they are substances emitting light that makes observation possible under a prescribed condition. As a luminescent substance, a fluorescent substance that can emit fluorescence when irradiated with excitation light is particularly preferable. Examples of fluorescent substances may include a fluorescent protein such as a green fluorescent protein (GFP), an enhanced green fluorescent protein (EGFP), a red fluorescent protein (RFP), TagRFP, etc.; fluorescent dye such as Atto (trademark) 488, Atto (trademark) 550, Dylight (trademark) 488, Dylight (trademark) 550, CF (trademark) dye (CF680R, CF488A, CF543, etc.), etc.; and a quantum dot.

[0107] In an imaging step of the present invention, a mixture of a medium, a probe and an observation target sample is preferably exposed at intervals that are sufficiently shorter than a period of time between when the luminescent substance starts to be exposed to the prescribed condition and when it photobleaches the light (photobleaching period of time) or may be exposed consecutively, in the prescribed condition (irradiation with excitation light for example) for making a luminescent substance included in the probe emit light. For example, in an embodiment in which a luminescent substance is a fluorescent substance, a mixture of a medium, a probe and an observation target sample may be irradiated with excitation light at intervals that are sufficiently shorter than a photobleaching period of time or may be irradiated consecutively.

[0108] A luminescent substance in a probe and a binding substance can be linked through chemical binding via an appropriate linker component as necessary. Examples of chemical binding may include covalent binding, coordinate binding, etc., and covalent binding is preferable in view of stability. When a luminescent substance and a binding substance are both polypeptides, a probe can be formed as a fused polypeptide resulting from the luminescent substance and the binding substance linking to each other through normal peptide binding.

[0109] An imaging step of the present invention is performed in a state in which a medium containing a probe and a sample are in contact with each other. Media are not particularly limited as long as they allow a probe and a target substance to maintain the above binding characteristics, however they are usually liquid, and the liquid is preferably an aqueous solution, is more preferably an aqueous buffer solution adjusted to an appropriate pH, is more preferably an aqueous buffer solution adjusted to a pH of 6.1 to pH 7.5, and more preferably contains active oxygen remover. As an active oxygen remover, at least one type or two or more types selected from glucose oxidase, catalase, 2-mercaptoethanol, glucose, etc. can be used. An example of a proper amount of glucose oxidase may be 200 µg/ml, an example of a proper amount of catalase may be 35 µg/ml, an example of a proper amount of 2-mercaptoethanol may be 0.5%, and an example of a proper amount of glucose may be 4.5 mg/ml. As a buffer component for preparing an aqueous buffer solution, HEPES, Tris, etc. can be used.

[0110] It is preferable that the concentration of probes in a medium have appropriately been adjusted to a concentration that allows identification of emission of light of a luminescent substance as a separate speckle for each molecule in one speckle image.

[0111] Temperature conditions in a case when a medium containing a probe is brought into contact with a sample are not particularly limited, and for example 20 degrees Celsius through 30 degrees, and more preferably an ambient temperature, specifically 25 degrees Celsius, can be adopted.

4. Embodiment of Observation Method of Present Invention that Uses Microscope

[0112] Devices used for an observation method of the present invention are not particularly limited.

[0113] FIG. 4 shows an example of a device that can be used for an embodiment of the present invention in which a luminescent substance contained in a probe is a fluorescent substance that emits fluorescence when irradiated with excitation light.

[0114] A microscope apparatus 1 shown in FIG. 4 includes excitation illumination devices 10 (including a first excitation illumination device 11 and a second excitation illumination device 12), a microscope body 20, a camera 30, a camera controller 40, a control unit 50, a display device 60 and a storage unit 70.

[0115] The excitation illumination devices 10 (including the first excitation illumination device 11 and the second excitation illumination device 12) are devices that provide the microscope body 20 with excitation light for making a fluorescent substance emit light. Any device can be used as the excitation illumination devices 10 (including the first excitation illumination device 11 and the second excitation illumination device 12) as long as it can provide excitation light of a wavelength in accordance with the fluorescent substance. Although they are not shown, each of the first excitation illumination device 11 and the second excitation illumination device 12 may be a system combining a plurality of devices and usually includes a laser beam source, a shutter, and a total reflection mirror. The laser beam source is a light source that emits excitation light. The shutter is a device that switches between supply and suspend of excitation light to the microscope body 20. The total reflection mirror is a mechanism for totally reflecting excitation light emitted from the laser beam source toward a stage 21 of the microscope body 20.

[0116] The microscope body 20 may be an inverted microscope for example. The microscope body 20 includes the stage 21 for mounting thereon a sample that is to be observed. To the microscope body 20, the camera 30 for picking up a fluorescence image of the sample mounted on the stage 21 is connected. As the camera 30, for example a CCD camera having a plurality of pixels can be used.

[0117] The microscope body 20 is provided with a an objective lens for issuing excitation light toward the stage 21, an imaging lens for condensing, on the light receiving plane of the camera 30, fluorescence radiated from a fluorescent substance in the sample, etc. The above objective lens and the imaging lens constitute an imaging optical system.

[0118] The microscope body 20 including the stage 21 and the imaging optical system is configured to be able to provide total internal reflection illumination by which excitation light is totally reflected on the interface between the glass in contact with the sample and the sample. The total internal reflection illumination makes it possible to illuminate the sample with evanescent light that leaks to the sample side from the glass when the excitation light is totally reflected. In the shown embodiment, the second excitation illumination device 12 is equivalent to a device for providing excitation light through total internal reflection illumination.

[0119] The microscope body 20 is further configured to be able to switch between the above total internal reflection illumination and epi-illumination to use them. In the shown embodiment, the first excitation illumination device 11 is equivalent to a device for providing excitation light based on epi-illumination. Using a speckle image of fluorescence based on the total internal reflection illumination of excitation light and a speckle image of fluorescence based on epi-illumination makes it possible to calculate the position in the depth direction (z direction) in an observation image. Alternatively, it is also possible to make a probe emit intensive fluorescence by issuing excitation light based on

epi-illumination and excitation light based on total internal reflection illumination simultaneously.

[0120] The excitation illumination devices 10 do not always have to include the first excitation illumination device 11 and the second excitation illumination device 12 and may have only one of the first and second excitation illumination devices 11 and 12.

[0121] The control unit 50 is a computer that totally controls the microscope apparatus 1, and is connected to the excitation illumination devices 10 (first excitation illumination device 11 and second excitation illumination device 12), the display unit 60, the storage unit 70, and the camera controller 40. The control unit 50 has at least a control signal generation function of generating a control signal for controlling these devices, a speckle image obtainment function of obtaining a speckle image via the camera controller 40, an image analysis function of analyzing an obtained speckle image, a light source control function of controlling driving of a light source included in the excitation illumination devices 10, and an image forming function of generating an observation image from a plurality of speckle images. The control unit 50 constitutes an image generation unit that generates an observation image as the overall function.

[0122] The camera controller 40 performs driving control of the camera 30. The camera controller 40 operates the camera 30 on the basis of a control signal input from the control unit 50 so as to obtain a speckle image of fluorescence, and outputs the obtained speckle image to the control unit 50.

[0123] The display unit 60 is a display (display device), a printer (printing device), etc., and provides a function of displaying and printing an image based on image data output from the control unit 50 (data of a speckle image, data of an observation image, etc.).

[0124] The storage unit 70 can be configured by a storage device such as a semiconductor memory, a hard disk etc. A program used in the control unit 50 and data provided from the control unit 50 (such as a speckle image etc.) are stored in a state in which they can be read by the control unit 50.

[0125] Hereinafter, explanations will be given for the observation method of the present invention, by the microscope apparatus 1, that uses a fluorescent substance emitting fluorescence when irradiated with excitation light as a luminescent substance contained in a probe.

[0126] An imaging step using the microscope apparatus 1 is a step in which a step is performed a plurality of times at different times respectively, where a sample that has been brought into contact with a medium containing a probe on the stage 21 is irradiated with excitation light by using the excitation light illumination devices 10 (one or both of the first excitation illumination device 11 and the second excitation illumination device 12) and a speckle image including, as a speckle of fluorescence, fluorescence emitted from a substance contained in a probe that has bound to a target substance in the sample is obtained by the camera 30, and thereby a plurality of speckle images are obtained. A step of obtaining each speckle image will be referred to as a "frame imaging step". In each frame imaging step, the camera 30 picks up a speckle image of fluorescence emitted from a fluorescent substance each time excitation light is issued. A speckle image has a different pattern for each frame.

[0127] In each frame imaging step, the control unit 50 operates the camera 30 via the camera controller 40 so as to pick up an image of fluorescence emitted from a fluorescent

substance, and thereby obtains a speckle image. An obtained speckle image is output to the control unit 50 from the camera controller 40. The control unit 50 also stores a thus-obtained speckle image in the storage unit 70.

[0128] In an imaging step, the control unit 50 repeatedly performs the above frame imaging step at appropriate intervals. Examples of the number of times of the frame imaging step may include hundreds of times through hundreds of thousands of times including for example 1,000 to 999,000 times.

[0129] An observation image generation step in the present invention is a step in which an observation image of a target substance that binds to the probe in the sample is generated from a plurality of speckle images obtained in the above imaging step. Specifically, the control unit 50 executes an appropriate computer program so as to obtain, for each of the plurality of speckle images recorded in the storage unit 70, information of position of a speckle included in a speckle image so as to generate an observation image by integrating the pieces of information from the plurality of speckle images. In this example, information of the position of each speckle is typically information of the central position (position of the center of gravity) of each speckle, and can be obtained by using for example DAOSTORM, a computer program (Nature methods, 8 279-280, 2011). In the above, in order to increase the accuracy, only a speckle of luminance that is equal to or higher than a prescribed threshold may be used for generating an observation image in each of the speckle images. When generating an observation image, an observation image can be generated by drawing, at the central position of each speckle of a blank image, a point having an appropriate size. Specifically, by turning the size of pixels of a blank image into an appropriate size (for example a square pixel with the length of each side being 5 nm through 20 nm) and by plotting the central position of each speckle to each pixel, an observation image can be generated. The control unit 50 records the data of the generated observation image in the storage unit 70 and also displays it in the display unit 60.

[0130] When a plurality of target substances exist in one sample, it is sufficient to perform the above imaging step sequentially by using probes that are specific to the respective target substances. "Probes that are specific to respective target substances" refer to probes that repeatedly bind to and dissociate from the respective target substances directly and specifically. It is preferable that the sample be washed sufficiently between the respective imaging steps. Then, the control unit 50 can generate an observation image of each target substance in the sample from a plurality of speckle images, stored in the storage unit 70, obtained in the respective imaging steps. The control unit 50 can also synthesize pieces of data of observation images of a plurality of target substances in the same sample so as to output the result to the display unit 60 and display a multiple-observation image resulting from superposing observation images of a plurality of target substances.

[0131] The above is an embodiment of the observation method of the present invention that uses a probe containing a fluorescent substance as a luminescent substance. When a luminescent substance is a luminescent substance that is not a fluorescent substance, the method of the present invention can be implemented through the same procedures as that described above if means for providing a condition in a sample mounted on the stage 21 for that luminescent sub-

stance to radiate light instead of the excitation illumination devices **10** in the microscope apparatus **1** are arranged and the camera **30** that can pick up an image of light emitted by the luminescent substance in the probe that has bound to a target substance in the sample as a speckle is used.

5. Probe and Kit

[0132] The present invention also provides the above probe itself and a kit including at least the above probe.

[0133] The above probe may be provided solely in solid form such as powder, may be provided in liquid form that has dispersed or has been dissolved in an appropriate liquid medium or may be provided in solid form such as powder together with an appropriate solid component (such as a diluting agent etc.). In other words, a probe of the present invention may be provided solely by itself or may be provided as a probe-containing composition that includes at least a probe and may include a different complementary component.

[0134] Any kit may be used as the above kit as long as it includes at least a probe of the above various forms and it may further include a different element used for observing a target substance. Examples of a different element for the above kit may include a medium such as a liquid medium for dissolving or dispersing the probe etc., and a reagent to be used for a process of observing a sample etc. The above medium may be a medium that includes a probe and that is used for bringing it into contact with a sample for performing observation. The probe and a different element in the above kit are usually wrapped separately so that they will not be mixed physically.

6. Screening Method

[0135] The present invention also provides a screening method of a site (also referred to as a target substance identification site or a binding substance) in which a target substance is identified in the probe accordingly, the screening method including:

[0136] an immobilization step in which a candidate substance of the site or a substance partially containing the candidate substance is fixed to a solid support;

[0137] an observation step in which a target substance linked to a luminescent substance and a solid support obtained in the immobilization step are observed in a medium while the target substance linked to a luminescent substance and the solid support obtained in the immobilization step are kept in contact, in a condition that allows observation, in units of 1 molecule, of light emission from the luminescent substance in a probe-target complex formed by binding between the target substance and the candidate substance, and

[0138] a screening step in which the candidate substance resulting in a half-life of the probe-target complex that is equal to or more than 10 milliseconds and equal to or less than 3 seconds is selected as the site on the basis of observation in the observation step.

[0139] According to this embodiment of the present invention, it is possible to efficiently perform screening for a substance that presents a desired binding tendency to a target substance from a library of candidate substances.

[0140] Examples of a solid support used for an immobilization step may include an inner wall surface of each well of for example a multiwell plate.

[0141] Means for fixing a candidate substance to a solid support in an immobilization step are not particularly limited, and when for example a candidate substance is an antibody or an antibody fragment, it is preferable that a Fab domain of an antibody or an antibody fragment be immobilized to the solid support in a state in which it can bind to the target substance. For example, by fixing to a solid support a protein having a binding capability to an Fc domain of an antibody (for example, Protein G) and then immobilizing the antibody to the solid support to which the protein has been immobilized, it is possible to immobilize the antibody to the solid support in a state in which the Fab domain can bind to the target substance. Methods of immobilizing the protein to a solid support are not particularly limited, however it is possible to immobilize the protein via a functional group introduced to a surface of a solid support.

[0142] As a medium used in an observation step, the same medium as that used in an imaging step of the present invention can be used.

[0143] Types of luminescent substances are not particularly limited in a target substance linked to the luminescent substance used in an observation step, however a fluorescent substance that can emit fluorescence when irradiated with excitation light is particularly preferable. Specific examples of a fluorescent substance are the same as those of the fluorescent substance described above for a probe.

[0144] In an observation step, in a medium, a target substance linked to a luminescent substance and a solid support to which a candidate substance is fixed are observed in a state in which they are in contact with each other, in a condition that allows observation, in units of 1 molecule, of emission of light by the luminescent substance in a probe-target complex of the target substance and the candidate substance. In this step, the fact that light emission of a target substance that has not bound cannot be detected because it causes thermal motions randomly in the medium, whereas emission of light from the target substance that has bound to the candidate substance can be detected and is utilized. In order to observe emission of light by the luminescent substance in units of 1 molecule, it is effective to reduce the concentration of the target substance in the medium. Even when a candidate substance is a bivalent antibody or antibody fragment having two antigen binding sites, reducing the concentration of the target substance in the medium makes it possible to observe binding of the target substance individually in each antigen binding site.

[0145] Observation means in an observation step may appropriately be selected in accordance with the luminescent substance. When the luminescent substance is a fluorescent substance, observation is possible by using a fluorescence microscope (for example, a TIRF fluorescence microscope).

[0146] In a screening step, on the basis of observation in the observation step, the candidate substance leading to the half-life of the probe-target complex that is equal to or more than 10 milliseconds and equal to or less than 3 seconds is selected as a target substance identification site. The half-life is more preferably equal to or more than 10 milliseconds and equal to or less than 2 seconds, more preferably equal to or more than 10 milliseconds and equal to or less than 1 second, more preferably equal to or more than 10 milliseconds and equal to or less than 900 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 800 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 700 milliseconds,

more preferably equal to or more than 10 milliseconds and equal to or less than 600 milliseconds, more preferably equal to or more than 10 milliseconds and equal to or less than 500 milliseconds, more preferably equal to or more than 20 milliseconds and equal to or less than 300 milliseconds, and particularly preferably equal to or less than 250 milliseconds. The half-life is defined by a period of time before the number of target substances that have bound to candidate substances on solid supports at a given moment is reduced to half through dissociation. Measurement procedures for the half-life are as below. A medium containing a target substance labeled with a luminescent substance is brought into contact with a solid support to which the candidate substance has been fixed, and a period of time between when a light emission speckle appears and when it disappears is measured, for each light emission speckle, while performing observation under a condition used for observation. Then, periods of time between the appearance and disappearance of the speckles are plotted in accordance with a complementary cumulative relative frequency function (1-Ndissociation). Then, by fitting the above complementary cumulative relative frequency with an exponent function, that half-life is calculated.

[0147] A period of time between appearance and disappearance of a light emission speckle based on a labeled target substance is measured by the following procedures. Specifically, while keeping a medium containing a labeled target substance in contact with a solid support to which the candidate substance has been immobilized and providing a prescribed condition necessary for emitting light (for example irradiation with excitation light), speckle images including speckles based on the light emission of the labeled target substance are consecutively picked up with an exposure time of X seconds (for example 0.050 seconds, 0.100 seconds, etc.) and at a frame rate of 1/XHz. Then, a period of time is measured between when a target substance that has bound to a candidate substance appears in a speckle image and when it disappears through dissociation. In the above, observation is performed for Y consecutive frames, and for a speckle not observed in frames before and after them, the period of time between the appearance and disappearance of that speckle can be treated as YX seconds.

[0148] In a screening method of the present invention, the candidate substance is more preferably an antibody or a fragment of an antibody that binds to a prescribed target substance and fixes the antibody to a solid support in the immobilization step. When screening is performed for antibodies binding to prescribed target substances, ELISA is generally used in which target substances are immobilized to solid supports and candidate antibodies are brought into contact with the solid supports. In a screening method according to the present embodiment, contrary to ELISA, an antibody (the candidate substance) is solid phased. Then, in the medium, an antigen (target substance) linked to a luminescent substance and the solid support are observed in a state in which they are in contact with each other, in a condition that allows observation, in units of 1 molecule, of emission of light by the luminescent substance in a probe-target complex formed by binding of the target substance and the candidate substance. This method has advantages as described below.

[0149] 1) It is possible to measure the half-life of binding of an antibody and an antigen. Because binding/dissociation between an antibody and an antigen is visualized through the

appearance and disappearance of a fluorescent single molecule, the half-life of that binding can be measured directly for each single molecule of an antigen.

[0150] 2) It is possible to measure univalent binding/dissociation of an antigen and an antibody. Epitope that allows identification of a monoclonal antibody is at one location in an antigen. Accordingly, an antigen that has bound to 1 antigen binding site of a solid-phased antibody cannot bind to a different antigen binding site existing nearby, and an antigen inevitably binds with the antibody molecule in a univalent state. When an antigen has been solid phased, an antibody can bind to one or two antigens, making it impossible to measure univalent binding/dissociation of an antigen and an antibody. A method in which this antibody is solid phased makes it possible to estimate the half-life of binding of an antibody fragment such as a Fab fragment that binds in a univalent manner without producing it.

[0151] 3) It is possible to perform screening for an antibody fragment from a library of hybridoma in an inexpensive and simple manner. In one embodiment of this screening method, main structure factors are a solid support resulting from immobilizing an antibody contained in each supernatant of a hybridoma library and a labeling body of antigen epitope. The half-life of binding of an antibody fragment and an antigen can be estimated even without producing a light-emission-labeled antibody fragment from an antibody of each supernatant of a library, making it possible to perform screening on candidate substances at a very low cost. Also, visualization is performed in units of molecules, measurement is possible even when the amount of antibodies that have been solid phased is small. Thereby, immense amounts of labor and time taken for cultivation of a hybridoma library can be saved.

[0152] The present screening method has solved a major problem in evaluation of the affinity of an antibody by ELISA. 1) In ELISA, an antigen is solid phased, and accordingly an antibody may bind to an antigen regardless of whether the binding is univalent or bivalent. Further, by a secondary antibody used for measuring a binding amount of primary antibodies, a primary antibody is crosslinked. This prevents measurement of the half-life of univalent binding of an antibody and an antigen. 2) The binding amount of antibodies obtained in ELISA is influenced not only by the affinity of antibodies but also by the amount of antibodies produced by hybridoma. This makes it difficult to evaluate the property of binding of one antibody in ELISA. 3) In addition, because washing is conducted a plurality of times in a reaction step in ELISA, there is a risk that an antibody that has bound with a short half-life that is equal to or less than 3 seconds, which is needed in the present invention, may be lost before being visualized. The method of the present embodiment has solved these problems by solid phasing an antibody instead of an antigen and visualizing binding/dissociation of an antigen and an antibody in actual time. The method of the present embodiment is more suitable for selecting an antibody having a short binding half-life, which is the aim of the present invention, than ELISA in principle in that it can evaluate univalent binding/dissociation, it is not influenced by the production amount of antibodies, and it can also measure a short binding half-life.

[0153] According to a screening method of the present invention, a library of polypeptides and a library of phage display can be solid phased as candidate substances for a

target substance identification site of a probe in addition to an antibody or an antibody fragment. A screening method of the present invention can also be applied to solid-phasing of these for performing screening for a polypeptide that binds to and dissociates from a target substance.

[0154] The present invention will be explained in more detail by the following examples, however they are just exemplary and do not limit the present invention. All the experiments below were conducted at an ambient temperature, i.e., 25 degrees Celsius, unless limitations are particularly given.

1. Experiment 1

Methods

Plasmid and Reagent

[0155] Expression plasmid (pFLAG-EGFP-C1) encoding EGFP (sequence number 1) having an N terminus tagged with FLAG (sequence number 2) and an expression plasmid (p3xFLAG-EGFP-N3) encoding EGFP having a C terminus tagged with 3xFLAG (sequence number 3) were constructed respectively by using a pEGFP-C1 vector and a pEGFP-N3 vector (Clontech Laboratories, Inc).

[0156] EST clones encoding mouse MAP4, human Tau isoforms 3 and 4, mouse KIF1A, human plectin-1 and *Xenopus laevis* talin-1 were purchased from OpenBiosystems.

[0157] cDNA encoding human FAK was purchased from the DNASU Plasmid Repository.

[0158] The GenBank/EMBL/DDBJ accession numbers for each sequence are as follows:

[0159] BC055332 (MAP4), BC114948 (Tau isoform 3), BC101936 (Tau isoform 4), BC062891 (KIF1A), BM559026 (Plectin-1), CF282569 (Talin1) and BC035404 (FAK).

[0160] cDNAs encoding human EB1, rat CLIP-170, human CLASP2 γ and human APC were provided by Y. Mimori-Kiyosue (RIKEN).

[0161] As plasmids encoding human paxillin, chicken Src and human vinculin, those described in documents 31 to 33 were used.

[0162] Each cDNA was inserted into a pFLAG-EGFP-C1 vector or a p3xFLAG-EGFP-N3 vector by using a PCR. For a probe having an N terminus at the fusion position of GFP in table 1 below, a pFLAG-EGFP-C1 vector was used. For a probe having a C terminus at the fusion position of GFP in table 1 below, a p3xFLAG-EGFP-N3 vector was used.

[0163] An expression plasmid encoding PIPK1 γ fragment (amino acid residues 641-668) having an N terminus tagged with FLAG-EGFP was constructed by inserting a synthetic cDNA encoding PTDEKSWVYSPLHYSAQAPPASDGEEDT (sequence number 18) into a pFLAG-EGFP-C1 vector.

[0164] Lifeact peptide (MGVADLIKKFESISKEE (sequence number 19)) with an Atto 488 fluorescent body linked to an N terminus was purchased from Sigma-Aldrich.

[0165] The amino acid sequence of EGFP is denoted by sequence number 1.

[0166] The amino acid sequence of FLAG is denoted by sequence number 2.

[0167] The amino acid sequence of 3xFLAG is denoted by sequence number 3.

[0168] The amino acid sequence of mouse MAP4 is denoted by sequence number 4.

[0169] The amino acid sequence of human Tau isoform 3 is denoted by sequence number 5.

[0170] The amino acid sequence of human Tau isoform 4 is denoted by sequence number 6.

[0171] The amino acid sequence of mouse KIF1A is denoted by sequence number 7.

[0172] The amino acid sequence of human Plectin-1 is denoted by sequence number 8.

[0173] The amino acid sequence of *Xenopus laevis* Talin1 (amino acid residues 1-2353) is denoted by sequence number 9.

[0174] The amino acid sequence of human FAK is denoted by sequence number 10.

[0175] The amino acid sequence of human EB1 is denoted by sequence number 11.

[0176] The amino acid sequence of rat CLIP-170 is denoted by sequence number 12.

[0177] The amino acid sequence of human CLASP2 γ is denoted by sequence number 13.

[0178] The amino acid sequence of human APC is denoted by sequence number 14.

[0179] The amino acid sequence of human Paxillin is denoted by sequence number 15.

[0180] The amino acid sequence of chicken Src is denoted by sequence number 16.

[0181] The amino acid sequence of human Vinculin is denoted by sequence number 17.

[0182] The amino acid sequence of human PIPK1 γ -90 fragment (amino acid residues 641-668) is denoted by sequence number 18.

[0183] The amino acid sequence of Lifeact peptide is denoted by sequence number 19.

Production and Screening of Exchangeable Protein Probes

[0184] In order to find exchangeable probes for super-resolution images of microtubules, intermediate filaments and focal adhesions, a test was conducted by using, as candidate molecules for probes, a protein (polypeptide) and a protein fragment that are known to be able to localize in each target structure. "Exchangeable" used herein refers to being able to repeatedly bind to and dissociate from a target structure.

[0185] Table 1 shows the tested probe candidates. The test was conducted a plurality of times by using, as a probe candidate, a protein or a fragment of a protein mentioned in the section of "plasmid and reagent".

[0186] An expression plasmid for expression as a molecule made to fuse with EGFP was constructed for each probe candidate in accordance with the documents described in table 1.

[0187] HEK (Human embryonic kidney)-293F cells were transfected with a plasmid encoding a probe candidate protein tagged with FLAG-EGFP or 3xFLAG-EGFP. 3 to 4 days later, the cells were dissolved in a cell lysis buffer (10 mM HEPES, pH 7.2, 90 mM KCl, 3 mM MgCl₂, 0.2% Triton X-100, 100 μ M DTT) containing a protease inhibitor cocktail (Nacalai Tesque). Centrifugal separation was performed on the lysate and the supernatant liquid was collected. In order to perform screening for a probe for IRIS, the binding capacity of a probe candidate in the supernatant liquid with respect to the structure of an XCT cell that was fixed with paraformaldehyde (PFA) and that received a

permeabilization process was tested. The appearance and disappearance of single molecule speckles (Single-molecule speckles, SiMS) in the structure was tested. Screening was performed for a probe for IRIS in accordance with the following criteria:

- (1) It is possible to confirm a distribution of target substances in the structure in an image resulting from adding SiMS images;
- (2) It is possible to wash and remove a probe after SiMS imaging (imaging step);
- (3) A probe that has bound can dissociate from the target substance swiftly (half-life is equal to or less than 500 ms); and
- (4) It is possible to reconstruct an image of a target substance by integrating the central positions of the respective speckles.

[0188] Table 1 describes that “localization” is positive (P) when above criterion (1) is met, that “washability” is positive (P) when above criterion (2) is met, and that “IRIS image” is positive (P) when above criterion (3) is met.

[0189] Probes used for an IRIS experiment were purified in the following procedures. Specifically, each probe was overexpressed in HEK-293F cells and collected with anti-DYKDDDDK (Flag) antibody beads (Wako). The beads were washed four times with an excess amount of HEPES-buffered solution (10 mM Hepes pH 7.2, 90 mM KCl, 3 mM MgCl₂, 100 μ M DTT). Proteins that had bound to beads after the washing were processed with the HEPES-buffered solution containing 0.5 mg/ml DYKDDDDK (Flag) peptide (Wako) or 3 \times FLAG peptide (Sigma-Aldrich) for 30 minutes and were eluted.

[0190] The localization test of the above (1), whose result is shown in table 1, was performed in the following procedures. First, the supernatant liquid of lysate of a cell that expressed each probe candidate was brought into contact with an XCT cell that was fixed on a coverglass and that received a permeabilization process. A coverglass on which the sample was mounted was arranged in the observation chamber of a fluorescence microscope apparatus, which will be described in detail in the section of “procedures for imaging of multicolor super-resolution by IRIS” below, and speckle images of 10000 frames were obtained with an exposure time of 50 ms or 100 ms for one frame and at a frame rate of 20 Hz (20 frames per second) or with an exposure time of 100 ms and at a frame rate of 10 Hz (10 frames per second) while irradiating the sample with a 488 nm-laser beam line (with a main body output of 50 mW but reaching the sample after being attenuated by AOTF etc.) for total internal reflection fluorescence observation. The obtained speckle images of 10000 frames were integrated so as to confirm whether or not a distribution of target substances in the cell (microtubule, intermediate filament, focal adhesion or actin filament) was able to be confirmed. When a distribution of a target substances was able to be confirmed, the result of the localization test was treated as P (positive).

[0191] The localization test of the above (2), whose result is shown in table 1, was performed in the following procedures. After the picking up of speckle images explained in the previous paragraph, 1 ml of the supernatant liquid containing respective probe candidates in the observation chamber was aspirated by using an aspirator and 1 ml of an imaging solution (however, it was not supplemented with active oxygen-scavenging mix) that does not contain a probe

and that will be described below was added. This switching of imaging solution was conducted slowly so that the observation position would not shift. Next, an imaging solution not containing a probe and not containing the active oxygen-scavenging mix was switched 10 to 20 times. After the switching, for confirmation, for an XCT cell sample in the observation chamber, speckle images of 10 frames were picked up under a similar condition to that of the localization test described in the previous paragraph, and when the number of speckles confirmed in the speckle images (i.e., the number of probes that had bound) became sufficiently smaller (equal to or less than about 10%) than the number of the speckles in the speckle images before the washing operation, washing was determined to be possible and the result was treated as P (positive).

[0192] The measurement of binding half-life, whose result is shown in table 1, was performed in the following procedures. Speckle images of 10000 frames picked up in the localization test were used in order to measure a period of time between the appearance of a probe that has bound to a target in a speckle image and disappearance through dissociation in a semi-manual mode by using Speckle TrackerJ, an ImageJ plug-in. In case of for example picking up of images with an exposure time of 50 ms and at a frame rate of 20 Hz, it is assumed that a period of time between the appearance and disappearance of a speckle that is observed in only one of consecutive frames is 50 ms and a period of time between the appearance and disappearance of a speckle observed in only two consecutive frames is 100 ms. Then, the number of binding probes with respect to periods of time between the appearance and disappearance was plotted in accordance with a complementary cumulative relative frequency function (1-Ndissociation). Ndissociation is a cumulative relative frequency of probes that dissociated. Then, by fitting the complementary cumulative relative frequency function with an exponent function, the half-life was calculated. However, the binding half-life with respect to an actin filament of Atto488-Lifeact was measured by using a method that is described in detail in the section for FIG. 9 in the Brief Description of the Drawings.

[0193] The IRIS image test of the above (3) whose result is described in table 1 was performed in the following procedures. By using all the speckle images of 10000 frames picked up in the localization test, the central point of a probe that has bound to a target in each frame of the speckle images was determined with nanometer accuracy by using DAOSTORM. By adding a large number of pieces of central point information in the fluorescence images of 10000 frames, a reconstruction image (observation image) was generated. Whether or not a distribution of a target molecule was able to be observed in that reconstruction image was determined, and when a distribution of a target molecule was observed, the result was treated as P (positive).

Procedures for Imaging of Multicolor Super-Resolution by IRIS

[0194] *Xenopus laevis* XTC cells were cultured in 70% Leibovitz's L15 medium supplemented with 10% fetal bovine serum. A multicolor super-resolution image was produced from a large number of fluorescence single molecule speckle (SiMS) images that were sequentially obtained from a fixed and XTC cell (20,000 to 500,000 frames per probe) with various exchangeable probes. The cells were allowed to spread on a 0.1 mg/ml poly (L-lysine) and 10

g/ml fibronectin-coated coverglass in 70% Leibovitz's L15 medium without serum and distinct stress fibers and focal adhesions were formed (document 33). 2 hours later, the cells were fixed and received a permeabilization process with a cytoskeleton buffer containing 3.7% PFA and 0.5% Triton X-100 in (10 mM Mes pH6.1, 90 mM KCl, 3 mM MgCl₂, 2 mM glycol ether diamine tetraacetic acid (EGTA)). After performing blocking with 4% bovine serum albumin for 30 minutes, the purified IRIS probes were brought into contact with the cells in an imaging solution including the HEPES-buffered solution (10 mM Hepes pH 7.2, 90 mM KCl, 3 mM MgCl₂, 100 μM DTT) with an oxygen-scavenging mix (200 μg/ml glucose oxidase, 35 μg/ml catalase, 4.5 mg/ml glucose, 0.5% 2-mercaptoethanol) (document 34). The concentration of the probe was 1 nM through 100 nM. When the oxygen-scavenging mix was not used, laser-induced photodamage was apparent after obtaining SiMS images several tens of thousands of times.

[0195] For the imaging of actin filaments in vitro, monomeric actin was prepared from rabbit skeletal muscle in a method described in documents 33, 35 and 36. Phalloidin-stabilized F-actin was observed on a 1 mg/ml poly (L-lysine)-coated coverglass in the imaging solution.

[0196] SiMS images were obtained by using an inverted microscope (Olympus IX83-ZDC) equipped with an Olympus PlanApo 1.45×100 through a numerical aperture (NA) objective lens, a 2×intermediate lens and an EM-CCD camera (Evolve 512, Roper), and controlled by MetaMorph software (Molecular Device). The focus was automatically maintained at the bottom of the cell by a z drift compensation system during the long-term imaging. The IRIS probe was alternately excited with a 473-nm laser beam (50 mW) for epi-illumination microscopy and a 488-nm laser beam (50 mW) for epi-fluorescence observation in the following procedures. In the epi-illumination mode, the incidence angle of the 473-nm laser beam was tilted so as to reduce background fluorescence from out-of-focus probes that have not bound. The epi-fluorescence image and TIRF image were used to estimate the z position of the target object (see below). Specifically, images were picked up by repeating the following procedures:

- (a) imaging in bright field;
- (b) imaging of a SiMS image (speckle image) with epi-fluorescence (exposure time for one frame: 50 ms, frame rate: 20 Hz (20 frames per second), number of frames picked up consecutively: 250 frames); and
- (c) imaging of SiMS image (speckle image) with TIRF (exposure time for one frame: 50 ms, frame rate: 20 Hz (20 frames per second), number of frames picked up consecutively: 250 frames).

[0197] An image obtained in a bright field was used to correct a drift of the microscope stage in a lateral direction (see below) In case of observation of focal adhesions, epi-fluorescence observation was not performed and TIRF observation was performed (frame rate: 20 Hz, 500 frames). Each one of the procedures required 27 seconds and was repeated 160 to 240 times (probe/target substance=CLIP-170 fragment) (amino acid residues 3-309 of sequence number 12)/microtubule, 40 times (probe/target substance=PIPK1γ fragment (641-668) (sequence number 18), Paxillin (overall length of sequence number 15) and the Src fragment (amino acid residues 3-251 of sequence number 16)/focal adhesion), 800 times (probe/target substance=Lifect (sequence number 19)/actin filament),

and 400 to 480 times (probe/target substance=Plectin-1 fragment (amino acid residues 4022-4364 of sequence number 8)/intermediate filament). In order to maintain the oxygen-scavenging capacity in the imaging solution, an imaging solution containing the probe was replaced with a fresh imaging solution every 40 sets when the CLIP-170 fragment, the PIPK1γ fragment, paxillin and the Src fragment were used and every 80 sets when Lifect and the plectin-1 fragment were used. For multicolor imaging of three types of cytoskeletons and focal adhesions, a pick up of SiMS images was conducted in the order of the CLIP-170 fragment, the PIPK1γ fragment (or the Src fragment and paxillin), Lifect and the plectin-1 fragment. After obtaining SiMS images on the basis of each probe, washing was conducted 10 times by using the HEPES-buffered solution. The remaining fluorescence of a probe was completely photobleached in the HEPES-buffered solution supplemented with an oxygen-scavenging mix, and the next probe was made to react.

[0198] In the present example, imaging steps of obtaining observation images were conducted under the above conditions unless otherwise described.

Procedures for Image Reconstruction in IRIS

[0199] A super-resolution image was reconstructed by plotting the central points of each fluorescent speckle on a blank image with subpixel accuracy. The number of plotted points was typically 10⁶ to 10⁸. A central point was estimated with subpixel accuracy by fitting of a point-spread function (PSF) of this microscope using a computer program known as DAOSTORM (document 14). In order to correct a stage drift of the microscope, the drift distance was calculated by an autocorrelation function, i.e., $A_N(x_{drift}, y_{drift})$ of the bright-field images obtained at each set of imaging procedures.

$$A_N(x_{drift}, y_{drift}) = \quad \text{[Numerical expression 1]}$$

$$\sum_{y=y_0}^{y_m} \sum_{x=x_0}^{x_m} [I_0(x, y) \times I_N(x + x_{drift}, y + y_{drift})]$$

where x_{drift} and y_{drift} are drift distances in the directions of the x axis and y axis, respectively. $I_0(x, y)$ and $I_N(x + x_{drift}, y + y_{drift})$ are intensities at the pixel positions (x, y) and (x + x_{drift} , y + y_{drift}) in the bright field images obtained in the 1st and Nth sets, respectively.

[0200] The product of $I_0(x, y)$ and $I_N(x + x_{drift}, y + y_{drift})$ is integrated within a prescribed region in the bright-field image. $A_N(x_{drift}, y_{drift})$, which is a function of variables x_{drift} and y_{drift} , becomes the maximum when the two bright-field images coincide. The x_{drift} and y_{drift} values leading to a maximum value as $A_N(x_{drift}, y_{drift})$ were calculated by using a customized plug-in in ImageJ software (<http://rsb.info.nih.gov/ij/>). First, the drift of the bright-field image in the Nth set was corrected using the x_{drift} and y_{drift} values with pixel accuracy. To further determine the drift distance with subpixel accuracy, the bright-field image and the corrected image in the first and Nth sets were enlarged by using a bicubic method. Using the $A_N(x_{drift}, y_{drift})$ of the enlarged images, the drift distances were determined with subpixel accuracy. The central positions of speckles in the SiMS images in the Nth set were corrected with the drift distances.

By plotting the corrected central positions, a super-resolution image was produced. The positions of speckles consecutively observed in 10 or more frames or in 20 or more frames were not used when generating a reconstruction image (observation image) based on Lifeact or generating a reconstruction image (observation image) based on the plectin-1 fragment. These two probes in some cases bound to a target substance when strong excitation laser output was employed.

Image Process for Mapping z Position of Observation Target Object

[0201] Because a TIRF excitation light intensity exponentially decreases with increasing distance from the coverglass, the height of the observation target object was estimated from the ratio of a TIRF image and an epi-fluorescence image. The z position from the coverglass surface was measured by the method, described in document 18, using fluorescent microtubules that were tilted with respect to the coverglass in a low-melting-point agarose gel. HyLight 488-labeled tubulin was purchased from Cytoskeleton. The labeled microtubules were prepared according to the method described in document 18. Images of the tilted microtubules each having one end touching the coverglass were picked up by TIRF and epi-fluorescence. Epi-fluorescence images were obtained as z stack images (0.2- μm step size) (FIG. 14 (a)). In the intensity line profile along the microtubules of epi-fluorescence images, the x-y position of the highest intensity was used to determine the intersection of the tilted microtubule and the focal plane (FIG. 14 (a) arrows). By connecting the intersections among the z-stacked images, the z-directional distance of each point along the tilted microtubule was obtained. The z profile of the excitation light intensity based on a TIRF method was determined by associating the z-directional distance of each point with the ratio of the intensity of the microtubule in a TIRF image and the intensity of the microtubule in an epi-fluorescence image (FIG. 14 (b)). This z profile was fitted with a single exponential decay function (FIG. 14 (b)). The scale of the z positions were again adjusted by a factor of 0.82, taking into consideration the difference in the refractive index between the immersion oil and the imaging solution (document 18). The inverse function of the exponential function was used to determine the z position of the observation target object by the following numerical expression.

$$z = -\alpha_z \ln \left(\beta \frac{F^{TIRF}}{F^{Epi}} \right) \quad \text{[Numerical expression 2]}$$

(where α_z is the z position at which the intensity of TIRF illumination is $1/e$, β is a parameter for calibrating the

difference in the laser output between TIRF observation and epi-fluorescence observation, and F^{TIRF} and F^{Epi} are the fluorescence intensities of the target object in a TIRF image and an epi-fluorescence image, respectively)

[0202] The z-position maps of three types of cytoskeletons were converted from the image of a ratio of the IRIS image (observation image) based on TIRF to the IRIS image (observation image) based on an epi-fluorescence image. For this purpose, the peak intensity of the fluorescent speckle was also fitted by using DAOSTORM. The IRIS images were reconstructed by plotting the peak intensity at the central position of the speckle. In the obtained z-position map, the fluorescence intensity in an image resulting from adding the IRIS image based on TIRF and the IRIS image based on epi-fluorescence was masked by a threshold and noise in a region having no cytoskeletons was removed. In a layered structure such as an actin stress fiber, etc., the calculated z position represents the height of the center of gravity of the structure in the z axial direction.

[0203] The z position of the microtubule plus end traced by live-cell imaging of EB1-EGFP was obtained by converting a ratio of an average intensity in a 0.4- μm -diameter region of the microtubule plus end in the TIRF image to that in a corresponding region in the epi-fluorescence image.

Live-Cell Imaging of the Movement of Microtubule Plus End

[0204] XTC cells were transfected with an expression plasmid of EGFP fused EB1. 3 to 4 days later, live-cell imaging of EB1-EGFP was conducted on the cells at one-second intervals. In each interval term, an imaging with an exposure time of 100 ms of fluorescence by excitation light for epi-fluorescence observation and imaging with an exposure time of 100 ms of fluorescence by excitation light for total reflection fluorescence observation were conducted. In the above, each excitation light was emitted under the above conditions for IRIS super-resolution except that the excitation light was emitted after having its laser power reduced to about 20% of that used for an IRIS super-resolution imaging in order to avoid damaging live cells. At each imaging time point, two fluorescence images were obtained by alternately using total internal reflection illumination and epi-illumination with an exposure time of 100 milliseconds. An EB1-labeled microtubule plus end was traced by using Speckle TrackerJ, an ImageJ plug-in (documents 33 and 37). The z position of the traced microtubule plus end was calculated by the above method. The speed of the site of the microtubule plus end was calculated with a linear approximation of its x-y positions at five consecutive imaging time points.

Results

[0205] The results of screening tests on probe candidates were as below.

TABLE 1

TARGET SUBSTANCE	PROBE	SEQUENCE NUMBER	AMINO ACID NUMBER	EGFP FUSION POSITION	LOCALIZATION	WASH-ABILITY	HALF-LIFE	IRIS IMAGE	DOCUMENT
MICROTUBULE	CLIP-170	12	3-309	N TERMINUS	P	P	44 ms	P	41
	APC	14	2536-2843	C TERMINUS	P	P	100 ms	P	43.44
	APC	14	2781-2819	N TERMINUS	P	P	27 ms	P	43.44
	MAP4	4	1-908	N TERMINUS	P	P	109 ms	P	47

TABLE 1-continued

TARGET SUBSTANCE	PROBE	SEQUENCE NUMBER	AMINO ACID NUMBER	EGFP FUSION POSITION	LOCALIZATION	WASHABILITY	HALF-LIFE	IRIS IMAGE	DOCUMENT
	MAP4	4	659-908	N TERMINUS	P	P	106 ms	P	47
	TAU ISOFORM 3	5	1-383	N TERMINUS	P	P	110 ms	P	48,49
	TAU ISOFORM 4	6	1-352	N TERMINUS	P	P	60 ms	P	48,49
INTERMEDIATE FILAMENT	Plectin-1	8	3777-4684	N TERMINUS	P	P	457 ms	P	16
	Plectin-1	8	3777-4364	N TERMINUS	P	P	59 ms	P	16
	Plectin-1	8	3777-4313	N TERMINUS	P	P	52 ms	P	16
	Plectin-1	8	4022-4364	N TERMINUS	P	P	103 ms	P	16
	Plectin-1	8	4066-4364	N TERMINUS	P	P	7.7 s	NA	16
FOCAL ADHESION	Paxillin	15	1-557	N TERMINUS	P	P	196 ms	P	50
	Paxillin	15	54-557	N TERMINUS	P	P	246 ms	P	50
	Paxillin	15	54-498	N TERMINUS	P	P	91 ms	P	50
	Paxillin	15	167-557	N TERMINUS	P	P	138 ms	P	50
	Src	16	1-251	C TERMINUS	P	P	161 ms	P	59
	Src	16	3-251	C TERMINUS	P	P	141 ms	P	59
	PIPKly90	18	641-668	N TERMINUS	P	P	496 ms	P	17
ACTIN FILAMENT	Atto488-Lifect	19	—	—	—	—	23 ms	—	—

[0206] In the table, P represents positive while NA indicates that measurement was not conducted (not accessed).

[0207] In the sequence table, sequence number 18 represents an amino acid sequence of fragments 641-668 of PIPKly90.

[0208] A probe candidate that had not passed either a localization test or a washing test received neither a test of an IRIS image nor half-life measurement, and thus is not described in the above table 1. A probe candidate that did not pass a test is estimated to have a very long or very short binding half-life to a target substance or to not bind to a target substance.

[0209] Because the test described in detail in FIG. 9 and the explanations for the figure have independently confirmed that Atto488-Lifect is effective for an actin filament and that the half-life thereof is 23 nm, these facts are described in the above table as reference results.

Super-Resolution of Actin Filament in High Labeling Density by IRIS

[0210] The present inventors confirmed whether or not a super-resolution image can be generated through the IRIS method by using Lifect, which is a widely used actin marker. Lifect is a short peptide that stains an actin filament in a live cell or a fixed cell (document 10). Lifect has a property of being exchanged within 0.4 seconds on an actin filament (document 10). The high-speed exchangeability of Lifect is confirmed through single-molecule speckle (SiMS) microscopy (documents 11 to 13). The dwell time of Atto488-labeled Lifect showed a single exponential decay of a half-life of 23 milliseconds (FIG. 9 (a)). The present inventors selected a concentration of 2.4 nM as the Atto488-labeled Lifect concentration in the imaging solution and obtained 2×10^5 SiMS images of Lifect on an actin filament in vitro through consecutive imaging using a total internal reflection illumination of 488 nm with an exposure time of 50 ms/frame and at a frame rate of 20 Hz. The central position of each fluorescent speckle was determined by using a computer program named DAOSTORM (document 14) (FIG. 9 (b)). Pieces of position information from a great number of Lifect speckles in the above speckle images of 2×10^5 frames were integrated so as to reconstruct an image of actin filaments (FIG. 5 (b) and FIG. 9 (c)). The average

width of single actin filaments was 23 nm as the full-width at half-maximum (FWHM) in an image reconstructed by using only high-brightness speckles (highest 12% approximately) in order to guarantee a high accuracy of localization (FIG. 5 (c) and FIG. 9 (d)).

[0211] A major problem in conventional super-resolution microscopy is that using an antibody and a photoactivatable fluorescent protein makes it difficult to label an observation target structure in a sufficient density. The actin subunit and the antibody have the widths of 6 nm and 12 nm, respectively, and accordingly a single actin filament having 360 subunits per 1 μ m can only be used for labeling up to a density of at most 180 subunits per 1 μ m in the labeling of antibodies. This labeling density is equivalent to the labeling density in an observation image reconstructed from speckle images of 2×10^3 frames in the IRIS method of the present inventors. As shown in the left view of FIG. 5 (d) and in FIG. 5 (e), actin filaments show a pattern that is not continuous in the longitudinal direction in an observation image reconstructed from speckle images of 2×10^3 in the IRIS method. Even a labeling density that was 6.5 times its original was not sufficient for continuous staining of actin filaments (FIG. 5 (d)), however it was possible to achieve a labeling density of 1.2×10^4 for an observation image reconstructed from speckle images of 2×10^5 frames and to obtain consecutive super-resolution images of actin filaments (FIG. 5 (d) and FIG. 5 (e)). As described above, it was made obvious that the IRIS method of the present invention can eliminate the problem of a labeling density that had conventionally been an obstacle to dissolving two or three types of target substances coexisting close to each other.

[0212] For a cell that was fixed and had received a permeabilization process, in an observation image generated by using the IRIS method that utilizes a Lifect probe of an actin filament, it was possible to dissolve two parallel actin bundles that were apart by 50 nm (FIG. 5 (d) through FIG. 5 (f)). By contrast, in an image resulting from adding SiMS images (speckle images) (which is equivalent to an image that can be obtained by a conventional immunofluorescence method), it was not possible to dissolve two such actin bundles (lower left view of FIG. 5 (g)). According to the IRIS method, it is possible to obtain consecutive observation

images of an actin filament or a microtubule (below). This is remarkable progress from the conventional super-resolution microscopy (FIG. 10).

Establishment of Screening Method of IRIS Probe

[0213] The present inventors established an effective method for swiftly determining an IRIS probe for a different cell structure while taking into consideration a necessary molecular characteristic determined by data using a Lifeact probe. A probe candidate was generated by making a protein and EGFP fuse with each other, the protein and the EGFP being known to bind to a target substance. Live-cell fluorescence single molecule speckle (SiMS) microscopy (documents 11 and 12) is also effective for testing a probe candidate. However, the present inventors discovered that the binding specificity and dissociation dynamics of a probe candidate can easily be clarified by bringing crude lysate of a cell that expresses EGFP fused probe candidate into contact with a fixed cell. The inventors selected an IRIS probe in the following four steps:

- (i) SiMS images (speckle images) of 10,000 frames were picked up at intervals of 50 ms or 100 ms, and probe candidates that were not able to localize in targets in an integrated SiMS image were excluded;
- (ii) probe candidates that were not able to be easily removed through washing were excluded;
- (iii) probe candidates having high dissociation speeds (with a half-life of 10 ms through 500 ms, see FIG. 11) were selected; and
- (iv) it was confirmed, in a reconstruction IRIS image (observation image), that probe candidates were able to localize in targets.

[0214] As shown in table 1, 18 types were selected by the above screening method from among the probe candidates. By a plurality of probes for microtubule and focal adhesion, different sites of their structure bodies were able to be visualized (FIG. 12 and FIG. 13). Also, independently from the above, it has been confirmed that Atto488-Lifeact is effective as a probe for an actin filament and is effective for the visualization of an actin filament (FIG. 9 etc.). This suggests that IRIS is effective for mapping a distribution of probe identification sites in one structure body.

Super-Resolution Image of a Plurality of Target Substances by IRIS

[0215] It was discovered that Lifeact, a CLIP-170 fragment (residues 3-309), a Pleclin-1 fragment (residues 4022-4364) and a phosphatidylinositol(4)-phosphate 5-kinase type I γ -90 (PIPKI γ) fragment (residues 641-668) are preferable respectively for observation of actin, microtubules, intermediate filaments and focal adhesion. By utilizing the exchangeability of IRIS probes, images in which the IRIS probes bound to 4 different types of cytoskeleton structures were obtained sequentially.

[0216] Further, the present inventors investigated a three-dimensional (3D) network of an actin filament, a microtubule and an intermediate filament. An image based on each IRIS probe was obtained by alternately using total internal reflection illumination and epi-illumination, and super-resolution images at the bottom and in the entire peripheral region of the cell were reconstructed respectively by using the obtained images. These images show that actin arcs running parallel to the cell contour gradually rise as if they

were climbing on the radial actin bundles localizing at the bottom (arrows in FIG. 6 (a)) with decreasing distance to the center. It was confirmed that microtubules and intermediate filaments behind the lamellipodium (lobopodium) base were eliminated from the cell bottom at several locations (arrows in FIG. 6 (b) and FIG. 6 (c)).

[0217] The z position of each observation target object was estimated by using a signal ratio between an IRIS image based on TIRF and an IRIS image based on epi-fluorescence on the basis of measurements that used images of tilted fluorescent microtubules described in an above method (document 18). The 3D images as shown in FIG. 15 clarified the structure of the target substance. Lamellipodia (LP), stress fibers (SF) and actin arcs (Arc) were distributed with height positions of their centers of gravity (average \pm S.D.) of 42 ± 43 nm, 34 ± 30 nm and 217 ± 132 nm, respectively (arrows in FIG. 15 (a) and FIG. 15 (d)). The z positions of the microtubules became lower from 150 nm through 200 nm to 50 nm through 100 nm in the vicinity of the cell perimeters (arrowheads in FIG. 15 (b) and FIG. 15 (e)). Intermediate filaments formed mesh-shaped structures throughout the cell body, and some of the filaments were located at a height of approximately 200 nm behind the lamellipodium (ellipse in FIG. 15 (c)).

[0218] An IRIS image makes it possible to observe a spatial relationship between a plurality of cytoskeletal structures in a single cell at a resolution capability exceeding the diffraction limit. In a lamellar region, intermediate filaments were tangled with actin stress fibers in many cases, but they are not tangled with microtubules (FIG. 7 (a) through FIG. 7 (c)). The tangled actins and intermediate filaments appear to be linked to each other. Their cross-sectional profiles show that actin stress fibers overlapped intermediate filaments (arrows in FIG. 7 (e)). In peripheral regions, intermediate filaments did not tangle with actin stress fibers (FIG. 7 (g) and FIG. 7 (i)), whereas some intermediate filaments ran along microtubules (arrows in FIG. 7 (h)). The cross-sectional profiles show that intermediate filaments overlapped microtubules but did not overlap actin filaments (arrows in FIG. 3 (i), and FIG. 3 (j)). Thus, IRIS can reveal a region-specific proximity between 4 cytoskeletal components.

Behavior of Plus End of Microtubule in Live Cell and Comparison with Super-Resolution of Cytoskeleton Network

[0219] The present inventors discovered that the heights of microtubules locally change in the vicinity of focal adhesions and stress fibers. When microtubules crossed focal adhesions and stress fibers, they were lifted to positions at 200 nm from positions at 100 nm from the glass surface (arrowheads in FIG. 8 (a), FIG. 8 (b) and FIG. 16). Components of focal adhesions are located at a height of 30 nm through 80 nm from the glass surface (document 19). It appears that the lifted microtubules climbed on actin stress fibers without touching focal adhesions.

[0220] The present inventors further investigated a relationship between a microtubule plus end and a cytoskeleton network in that portion by observing the behavior of EB1-EGFP in a live cell and then reconstructing a super-resolution image of a 3D cytoskeleton network after fixation. The imaging system of the present inventors makes it possible to perform live-cell 3D imaging with acousto-optic tunable filters for swiftly switching between total internal reflection illumination and epi-illumination. The trajectory of the tip of

the EB1-labeled microtubule was compared with focal adhesions and stress fibers in a super-resolution image (asterisk in FIG. 8 (c)). When the tip of the microtubule glowing was brought into contact with a stress fiber, the tip moved upward so as to move away from a focal adhesion existing below (FIG. 8 (d)). When the tip of the EB1-labeled microtubule was brought into contact with a stress fiber, the growth slowed down and the moving direction changed (arrow in FIG. 8 (d)). These pieces of data suggest that the speed and the direction of growth of a microtubule are greatly influenced by the collision and subsequent interaction with an actin stress fiber. As described above, a combination between IRIS and live-cell imaging makes it possible to clarify formation processes of a plurality of cytoskeletal structures that dynamically interact with each other.

Consideration

[0221] Consideration 1: Comparison with PAINT

[0222] Document 20 (Proceedings of the national Academy of Sciences of the United States of America 103, 18911-18916 (2006)) has reported a method called PAINT (point accumulation for imaging in nanoscale topography). This document has reported that super-resolution observation based on PAINT was conducted on lipid bilayer morphology by using Nile-red, which is a fluorescent dye that rapidly shuttles between an aqueous solution and the lipid bilayer. However, Nile-red is not a probe that binds to and dissociates from a specific bilayer molecule type. This prevents highly accurate determination of the position of a specific lipid molecule type that constitutes a lipid layer. By contrast, a probe of IRIS can bind to and dissociate from a specific target protein from among various types of proteins existing in a cell so as to make it possible to determine the position of the molecule thereof highly accurately. In addition, the concept of PAINT does not include a concept of obtaining a distribution of a plurality of target substances through probe exchange of IRIS. High resolution-capability imaging of various target substances by IRIS cannot be realized by the concept of PAINT. Also, this document has does not disclose improving of resolution by increasing the number of times of observation in PAINT. The reason appears to be that even an increased number of times of observation does not fix a lipid layer, which is a target substance, and thus the resolution does not improve through a change in the shape. In addition, the reason also seems to be that the problem of labeling density first became widely recognized a few years later than the document as an unsolved problem among those skilled in the art (documents 4 through 7).

[0223] The problem of labeling density was proposed in 2008 (document 6), whereas it had long been an unsolved problem in super-resolution microscope observation (documents 4, 5 and 7). As a method of increasing a target density, the review of 2013 (document 5) introduced an attempt to make fluorescent dye, which is an organic compound that is smaller than an antibody and a fluorescent protein, stably bind to a target substance and to make a single domain antibody such as Nanobody etc., which stably binds, stably bind to a target substance. This means that PAINT had not been recognized as a method of increasing a target density. Also, even the method introduced in the above review did not achieve a labeling density equivalent to that realized by the present invention, and the problem of a labeling density

had remained unsolved. From these points, it can be said that there had not been an idea of using a binding and dissociation probe for solving the problem of a labeling density. In FIG. 10, an observation image of a microtubule based on IRIS and those based on STORM and Exchange-PAINT, which are conventionally known super-resolution microscopy, are compared (documents 21 to 23). According to the line profiles of label intensities along the longitudinal direction of microtubules, the observation image based on IRIS shows a more continuous pattern than those of observation images based on STORM and Exchange-PAINT (FIG. 10 (b)). Also, it is easy for the labeling density of actin filaments based on IRIS to become 60 times the maximum density of antibodies that bind to actin filaments. From these results, it is obvious that IRIS according to the present invention can solve the problem of labeling density that has caused a deterioration in reliability of the conventional super-resolution microscopy.

[0224] In addition to the ability to overcome the problem of labeling density, according to IRIS of the present invention, it is easy to visualize a plurality of target substances by protein-based exchangeable probes and there are no limitations on the number of target substances. In super-resolution microscopy that has conventionally been known and available, only up to 2 target substances can be observed (documents 21, 22, 24 and 25). Recently, Exchange-PAINT, which makes it possible to conduct super-resolution observation of a plurality of target substances, was announced (document 23 (Nature methods, 11, 313-318, 2014)). Exchange-PAINT uses characteristic labeling means of hybridizing short and fluorescence-labeled DNA with complementary DNA conjugated to antibody molecules. Exchange-PAINT is similar to IRIS in that it sequentially visualizes a plurality of target substances one by one by sequentially hybridizing pairs of various types of oligonucleotides. However, in Exchange-PAINT, uneven labeling and/or interference between antibodies sometimes lead to inaccurate analysis of the distribution of labeling substance (FIG. 10 (b)). By contrast, an IRIS probe is washed and removed and thereafter the next IRIS probe is brought into contact with the sample, resulting in no interference between a plurality of probes that are respectively for a plurality of target substances. The data of the present inventors shows the region-specific proximity of three types of cytoskeleton structures and focal adhesions with an accuracy exceeding the diffraction limit. In principle, the number of target substances that can be observed by IRIS is not limited even when they coalesce in a narrow region. This effect is remarkably advantageous in comparison with existing super-resolution microscopy.

Consideration 2: Probe

[0225] As shown in the above table, the present inventors have confirmed that probes of Document 19 are actually effective for the IRIS method. The above probes have been probes that use a binding partner known to associate with a target substance material, however the scope of the invention is not limited to this example. Using a phage display or an assay that confirms interactions between proteins of yeast-two hybrid system etc. makes it possible to perform screening for useful IRIS probes.

[0226] A plurality of IRIS probes for a microtubule and a focal adhesion visualized different sites of their structure bodies. A MAP4 fragment and a tau protein weakly visualized a microtubule plus end when EB1 existed (FIG. 12 (e)).

A CLIP-170 fragment strongly labeled a microtubule tip when EB1 existed and continuously labeled the entire microtubule when EB1 did not exist (FIG. 12 (c) and FIG. 12 (e)). The result corresponds to in-vitro data (document 26). Further, Paxillin and the Src fragment visualized different portions of focal adhesion (FIG. 13). Thus, IRIS can also be applied to mapping analysis of a site in which a plurality of protein fragments bind. Development of an IRIS probe that binds to a molecule in a specific state also makes it possible to perform super-resolution mapping on the molecule in the specific state.

2. Experiment 2

[0227] 2.1. Method of Producing a Fab Probe from a Polyclonal Antibody

[0228] An anti-p40 antibody including antiserum was produced from a rabbit by using an antigen *Xenopus laevis* derived p40 produced in the present inventors' laboratory. Rabbit antiserum was put into an affinity column filled with antigens and polyclonal antibodies were made to adsorb on the column. pH in the column was reduced in a stepwise manner (pH5 to pH2) and the antibodies made to adsorb were eluted. 2 ml of fraction eluted at a high pH (pH2 through pH3.5) was added to 1.4 ml of a 50% slurry solution of Protein A Protein beads (Protein A Sepharose CL-4B, GE) and the antibodies were made to adsorb on the beads over 1 night at 4 degrees Celsius. After using PBS to wash and remove antibodies that had not been made to adsorb, the beads were suspended in 1 ml of PBS. To this suspension of beads, 50 μ l of 1 μ g/ μ l DyLight 488 NHS Ester (Thermo-Scientific) in which DMSO was dissolved was added, and antibodies were fluorescence-labeled at an ambient temperature over one hour. DyLight 488 NHS Ester that had made no reactions was washed and removed by using PBS and only beads were collected through centrifugation. In order to produce a Fab fragment from an antibody that was made to adsorb on beads, 1 ml of 7 μ g/ml papain dissolved in a Digestion Buffer (50 mM Tris-HCl pH8.0, 10 mM Cysteine-HCl, 2 mM EDTA) was added and reactions were caused for 1 hour at 37 degrees Celsius. After the centrifugation, supernatant containing a Fab fragment was collected and 1 μ l of 1 mg/ml leupeptin was added in order to inhibit the activity of papain. The obtained Fab fragment was put into an affinity column filled with antigens, and the Fab fragment was made to adsorb. pH in the column was reduced in a stepwise manner and a Fab fragment of a fraction eluted at a high pH (pH 5 to pH 4) was obtained. As described above, in the production of a Fab fragment from a polyclonal antibody, column purification was conducted twice. By using the affinity column of the first time, an antibody having a weak antigen-binding force was purified. From that antibody, a Fab fragment having a further weak antigen-binding force was produced. By using the affinity column of the second time, a Fab fragment in which an antigen-binding force still remained was purified. A Fab fragment prepared in this method was derived from IgG.

2.2. Iris Super-Resolution Imaging of Actin

[0229] Preparation of Arp2/3 complex observation cell sample By following the procedures described in detail in "procedures for imaging of multicolor super-resolution by IRIS" of experiment 1, a *Xenopus laevis* XTC cell was fixed and received a permeabilization process. After a blocking

step with 4% bovine serum albumin for 30 minutes, the Fab probes were brought into contact with the cells in an imaging solution comprising the HEPES-buffered solution (10 mM Hepes pH 7.2, 90 mM KCl, 3 mM MgCl₂, 100 μ M DTT, 0.1% Triton X-100) supplemented with an oxygen-scavenging mix (200 μ g/ml glucose oxidase, 35 μ g/ml catalase, 4.5 mg/ml glucose, 0.5% 2-mercaptoethanol). The Fab probe concentration was 100 nM in the imaging solution.

Imaging and Image Reconstruction

[0230] Similarly to "procedures for imaging of multicolor super-resolution by IRIS" of experiment 1, SiMS images (speckle images) were obtained by using an inverted microscope (Olympus IX83-ZDC) equipped with an Olympus PlanApo 100 \times /1.45-numerical aperture (NA) objective lens, a 2 \times intermediate lens and an EM-CCD camera (Evolve 512, Roper) and controlled by MetaMorph software (Molecular Device). The Fab probe fluorescence-labeled by being irradiated with a 488 nm laser beam (50 mW) was excited for TIRF observation. SiMS images (speckle images) of a total of 33,750 frames were picked up by conducting consecutive imaging of 250 frames for each under a condition that the exposure time was 100 milliseconds for 1 frame and the frame rate (imaging speed) was 10 Hz (10 frames per second).

[0231] The procedures for image reconstruction from the above SiMS images of 33,750 frames are as described in detail in "procedures for image reconstruction in IRIS" of experiment 1.

[0232] The half-life of a probe-target complex formed between a purified Fab probe and a target antigen was obtained in a similar method to that in Experiment 1. In other words, by using the SiMS images picked up in the above, a period of time between when the Fab probe that had bound to the target appeared in a speckle image and it disappeared through dissociation was measured in a semi-manual mode by using Speckle TrackerJ, a plug-in of ImageJ. Then, the number of binding probes with respect to the period of time between the appearance and disappearance was plotted in accordance with a complementary cumulative relative frequency function (1-Ndissociation). Then, by fitting the complementary cumulative relative frequency function with an exponent function, the half-life was calculated. The result showed that the half-life of a probe-target complex formed between the Fab probe and a target antigen was 216 milliseconds.

2.3. Results

[0233] FIG. 17 shows a reconstruction image based on the Fab probe generated from a polyclonal antibody derived from a rabbit with respect to a p40 subunit of an Arp2/3 complex, which is an actin polymerization promotion factor. It is known that much p40 is distributed in a peripheral region of a cell and a distribution of Fab probes was visualized as such.

3. Experiment 3

[0234] Screening of Fab Probe

[0235] A library (1200 samples) of hybridoma was generated by using FLAG peptide (sequence number 2) as an antigen. After immunizing a mouse with an antigen, lymphocytes were collected from an iliac lymph node and were made to fuse with a mouse myeloma cell, and hybridoma

was generated. The library of hybridoma was made through dilution culture. Culture supernatant was collected for each sample from the hybridoma library and was made to react with a 96 well glass bottom plate in which Protein G was solid phased and thereby an antibody in the culture supernatant was fixed to the surface of the bottom glass.

[0236] The 96 well glass bottom plate in which Protein G was solid phased and immobilization of the antibody were conducted in the following procedures. (3-aminopropyl) triethoxysilane was dissolved in a mixture of methanol and acetic acid (mixing ratio 100:5) and a 3% solution was produced. This solution was put in a 96 well multi-well plate having each of its wells made of glass, the solution was incubated for 30 minutes at an ambient temperature, and the glass surfaces of the well bottoms were coated. (3-aminopropyl) triethoxysilane that had made no reactions was washed a plurality of times with methanol so as to remove it and thereafter the plate was air dried so that (3-aminopropyl) triethoxysilane was made to remain on the glass surfaces. Next, a 0.1% glutaraldehyde solution was added to each well so as to cause reactions for 30 minutes at an ambient temperature and glutaraldehyde was made to bind to the glass surfaces. A 50 ng/ μ l Protein G (Thermo Scientific) solution was added to each well and was kept in contact with the glass surfaces to which glutaraldehyde bound, for 1 night at 4 degrees Celsius. Protein G causes covalent bonding with (3-aminopropyl) triethoxysilane via glutaraldehyde and is solid phased on the glass surfaces. Blocking was conducted on the glass surfaces on which Protein G was solid phased, with a 3% BSA solution for 1 night at 4 degrees Celsius. Thereafter, a hybridoma culture supernatant containing an antibody was added to each well and was left to stand for 1 night at 4 degrees Celsius. An antibody contained in the culture supernatant binds, specifically and strongly, to Protein G via the Fc domain thereof and is fixed to the glass surface. After removing the hybridoma culture supernatant, the glass surfaces were washed with a cell lysis buffer (10 mM Hepes pH 7.2, 3 mM MgCl₂, 0.2% Triton X-100, 100 μ M DTT), and received an observation step, which will be described later. In order to obtain a fused protein of FLAG peptide and FLAG peptide that becomes an antigen, an HEK-293F cell was transfected with a plasmid that encoded FLAG-EGFP. After culturing the transfected cell for 3 to 4 days, it was dissolved in a cell lysis buffer (10 mM Hepes pH 7.2, 3 mM MgCl₂, 0.2% Triton X-100, 100 μ M DTT) containing a protease inhibitor cocktail (Nacalai Tesque). After centrifugally separating the lysate, the supernatant liquid was collected. The FLAG-EGFP concentration contained in the supernatant liquid was estimated from the light emission intensity of EGFP, and a 50 nM FLAG-EGFP solution was prepared by diluting it with a cell lysis buffer.

[0237] A 50 nM FLAG-EGFP solution was added to each well to which an antibody was fixed. By using a TIRF microscope, SIMS images (speckle images) of 500 frames were picked up with an exposure time of 50 ms for 1 frame and at a frame rate of 20 Hz (20 frames per second). As a result of observing 1200 wells, 10 to 300 speckles (0.006 to 0.178 speckles/ μ m²) were observed in the field of view of the microscope (41 μ m \times 41 μ m). Each speckle corresponds to EGFP of a single molecule. As examples of speckle images, FIG. 18A shows an image in which about 250 speckles were observed and FIG. 18B shows an image in which about 20 speckles were observed. In the case where 50 nM FLAG-EGFP solutions were added to wells in which

antibodies had not been solid phased, the numbers of the speckles in the field of view were 10 to 20 (0.006 to 0.012/ μ m²). Then, the half-lives of binding between antibodies and FLAG-EGFP were measured by treating, as positive examples, a 26 well in which 40 (0.023/ μ m²) or more speckles had been observed in the field of view. Each of the wells had different half-lives. The half-life in the well shown in FIG. 18A was 55 ms.

[0238] From the 26 well in which 40 or more speckles had been observed in the field of view of the microscope as the positive examples, antibodies having half-lives of the probe-target complex that were equal to or more than 10 milliseconds and equal to or less than 3 seconds were selected. Hybridoma producing these antibodies was converted into monoclonal and 2 ml to 10 ml of the culture supernatant was collected. 40 μ l of a 50% slurry solution of Protein A beads (Protein A Sepharose CL-4B, GE) was added to the culture superresolution, and the antibodies were made to adsorb on the beads for 1 night at 4 degrees Celsius. After using PBS to wash and remove antibodies that had not adsorbed, the beads were suspended in 100 μ l of PBS. To this suspension of beads, 16 μ l of 0.5 μ g/ μ l DyLight 488 NHS Ester (ThermoScientific) dissolved in DMSO was added, and the antibodies were fluorescence-labeled for 1 hour at an ambient temperature. DyLight 488 NHS Ester that had made no reactions was washed and removed with PBS and only beads were collected after centrifugation. In order to produce a Fab fragment from an antibody that bound to the beads, 20 μ l of 0.01 mg/ml papain dissolved in Digestion Buffer (50 mM Tris-HCl pH8.0, 10 mM Cysteine-HCl, 2 mM EDTA) was added and reactions were caused for 1 hour at 37 degrees Celsius. After the centrifugation, supernatant containing a Fab fragment was collected and 2 μ l of 0.01 mg/ml leupeptin was added in order to inhibit the activity of papain.

2.2. IRIS Super-Resolution Imaging of Actin

[0239] Preparation of a FLAG Tag Fused Actin Expression XTC Cell Sample

[0240] FLAG tag fused actin encoded by the base sequence described by sequence number 20 was expressed with an XTC cell.

[0241] Expression vector-(CLONETECH) encoding pEGFP-actin was cut with restriction enzyme Age-I, Bgl-II (NEB) to remove EGFP from the vector. Synthetic cDNA encoding FLAG peptide (DYKDDDDK) was inserted into the vector and an expression plasmid encoding an actin for which FLAG was tagged to an N terminus was constructed.

[0242] A *Xenopus laevis* XTC cell was transfected with the above expression plasmid in the following procedures. 3 μ l of an expression plasmid of 1 μ g/ μ l of FLAG fused actin was dissolved in 200 μ l of a serum-free medium (70% dilution Leibovitz's L-15 medium) and 8 μ l of Polyethyleneimine, linear, M.W. 25,000 (Polysciences) of 1 mg/ml was added and it was vortexed. After leaving it to stand for 30 minutes at an ambient temperature, a medium with serum of 1 ml (70% dilution Leibovitz's L-15 medium, 10% FCS supplement) was added. The solution containing this plasmid and the media of XTC cells spread in 6 wells were switched, and the cells were made to take in plasmids for 1 night.

[0243] 3 to 4 days later than the transfecting, the cells were fixed and received the permeabilization process in the procedures described in detail in "procedures for imaging of multicolor super-resolution by IRIS" of experiment 1. After

a blocking step with 4% bovine serum albumin for 30 minutes, the Fab probes were each brought into contact with the cells in an imaging solution comprising the HEPES-buffered solution (10 mM Hepes pH 7.2, 3 mM MgCl₂, 100 μM DTT, 1 μg/ml leupeptin) supplemented with an oxygen-scavenging mix (200 μg/ml glucose oxidase, 35 μg/ml catalase, 4.5 mg/ml glucose, 0.5% 2-mercaptoethanol). The Fab probe concentration was 3 nM in the imaging solution.

Imaging and Image Reconstruction

[0244] Similarly to “procedures for imaging of multicolor super-resolution by IRIS” of experiment 1, SiMS images (speckle images) were obtained by using an inverted microscope (Olympus IX83-ZDC) equipped with an Olympus PlanApo 100×/1.45-numerical aperture (NA) objective lens, a 2×intermediate lens and an electron-multiplying EM-CCD camera (Evolve 512, Roper) and controlled by MetaMorph software (Molecular Device). The Fab probe fluorescence-labeled by being irradiated with a 488 nm laser beam (50 mW) was excited for TIRF observation. SiMS images (speckle images) of a total of 120,000 frames were picked up by conducting consecutive imaging of 500 frames for each under a condition such that the exposure time was 50 milliseconds for 1 frame and the frame rate (imaging speed) was 20 Hz (20 frames per second).

[0245] The procedures for image reconstruction from the above SiMS images of 120,000 frames are as described in detail in “procedures for image reconstruction in IRIS” of experiment 1.

2.4. Results

[0246] FIG. 19 shows IRIS super-resolution images of FLAG fused actin by a Fab probe derived from an anti-FLAG monoclonal antibody selected by the above screening method. The half-life of a probe-target complex of a Fab probe and an FLAG fused actin used for the generation of these IRIS super-resolution images was 203 milliseconds.

[0247] The present screening method made it possible to select a Fab probe appropriate for IRIS super-resolution imaging.

4. Documents

[0248] The documents referred to herein are described below.

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- [0306] The present invention can provide a super-resolution microscope observation method that makes it possible to obtain, at a high density, position information of luminescent substances used for labeling and to generate a high resolution observation image exceeding the diffraction limit, and also it is made possible to clarify the formation process of a plurality of cytoskeleton structures that interact dynamically, by a combination between the present invention and a living-cell image technique, and thus the present invention has a high industrial applicability.
- [0307] All publications, patents and patent applications referred to herein are incorporated herein in their entirety by reference.

SEQUENCE LISTING

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Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe
 210 215 220
 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 225 230 235

<210> SEQ ID NO 2
 <211> LENGTH: 8
 <212> TYPE: PRT
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: FLAG

<400> SEQUENCE: 2

Asp Tyr Lys Asp Asp Asp Lys
 1 5

<210> SEQ ID NO 3
 <211> LENGTH: 22
 <212> TYPE: PRT
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: 3xFLAG

<400> SEQUENCE: 3

Asp Tyr Lys Asp His Asp Gly Asp Tyr Lys Asp His Asp Ile Asp Tyr
 1 5 10 15

Lys Asp Asp Asp Lys
 20

<210> SEQ ID NO 4
 <211> LENGTH: 1125
 <212> TYPE: PRT
 <213> ORGANISM: Mus musculus

<400> SEQUENCE: 4

Met Ala Asp Leu Ser Leu Val Asp Ala Leu Thr Glu Pro Pro Pro Glu
 1 5 10 15

Ile Glu Gly Glu Ile Lys Arg Asp Phe Met Ala Ala Leu Glu Ala Glu
 20 25 30

Pro Tyr Asp Asp Ile Val Gly Glu Thr Val Glu Lys Thr Glu Phe Ile
 35 40 45

Pro Leu Leu Asp Gly Asp Glu Lys Thr Gly Asn Ser Glu Ser Lys Lys
 50 55 60

Lys Pro Cys Leu Asp Thr Ser Gln Val Glu Gly Ile Pro Ser Ser Lys
 65 70 75 80

Pro Thr Leu Leu Ala Asn Gly Asp His Gly Met Glu Gly Asn Asn Thr
 85 90 95

Ala Gly Ser Pro Thr Asp Phe Leu Glu Glu Arg Val Asp Tyr Pro Asp
 100 105 110

Tyr Gln Ser Ser Gln Asn Trp Pro Glu Asp Ala Ser Phe Cys Phe Gln
 115 120 125

Pro Gln Gln Val Leu Asp Thr Asp Gln Ala Glu Pro Phe Asn Glu His
 130 135 140

Arg Asp Asp Gly Leu Ala Asp Leu Leu Phe Val Ser Ser Gly Pro Thr
 145 150 155 160

Asn Ala Ser Ala Phe Thr Glu Arg Asp Asn Pro Ser Glu Asp Ser Tyr
 165 170 175

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Gly Met Leu Pro Cys Asp Ser Phe Ala Ser Thr Ala Val Val Ser Gln
 180 185 190
 Glu Trp Ser Val Gly Ala Pro Asn Ser Pro Cys Ser Glu Ser Cys Val
 195 200 205
 Ser Pro Glu Val Thr Ile Glu Thr Leu Gln Pro Ala Thr Glu Leu Ser
 210 215 220
 Lys Ala Ala Glu Val Glu Ser Val Lys Glu Gln Leu Pro Ala Lys Ala
 225 230 235 240
 Leu Glu Thr Met Ala Glu Gln Thr Thr Asp Val Val His Ser Pro Ser
 245 250 255
 Thr Asp Thr Thr Pro Gly Pro Asp Thr Glu Ala Ala Leu Ala Lys Asp
 260 265 270
 Ile Glu Glu Ile Thr Lys Pro Asp Val Ile Leu Ala Asn Val Thr Gln
 275 280 285
 Pro Ser Thr Glu Ser Asp Met Phe Leu Ala Gln Asp Met Glu Leu Leu
 290 295 300
 Thr Gly Thr Glu Ala Ala His Ala Asn Asn Ile Ile Leu Pro Thr Glu
 305 310 315 320
 Pro Asp Glu Ser Ser Thr Lys Asp Val Ala Pro Pro Met Glu Glu Glu
 325 330 335
 Ile Val Pro Gly Asn Asp Thr Thr Ser Pro Lys Glu Thr Glu Thr Thr
 340 345 350
 Leu Pro Ile Lys Met Asp Leu Ala Pro Pro Glu Asp Val Leu Leu Thr
 355 360 365
 Lys Glu Thr Glu Leu Ala Pro Ala Lys Gly Met Val Ser Leu Ser Glu
 370 375 380
 Ile Glu Glu Ala Leu Ala Lys Asn Asp Glu Ser Ser Ala Glu Ile Pro
 385 390 395 400
 Val Ala Gln Glu Thr Val Val Ser Glu Thr Glu Val Val Leu Ala Ile
 405 410 415
 Glu Val Val Leu Pro Ser Asp Pro Ile Thr Thr Leu Thr Lys Asp Val
 420 425 430
 Thr Leu Pro Leu Glu Ala Glu Arg Pro Leu Val Thr Asp Met Thr Pro
 435 440 445
 Ser Leu Glu Thr Glu Met Thr Leu Gly Lys Glu Thr Ala Pro Pro Thr
 450 455 460
 Glu Thr Asn Leu Gly Met Ala Lys Asp Met Ser Pro Leu Pro Glu Ser
 465 470 475 480
 Glu Val Thr Leu Gly Lys Asp Val Val Ile Leu Pro Glu Thr Lys Val
 485 490 495
 Ala Glu Phe Asn Asn Val Thr Pro Leu Ser Glu Glu Glu Val Thr Ser
 500 505 510
 Val Lys Asp Met Ser Pro Ser Ala Glu Thr Glu Ala Pro Leu Ala Lys
 515 520 525
 Asn Ala Asp Leu His Ser Gly Thr Glu Leu Ile Val Asp Asn Ser Met
 530 535 540
 Ala Pro Ala Ser Asp Leu Ala Leu Pro Leu Glu Thr Lys Val Ala Thr
 545 550 555 560
 Val Pro Ile Lys Asp Lys Gly Thr Val Gln Thr Glu Glu Lys Pro Arg
 565 570 575

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Glu	Asp	Ser	Gln	Leu	Ala	Ser	Met	Gln	His	Lys	Gly	Gln	Ser	Thr	Val
			580					585					590		
Pro	Pro	Cys	Thr	Ala	Ser	Pro	Glu	Pro	Val	Lys	Ala	Ala	Glu	Gln	Met
		595					600					605			
Ser	Thr	Leu	Pro	Ile	Asp	Ala	Pro	Ser	Pro	Leu	Glu	Asn	Leu	Glu	Gln
	610					615					620				
Lys	Glu	Thr	Pro	Gly	Ser	Gln	Pro	Ser	Glu	Pro	Cys	Ser	Gly	Val	Ser
625					630					635					640
Arg	Gln	Glu	Glu	Ala	Lys	Ala	Ala	Val	Gly	Val	Thr	Gly	Asn	Asp	Ile
				645					650					655	
Thr	Thr	Pro	Pro	Asn	Lys	Glu	Pro	Pro	Ser	Pro	Glu	Lys	Lys	Ala	
			660					665					670		
Lys	Pro	Leu	Ala	Thr	Thr	Gln	Pro	Ala	Lys	Thr	Ser	Thr	Ser	Lys	Ala
		675					680					685			
Lys	Thr	Gln	Pro	Thr	Ser	Leu	Pro	Lys	Gln	Pro	Ala	Pro	Thr	Thr	Ser
	690					695					700				
Gly	Gly	Leu	Asn	Lys	Lys	Pro	Met	Ser	Leu	Ala	Ser	Gly	Ser	Val	Pro
705					710					715					720
Ala	Ala	Pro	His	Lys	Arg	Pro	Ala	Ala	Ala	Thr	Ala	Thr	Ala	Arg	Pro
				725					730					735	
Ser	Thr	Leu	Pro	Ala	Arg	Asp	Val	Lys	Pro	Lys	Pro	Ile	Thr	Glu	Ala
			740					745					750		
Lys	Val	Ala	Glu	Lys	Arg	Thr	Ser	Pro	Ser	Lys	Pro	Ser	Ser	Ala	Pro
		755					760					765			
Ala	Leu	Lys	Pro	Gly	Pro	Lys	Thr	Thr	Pro	Thr	Val	Ser	Lys	Ala	Thr
	770					775					780				
Ser	Pro	Ser	Thr	Leu	Val	Ser	Thr	Gly	Pro	Ser	Ser	Arg	Ser	Pro	Ala
					790					795					800
Thr	Thr	Leu	Pro	Lys	Arg	Pro	Thr	Ser	Ile	Lys	Thr	Glu	Gly	Lys	Pro
				805					810					815	
Ala	Asp	Val	Lys	Arg	Met	Thr	Ala	Lys	Ser	Ala	Ser	Ala	Asp	Leu	Ser
				820				825					830		
Arg	Ser	Lys	Thr	Thr	Ser	Ala	Ser	Ser	Val	Lys	Arg	Asn	Thr	Thr	Pro
		835					840					845			
Thr	Gly	Ala	Ala	Pro	Pro	Ala	Gly	Met	Thr	Ser	Thr	Arg	Val	Lys	Pro
	850					855					860				
Met	Ser	Ala	Pro	Ser	Arg	Ser	Ser	Gly	Ala	Leu	Ser	Val	Asp	Lys	Lys
	865				870					875					880
Pro	Thr	Ser	Thr	Lys	Pro	Ser	Ser	Ser	Ala	Pro	Arg	Val	Ser	Arg	Leu
				885					890					895	
Ala	Thr	Thr	Val	Ser	Ala	Pro	Asp	Leu	Lys	Ser	Val	Arg	Ser	Lys	Val
			900					905					910		
Gly	Ser	Thr	Glu	Asn	Ile	Lys	His	Gln	Pro	Gly	Gly	Gly	Arg	Ala	Lys
		915					920						925		
Val	Glu	Lys	Lys	Thr	Glu	Ala	Ala	Thr	Thr	Ala	Gly	Lys	Pro	Glu	Pro
	930					935					940				
Asn	Ala	Val	Thr	Lys	Ala	Ala	Gly	Ser	Ile	Ala	Ser	Ala	Gln	Lys	Pro
	945				950					955					960
Pro	Ala	Gly	Lys	Val	Gln	Ile	Val	Ser	Lys	Lys	Val	Ser	Tyr	Ser	His
				965					970					975	
Ile	Gln	Ser	Lys	Cys	Gly	Ser	Lys	Asp	Asn	Ile	Lys	His	Val	Pro	Gly

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	980		985		990														
Gly	Gly	Asn	Val	Gln	Ile	Gln	Asn	Lys	Lys	Val	Asp	Ile	Ser	Lys	Val				
	995						1000					1005							
Ser	Ser	Lys	Cys	Gly	Ser	Lys	Ala	Asn	Ile	Lys	His	Lys	Pro	Gly					
	1010					1015					1020								
Gly	Gly	Asp	Val	Lys	Ile	Glu	Ser	Gln	Lys	Leu	Asn	Phe	Lys	Glu					
	1025					1030					1035								
Lys	Ala	Gln	Ala	Lys	Val	Gly	Ser	Leu	Asp	Asn	Val	Gly	His	Leu					
	1040					1045					1050								
Pro	Ala	Gly	Gly	Ala	Val	Lys	Thr	Glu	Gly	Gly	Gly	Ser	Glu	Ala					
	1055					1060					1065								
Leu	Pro	Cys	Pro	Gly	Pro	Pro	Ala	Gly	Glu	Glu	Pro	Val	Ile	Pro					
	1070					1075					1080								
Glu	Ala	Ala	Pro	Asp	Ala	Gly	Ala	Pro	Thr	Ser	Ala	Ser	Gly	Leu					
	1085					1090					1095								
Ser	Gly	His	Thr	Thr	Leu	Ser	Gly	Gly	Gly	Asp	Gln	Arg	Glu	Pro					
	1100					1105					1110								
Gln	Thr	Leu	Asp	Ser	Gln	Ile	Gln	Glu	Thr	Ser	Ile								
	1115					1120					1125								

<210> SEQ ID NO 5
 <211> LENGTH: 383
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 5

Met	Ala	Glu	Pro	Arg	Gln	Glu	Phe	Glu	Val	Met	Glu	Asp	His	Ala	Gly
1				5					10					15	
Thr	Tyr	Gly	Leu	Gly	Asp	Arg	Lys	Asp	Gln	Gly	Gly	Tyr	Thr	Met	His
			20					25					30		
Gln	Asp	Gln	Glu	Gly	Asp	Thr	Asp	Ala	Gly	Leu	Lys	Ala	Glu	Glu	Ala
		35					40					45			
Gly	Ile	Gly	Asp	Thr	Pro	Ser	Leu	Glu	Asp	Glu	Ala	Ala	Gly	His	Val
	50					55					60				
Thr	Gln	Ala	Arg	Met	Val	Ser	Lys	Ser	Lys	Asp	Gly	Thr	Gly	Ser	Asp
65					70					75				80	
Asp	Lys	Lys	Ala	Lys	Gly	Ala	Asp	Gly	Lys	Thr	Lys	Ile	Ala	Thr	Pro
			85						90					95	
Arg	Gly	Ala	Ala	Pro	Pro	Gly	Gln	Lys	Gly	Gln	Ala	Asn	Ala	Thr	Arg
		100						105						110	
Ile	Pro	Ala	Lys	Thr	Pro	Pro	Ala	Pro	Lys	Thr	Pro	Pro	Ser	Ser	Gly
	115						120						125		
Glu	Pro	Pro	Lys	Ser	Gly	Asp	Arg	Ser	Gly	Tyr	Ser	Ser	Pro	Gly	Ser
	130					135					140				
Pro	Gly	Thr	Pro	Gly	Ser	Arg	Ser	Arg	Thr	Pro	Ser	Leu	Pro	Thr	Pro
145				150						155					160
Pro	Thr	Arg	Glu	Pro	Lys	Lys	Val	Ala	Val	Val	Arg	Thr	Pro	Pro	Lys
			165						170					175	
Ser	Pro	Ser	Ser	Ala	Lys	Ser	Arg	Leu	Gln	Thr	Ala	Pro	Val	Pro	Met
		180						185					190		
Pro	Asp	Leu	Lys	Asn	Val	Lys	Ser	Lys	Ile	Gly	Ser	Thr	Glu	Asn	Leu
	195					200						205			

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Pro Asp Leu Lys Asn Val Lys Ser Lys Ile Gly Ser Thr Glu Asn Leu
  195                               200                               205
Lys His Gln Pro Gly Gly Gly Lys Val Gln Ile Val Tyr Lys Pro Val
  210                               215                               220
Asp Leu Ser Lys Val Thr Ser Lys Cys Gly Ser Leu Gly Asn Ile His
  225                               230                               235                               240
His Lys Pro Gly Gly Gly Gln Val Glu Val Lys Ser Glu Lys Leu Asp
  245                               250                               255
Phe Lys Asp Arg Val Gln Ser Lys Ile Gly Ser Leu Asp Asn Ile Thr
  260                               265                               270
His Val Pro Gly Gly Gly Asn Lys Lys Ile Glu Thr His Lys Leu Thr
  275                               280                               285
Phe Arg Glu Asn Ala Lys Ala Lys Thr Asp His Gly Ala Glu Ile Val
  290                               295                               300
Tyr Lys Ser Pro Val Val Ser Gly Asp Thr Ser Pro Arg His Leu Ser
  305                               310                               315                               320
Asn Val Ser Ser Thr Gly Ser Ile Asp Met Val Asp Ser Pro Gln Leu
  325                               330                               335
Ala Thr Leu Ala Asp Glu Val Ser Ala Ser Leu Ala Lys Gln Gly Leu
  340                               345                               350

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<210> SEQ ID NO 7
<211> LENGTH: 1689
<212> TYPE: PRT
<213> ORGANISM: Mus musculus

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<400> SEQUENCE: 7

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Met Ala Gly Ala Ser Val Lys Val Ala Val Arg Val Arg Pro Phe Asn
  1                               5                               10                               15
Ser Arg Glu Met Ser Arg Asp Ser Lys Cys Ile Ile Gln Met Ser Gly
  20                               25                               30
Ser Thr Thr Thr Ile Val Asn Pro Lys Gln Pro Lys Glu Thr Pro Lys
  35                               40                               45
Ser Phe Ser Phe Asp Tyr Ser Tyr Trp Ser His Thr Ser Pro Glu Asp
  50                               55                               60
Ile Asn Tyr Ala Ser Gln Lys Gln Val Tyr Arg Asp Ile Gly Glu Glu
  65                               70                               75                               80
Met Leu Gln His Ala Phe Glu Gly Tyr Asn Val Cys Ile Phe Ala Tyr
  85                               90                               95
Gly Gln Thr Gly Ala Gly Lys Ser Tyr Thr Met Met Gly Lys Gln Glu
  100                              105                              110
Lys Asp Gln Gln Gly Ile Ile Pro Gln Leu Cys Glu Asp Leu Phe Ser
  115                              120                              125
Arg Ile Asn Asp Thr Thr Asn Asp Asn Met Ser Tyr Ser Val Glu Val
  130                              135                              140
Ser Tyr Met Glu Ile Tyr Cys Glu Arg Val Arg Asp Leu Leu Asn Pro
  145                              150                              155                              160
Lys Asn Lys Gly Asn Leu Arg Val Arg Glu His Pro Leu Leu Gly Pro
  165                              170                              175
Tyr Val Glu Asp Leu Ser Lys Leu Ala Val Thr Ser Tyr Asn Asp Ile
  180                              185                              190
Gln Asp Leu Met Asp Ser Gly Asn Lys Ala Arg Thr Val Ala Ala Thr

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195				200				205							
Asn	Met	Asn	Glu	Thr	Ser	Ser	Arg	Ser	His	Ala	Val	Phe	Asn	Ile	Ile
210						215					220				
Phe	Thr	Gln	Lys	Arg	His	Asp	Ala	Glu	Thr	Asn	Ile	Thr	Thr	Glu	Lys
225					230					235					240
Val	Ser	Lys	Ile	Ser	Leu	Val	Asp	Leu	Ala	Gly	Ser	Glu	Arg	Ala	Asp
				245					250					255	
Ser	Thr	Gly	Ala	Lys	Gly	Thr	Arg	Leu	Lys	Glu	Gly	Ala	Asn	Ile	Asn
			260					265					270		
Lys	Ser	Leu	Thr	Thr	Leu	Gly	Lys	Val	Ile	Ser	Ala	Leu	Ala	Glu	Met
		275					280						285		
Asp	Ser	Gly	Pro	Asn	Lys	Asn	Lys	Lys	Lys	Lys	Lys	Thr	Asp	Phe	Ile
290						295					300				
Pro	Tyr	Arg	Asp	Ser	Val	Leu	Thr	Trp	Leu	Leu	Arg	Glu	Asn	Leu	Gly
305					310					315					320
Gly	Asn	Ser	Arg	Thr	Ala	Met	Val	Ala	Ala	Leu	Ser	Pro	Ala	Asp	Ile
				325					330					335	
Asn	Tyr	Asp	Glu	Thr	Leu	Ser	Thr	Leu	Arg	Tyr	Ala	Asp	Arg	Ala	Lys
			340						345					350	
Gln	Ile	Arg	Cys	Asn	Ala	Ile	Ile	Asn	Glu	Asp	Pro	Asn	Asn	Lys	Leu
		355					360							365	
Ile	Arg	Glu	Leu	Lys	Asp	Glu	Val	Thr	Arg	Leu	Arg	Asp	Leu	Leu	Tyr
	370					375					380				
Ala	Gln	Gly	Leu	Gly	Asp	Ile	Thr	Asp	Met	Thr	Asn	Ala	Leu	Val	Gly
385					390					395					400
Met	Ser	Pro	Ser	Ser	Ser	Leu	Ser	Ala	Leu	Ser	Ser	Arg	Ala	Ala	Ser
				405					410					415	
Val	Ser	Ser	Leu	His	Glu	Arg	Ile	Leu	Phe	Ala	Pro	Gly	Ser	Glu	Glu
			420						425					430	
Ala	Ile	Glu	Arg	Leu	Lys	Glu	Thr	Glu	Lys	Ile	Ile	Ala	Glu	Leu	Asn
		435					440						445		
Glu	Thr	Trp	Glu	Glu	Lys	Leu	Arg	Arg	Thr	Glu	Ala	Ile	Arg	Met	Glu
	450				455						460				
Arg	Glu	Ala	Leu	Leu	Ala	Glu	Met	Gly	Val	Ala	Met	Arg	Glu	Asp	Gly
465					470					475					480
Gly	Thr	Leu	Gly	Val	Phe	Ser	Pro	Lys	Lys	Thr	Pro	His	Leu	Val	Asn
				485					490					495	
Leu	Asn	Glu	Asp	Pro	Leu	Met	Ser	Glu	Cys	Leu	Leu	Tyr	Tyr	Ile	Lys
			500						505				510		
Asp	Gly	Val	Thr	Arg	Val	Gly	Arg	Glu	Asp	Ala	Glu	Arg	Arg	Gln	Asp
		515					520						525		
Ile	Val	Leu	Ser	Gly	His	Phe	Ile	Lys	Glu	Glu	His	Cys	Ile	Phe	Arg
		530				535					540				
Ser	Asp	Ser	Arg	Gly	Gly	Gly	Glu	Ala	Val	Val	Thr	Leu	Glu	Pro	Cys
545					550					555					560
Glu	Gly	Ala	Asp	Thr	Tyr	Val	Asn	Gly	Lys	Lys	Val	Thr	Glu	Pro	Ser
				565					570					575	
Ile	Leu	Arg	Ser	Gly	Asn	Arg	Ile	Ile	Met	Gly	Lys	Ser	His	Val	Phe
			580						585				590		
Arg	Phe	Asn	His	Pro	Glu	Gln	Ala	Arg	Gln	Glu	Arg	Glu	Arg	Thr	Pro
		595					600							605	

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Cys Ala Glu Thr Pro Ala Glu Pro Val Asp Trp Ala Phe Ala Gln Arg
 610 615 620
 Glu Leu Leu Glu Lys Gln Gly Ile Asp Met Lys Gln Glu Met Glu Gln
 625 630 635 640
 Arg Leu Gln Glu Leu Glu Asp Gln Tyr Arg Arg Glu Arg Glu Glu Ala
 645 650 655
 Thr Tyr Leu Leu Glu Gln Gln Arg Leu Asp Tyr Glu Ser Lys Leu Glu
 660 665 670
 Ala Leu Gln Lys Gln Met Asp Ser Arg Tyr Tyr Pro Glu Val Asn Glu
 675 680 685
 Glu Glu Glu Glu Pro Glu Asp Glu Val Gln Trp Thr Glu Arg Glu Cys
 690 695 700
 Glu Leu Ala Leu Trp Ala Phe Arg Lys Trp Lys Trp Tyr Gln Phe Thr
 705 710 715 720
 Ser Leu Arg Asp Leu Leu Trp Gly Asn Ala Ile Phe Leu Lys Glu Ala
 725 730 735
 Asn Ala Ile Ser Val Glu Leu Lys Lys Lys Val Gln Phe Gln Phe Val
 740 745 750
 Leu Leu Thr Asp Thr Leu Tyr Ser Pro Leu Pro Pro Asp Leu Leu Pro
 755 760 765
 Pro Glu Ala Ala Lys Asp Arg Glu Thr Arg Pro Phe Pro Arg Thr Ile
 770 775 780
 Val Ala Val Glu Val Gln Asp Gln Lys Asn Gly Ala Thr His Tyr Trp
 785 790 795 800
 Thr Leu Glu Lys Leu Arg Gln Arg Leu Asp Leu Met Arg Glu Met Tyr
 805 810 815
 Asp Arg Ala Ala Glu Val Pro Ser Ser Val Val Glu Asp Cys Asp Asn
 820 825 830
 Val Val Thr Gly Gly Asp Pro Phe Tyr Asp Arg Phe Pro Trp Phe Arg
 835 840 845
 Leu Val Gly Arg Ala Phe Val Tyr Leu Ser Asn Leu Leu Tyr Pro Val
 850 855 860
 Pro Leu Val His Arg Val Ala Ile Val Ser Glu Lys Gly Glu Val Lys
 865 870 875 880
 Gly Phe Leu Arg Val Ala Val Gln Ala Ile Ser Ala Asp Glu Glu Ala
 885 890 895
 Pro Asp Tyr Gly Ser Gly Val Arg Gln Ser Gly Thr Ala Lys Ile Ser
 900 905 910
 Phe Asp Asp Gln His Phe Glu Lys Phe Gln Ser Glu Ser Cys Pro Val
 915 920 925
 Val Gly Met Ser Arg Ser Gly Thr Ser Gln Glu Glu Leu Arg Ile Val
 930 935 940
 Glu Gly Gln Gly Gln Gly Ala Asp Ala Gly Pro Ser Ala Asp Glu Val
 945 950 955 960
 Asn Asn Asn Thr Cys Ser Ala Val Pro Pro Glu Gly Leu Met Asp Ser
 965 970 975
 Pro Glu Lys Ala Ala Leu Asp Gly Pro Leu Asp Thr Ala Leu Asp His
 980 985 990
 Leu Arg Leu Gly Ser Thr Phe Thr Phe Arg Val Thr Val Leu Gln Ala
 995 1000 1005

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Ser	Ser	Ile	Ser	Ala	Glu	Tyr	Ala	Asp	Ile	Phe	Cys	Gln	Phe	Asn
1010						1015					1020			
Phe	Ile	His	Arg	His	Asp	Glu	Ala	Phe	Ser	Thr	Glu	Pro	Leu	Lys
1025						1030					1035			
Asn	Thr	Gly	Arg	Gly	Pro	Pro	Leu	Gly	Phe	Tyr	His	Val	Gln	Asn
1040						1045					1050			
Ile	Ala	Val	Glu	Val	Thr	Lys	Ser	Phe	Ile	Glu	Tyr	Ile	Lys	Ser
1055						1060					1065			
Gln	Pro	Ile	Val	Phe	Glu	Val	Phe	Gly	His	Tyr	Gln	Gln	His	Pro
1070						1075					1080			
Phe	Pro	Pro	Leu	Cys	Lys	Asp	Val	Leu	Ser	Pro	Leu	Arg	Pro	Ser
1085						1090					1095			
Arg	Arg	His	Phe	Pro	Arg	Val	Met	Pro	Leu	Ser	Lys	Pro	Val	Pro
1100						1105					1110			
Ala	Thr	Lys	Leu	Ser	Thr	Met	Thr	Arg	Pro	Ser	Pro	Gly	Pro	Cys
1115						1120					1125			
His	Cys	Lys	Tyr	Asp	Leu	Leu	Val	Tyr	Phe	Glu	Ile	Cys	Glu	Leu
1130						1135					1140			
Glu	Ala	Asn	Gly	Asp	Tyr	Ile	Pro	Ala	Val	Val	Asp	His	Arg	Gly
1145						1150					1155			
Gly	Met	Pro	Cys	Met	Gly	Thr	Phe	Leu	Leu	His	Gln	Gly	Ile	Gln
1160						1165					1170			
Arg	Arg	Ile	Thr	Val	Thr	Leu	Leu	His	Glu	Thr	Gly	Ser	His	Ile
1175						1180					1185			
Arg	Trp	Lys	Glu	Val	Arg	Glu	Leu	Val	Val	Gly	Arg	Ile	Arg	Asn
1190						1195					1200			
Thr	Pro	Glu	Thr	Asp	Glu	Ala	Leu	Ile	Asp	Pro	Asn	Ile	Leu	Ser
1205						1210					1215			
Leu	Asn	Ile	Leu	Ser	Ser	Gly	Tyr	Val	His	Pro	Ala	Gln	Asp	Asp
1220						1225					1230			
Arg	Thr	Phe	Tyr	Gln	Phe	Glu	Ala	Ala	Trp	Asp	Ser	Ser	Met	His
1235						1240					1245			
Asn	Ser	Leu	Leu	Leu	Asn	Arg	Val	Thr	Pro	Tyr	Arg	Glu	Lys	Ile
1250						1255					1260			
Tyr	Met	Thr	Leu	Ser	Ala	Tyr	Ile	Glu	Met	Glu	Asn	Cys	Thr	Gln
1265						1270					1275			
Pro	Ala	Val	Ile	Thr	Lys	Asp	Phe	Cys	Met	Val	Phe	Tyr	Ser	Arg
1280						1285					1290			
Asp	Ala	Lys	Leu	Pro	Ala	Ser	Arg	Ser	Ile	Arg	Asn	Leu	Phe	Gly
1295						1300					1305			
Ser	Gly	Ser	Leu	Arg	Ala	Thr	Glu	Gly	Asn	Arg	Val	Thr	Gly	Val
1310						1315					1320			
Tyr	Glu	Leu	Ser	Leu	Cys	His	Val	Ala	Asp	Ala	Gly	Ser	Pro	Gly
1325						1330					1335			
Met	Gln	Arg	Arg	Arg	Arg	Arg	Val	Leu	Asp	Thr	Ser	Val	Ala	Tyr
1340						1345					1350			
Val	Arg	Gly	Glu	Glu	Asn	Leu	Ala	Gly	Trp	Arg	Pro	Arg	Ser	Asp
1355						1360					1365			
Ser	Leu	Ile	Leu	Asp	His	Gln	Trp	Glu	Leu	Glu	Lys	Leu	Ser	Leu
1370						1375					1380			
Leu	Gln	Glu	Val	Glu	Lys	Thr	Arg	His	Tyr	Leu	Leu	Leu	Arg	Glu

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Pro	Arg	Ser	Leu	His	Pro	His	Val	Pro	Gly	Val	Thr	Asn	Leu	Gln	Val
		35					40					45			
Met	Arg	Ala	Met	Ala	Ser	Leu	Arg	Ala	Arg	Gly	Leu	Val	Arg	Glu	Thr
		50				55					60				
Phe	Ala	Trp	Cys	His	Phe	Tyr	Trp	Tyr	Leu	Thr	Asn	Glu	Gly	Ile	Ala
				70						75					80
His	Leu	Arg	Gln	Tyr	Leu	His	Leu	Pro	Pro	Glu	Ile	Val	Pro	Ala	Ser
				85					90						95
Leu	Gln	Arg	Val	Arg	Arg	Pro	Val	Ala	Met	Val	Met	Pro	Ala	Arg	Arg
			100					105						110	
Thr	Pro	His	Val	Gln	Ala	Val	Gln	Gly	Pro	Leu	Gly	Ser	Pro	Pro	Lys
							120						125		
Arg	Gly	Pro	Leu	Pro	Thr	Glu	Glu	Gln	Arg	Val	Tyr	Arg	Arg	Lys	Glu
		130					135					140			
Leu	Glu	Glu	Val	Ser	Pro	Glu	Thr	Pro	Val	Val	Pro	Ala	Thr	Thr	Gln
					150						155				160
Arg	Thr	Leu	Ala	Arg	Pro	Gly	Pro	Glu	Pro	Ala	Pro	Ala	Thr	Asp	Glu
				165					170						175
Arg	Asp	Arg	Val	Gln	Lys	Lys	Thr	Phe	Thr	Lys	Trp	Val	Asn	Lys	His
			180					185						190	
Leu	Ile	Lys	Ala	Gln	Arg	His	Ile	Ser	Asp	Leu	Tyr	Glu	Asp	Leu	Arg
			195				200						205		
Asp	Gly	His	Asn	Leu	Ile	Ser	Leu	Leu	Glu	Val	Leu	Ser	Gly	Asp	Ser
						215						220			
Leu	Pro	Arg	Glu	Lys	Gly	Arg	Met	Arg	Phe	His	Lys	Leu	Gln	Asn	Val
						230					235				240
Gln	Ile	Ala	Leu	Asp	Tyr	Leu	Arg	His	Arg	Gln	Val	Lys	Leu	Val	Asn
				245					250						255
Ile	Arg	Asn	Asp	Asp	Ile	Ala	Asp	Gly	Asn	Pro	Lys	Leu	Thr	Leu	Gly
			260					265						270	
Leu	Ile	Trp	Thr	Ile	Ile	Leu	His	Phe	Gln	Ile	Ser	Asp	Ile	Gln	Val
			275				280						285		
Ser	Gly	Gln	Ser	Glu	Asp	Met	Thr	Ala	Lys	Glu	Lys	Leu	Leu	Leu	Trp
						295						300			
Ser	Gln	Arg	Met	Val	Glu	Gly	Tyr	Gln	Gly	Leu	Arg	Cys	Asp	Asn	Phe
						310					315				320
Thr	Ser	Ser	Trp	Arg	Asp	Gly	Arg	Leu	Phe	Asn	Ala	Ile	Ile	His	Arg
				325						330					335
His	Lys	Pro	Leu	Leu	Ile	Asp	Met	Asn	Lys	Val	Tyr	Arg	Gln	Thr	Asn
			340					345						350	
Leu	Glu	Asn	Leu	Asp	Gln	Ala	Phe	Ser	Val	Ala	Glu	Arg	Asp	Leu	Gly
			355				360						365		
Val	Thr	Arg	Leu	Leu	Asp	Pro	Glu	Asp	Val	Asp	Val	Pro	Gln	Pro	Asp
						375						380			
Glu	Lys	Ser	Ile	Ile	Thr	Tyr	Val	Ser	Ser	Leu	Tyr	Asp	Ala	Met	Pro
						390					395				400
Arg	Val	Pro	Asp	Val	Gln	Asp	Gly	Val	Arg	Ala	Asn	Glu	Leu	Gln	Leu
				405						410					415
Arg	Trp	Gln	Glu	Tyr	Arg	Glu	Leu	Val	Leu	Leu	Leu	Leu	Gln	Trp	Met
				420					425					430	
Arg	His	His	Thr	Ala	Ala	Phe	Glu	Glu	Arg	Arg	Phe	Pro	Ser	Ser	Phe

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435				440				445							
Glu	Glu	Ile	Glu	Ile	Leu	Trp	Ser	Gln	Phe	Leu	Lys	Phe	Lys	Glu	Met
450						455					460				
Glu	Leu	Pro	Ala	Lys	Glu	Ala	Asp	Lys	Asn	Arg	Ser	Lys	Gly	Ile	Tyr
465					470					475					480
Gln	Ser	Leu	Glu	Gly	Ala	Val	Gln	Ala	Gly	Gln	Leu	Lys	Val	Pro	Pro
				485					490					495	
Gly	Tyr	His	Pro	Leu	Asp	Val	Glu	Lys	Glu	Trp	Gly	Lys	Leu	His	Val
			500					505					510		
Ala	Ile	Leu	Glu	Arg	Glu	Lys	Gln	Leu	Arg	Ser	Glu	Phe	Glu	Arg	Leu
		515					520					525			
Glu	Cys	Leu	Gln	Arg	Ile	Val	Thr	Lys	Leu	Gln	Met	Glu	Ala	Gly	Leu
530						535					540				
Cys	Glu	Glu	Gln	Leu	Asn	Gln	Ala	Asp	Ala	Leu	Leu	Gln	Ser	Asp	Val
545					550					555					560
Arg	Leu	Leu	Ala	Ala	Gly	Lys	Val	Pro	Gln	Arg	Ala	Gly	Glu	Val	Glu
				565					570					575	
Arg	Asp	Leu	Asp	Lys	Ala	Asp	Ser	Met	Ile	Arg	Leu	Leu	Phe	Asn	Asp
			580					585						590	
Val	Gln	Thr	Leu	Lys	Asp	Gly	Arg	His	Pro	Gln	Gly	Glu	Gln	Met	Tyr
			595				600					605			
Arg	Arg	Val	Tyr	Arg	Leu	His	Glu	Arg	Leu	Val	Ala	Ile	Arg	Thr	Glu
	610					615					620				
Tyr	Asn	Leu	Arg	Leu	Lys	Ala	Gly	Val	Ala	Ala	Pro	Ala	Thr	Gln	Val
625					630					635					640
Ala	Gln	Val	Thr	Leu	Gln	Ser	Val	Gln	Arg	Arg	Pro	Glu	Leu	Glu	Asp
				645					650					655	
Ser	Thr	Leu	Arg	Tyr	Leu	Gln	Asp	Leu	Leu	Ala	Trp	Val	Glu	Glu	Asn
			660				665						670		
Gln	His	Arg	Val	Asp	Gly	Ala	Glu	Trp	Gly	Val	Asp	Leu	Pro	Ser	Val
		675					680					685			
Glu	Ala	Gln	Leu	Gly	Ser	His	Arg	Gly	Leu	His	Gln	Ser	Ile	Glu	Glu
690						695					700				
Phe	Arg	Ala	Lys	Ile	Glu	Arg	Ala	Arg	Ser	Asp	Glu	Gly	Gln	Leu	Ser
705					710					715					720
Pro	Ala	Thr	Arg	Gly	Ala	Tyr	Arg	Asp	Cys	Leu	Gly	Arg	Leu	Asp	Leu
				725					730					735	
Gln	Tyr	Ala	Lys	Leu	Leu	Asn	Ser	Ser	Lys	Ala	Arg	Leu	Arg	Ser	Leu
			740					745					750		
Glu	Ser	Leu	His	Ser	Phe	Val	Ala	Ala	Ala	Thr	Lys	Glu	Leu	Met	Trp
		755					760					765			
Leu	Asn	Glu	Lys	Glu	Glu	Glu	Glu	Val	Gly	Phe	Asp	Trp	Ser	Asp	Arg
770						775					780				
Asn	Thr	Asn	Met	Thr	Ala	Lys	Lys	Glu	Ser	Tyr	Ser	Ala	Leu	Met	Arg
785					790					795					800
Glu	Leu	Glu	Leu	Lys	Glu	Lys	Lys	Ile	Lys	Glu	Leu	Gln	Asn	Ala	Gly
				805					810					815	
Asp	Arg	Leu	Leu	Arg	Glu	Asp	His	Pro	Ala	Arg	Pro	Thr	Val	Glu	Ser
			820					825					830		
Phe	Gln	Ala	Ala	Leu	Gln	Thr	Gln	Trp	Ser	Trp	Met	Leu	Gln	Leu	Cys
		835					840					845			

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Cys Cys Ile Glu Ala His Leu Lys Glu Asn Ala Ala Tyr Phe Gln Phe
 850 855 860

Phe Ser Asp Val Arg Glu Ala Glu Gly Gln Leu Gln Lys Leu Gln Glu
 865 870 875 880

Ala Leu Arg Arg Lys Tyr Ser Cys Asp Arg Ser Ala Thr Val Thr Arg
 885 890 895

Leu Glu Asp Leu Leu Gln Asp Ala Gln Asp Glu Lys Glu Gln Leu Asn
 900 905 910

Glu Tyr Lys Gly His Leu Ser Gly Leu Ala Lys Arg Ala Lys Ala Val
 915 920 925

Val Gln Leu Lys Pro Arg His Pro Ala His Pro Met Arg Gly Arg Leu
 930 935 940

Pro Leu Leu Ala Val Cys Asp Tyr Lys Gln Val Glu Val Thr Val His
 945 950 955 960

Lys Gly Asp Glu Cys Gln Leu Val Gly Pro Ala Gln Pro Ser His Trp
 965 970 975

Lys Val Leu Ser Ser Ser Gly Ser Glu Ala Ala Val Pro Ser Val Cys
 980 985 990

Phe Leu Val Pro Pro Pro Asn Gln Glu Ala Gln Glu Ala Val Thr Arg
 995 1000 1005

Leu Glu Ala Gln His Gln Ala Leu Val Thr Leu Trp His Gln Leu
 1010 1015 1020

His Val Asp Met Lys Ser Leu Leu Ala Trp Gln Ser Leu Arg Arg
 1025 1030 1035

Asp Val Gln Leu Ile Arg Ser Trp Ser Leu Ala Thr Phe Arg Thr
 1040 1045 1050

Leu Lys Pro Glu Glu Gln Arg Gln Ala Leu His Ser Leu Glu Leu
 1055 1060 1065

His Tyr Gln Ala Phe Leu Arg Asp Ser Gln Asp Ala Gly Gly Phe
 1070 1075 1080

Gly Pro Glu Asp Arg Leu Met Ala Glu Arg Glu Tyr Gly Ser Cys
 1085 1090 1095

Ser His His Tyr Gln Gln Leu Leu Gln Ser Leu Glu Gln Gly Ala
 1100 1105 1110

Gln Glu Glu Ser Arg Cys Gln Arg Cys Ile Ser Glu Leu Lys Asp
 1115 1120 1125

Ile Arg Leu Gln Leu Glu Ala Cys Glu Thr Arg Thr Val His Arg
 1130 1135 1140

Leu Arg Leu Pro Leu Asp Lys Glu Pro Ala Arg Glu Cys Ala Gln
 1145 1150 1155

Arg Ile Ala Glu Gln Gln Lys Ala Gln Ala Glu Val Glu Gly Leu
 1160 1165 1170

Gly Lys Gly Val Ala Arg Leu Ser Ala Glu Ala Glu Lys Val Leu
 1175 1180 1185

Ala Leu Pro Glu Pro Ser Pro Ala Ala Pro Thr Leu Arg Ser Glu
 1190 1195 1200

Leu Glu Leu Thr Leu Gly Lys Leu Glu Gln Val Arg Ser Leu Ser
 1205 1210 1215

Ala Ile Tyr Leu Glu Lys Leu Lys Thr Ile Ser Leu Val Ile Arg
 1220 1225 1230

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Gly	Thr	Gln	Gly	Ala	Glu	Glu	Val	Leu	Arg	Ala	His	Glu	Glu	Gln
1235						1240					1245			
Leu	Lys	Glu	Ala	Gln	Ala	Val	Pro	Ala	Thr	Leu	Pro	Glu	Leu	Glu
1250						1255					1260			
Ala	Thr	Lys	Ala	Ser	Leu	Lys	Lys	Leu	Arg	Ala	Gln	Ala	Glu	Ala
1265						1270					1275			
Gln	Gln	Pro	Thr	Phe	Asp	Ala	Leu	Arg	Asp	Glu	Leu	Arg	Gly	Ala
1280						1285					1290			
Gln	Glu	Val	Gly	Glu	Arg	Leu	Gln	Gln	Arg	His	Gly	Glu	Arg	Asp
1295						1300					1305			
Val	Glu	Val	Glu	Arg	Trp	Arg	Glu	Arg	Val	Ala	Gln	Leu	Leu	Glu
1310						1315					1320			
Arg	Trp	Gln	Ala	Val	Leu	Ala	Gln	Thr	Asp	Val	Arg	Gln	Arg	Glu
1325						1330					1335			
Leu	Glu	Gln	Leu	Gly	Arg	Gln	Leu	Arg	Tyr	Tyr	Arg	Glu	Ser	Ala
1340						1345					1350			
Asp	Pro	Leu	Gly	Ala	Trp	Leu	Gln	Asp	Ala	Arg	Arg	Arg	Gln	Glu
1355						1360					1365			
Gln	Ile	Gln	Ala	Met	Pro	Leu	Ala	Asp	Ser	Gln	Ala	Val	Arg	Glu
1370						1375					1380			
Gln	Leu	Arg	Gln	Glu	Gln	Ala	Leu	Leu	Glu	Glu	Ile	Glu	Arg	His
1385						1390					1395			
Gly	Glu	Lys	Val	Glu	Glu	Cys	Gln	Arg	Phe	Ala	Lys	Gln	Tyr	Ile
1400						1405					1410			
Asn	Ala	Ile	Lys	Asp	Tyr	Glu	Leu	Gln	Leu	Val	Thr	Tyr	Lys	Ala
1415						1420					1425			
Gln	Leu	Glu	Pro	Val	Ala	Ser	Pro	Ala	Lys	Lys	Pro	Lys	Val	Gln
1430						1435					1440			
Ser	Gly	Ser	Glu	Ser	Val	Ile	Gln	Glu	Tyr	Val	Asp	Leu	Arg	Thr
1445						1450					1455			
His	Tyr	Ser	Glu	Leu	Thr	Thr	Leu	Thr	Ser	Gln	Tyr	Ile	Lys	Phe
1460						1465					1470			
Ile	Ser	Glu	Thr	Leu	Arg	Arg	Met	Glu	Glu	Glu	Glu	Arg	Leu	Ala
1475						1480					1485			
Glu	Gln	Gln	Arg	Ala	Glu	Glu	Arg	Glu	Arg	Leu	Ala	Glu	Val	Glu
1490						1495					1500			
Ala	Ala	Leu	Glu	Lys	Gln	Arg	Gln	Leu	Ala	Glu	Ala	His	Ala	Gln
1505						1510					1515			
Ala	Lys	Ala	Gln	Ala	Glu	Arg	Glu	Ala	Lys	Glu	Leu	Gln	Gln	Arg
1520						1525					1530			
Met	Gln	Glu	Glu	Val	Val	Arg	Arg	Glu	Glu	Ala	Ala	Val	Asp	Ala
1535						1540					1545			
Gln	Gln	Gln	Lys	Arg	Ser	Ile	Gln	Glu	Glu	Leu	Gln	Gln	Leu	Arg
1550						1555					1560			
Gln	Ser	Ser	Glu	Ala	Glu	Ile	Gln	Ala	Lys	Ala	Arg	Gln	Ala	Glu
1565						1570					1575			
Ala	Ala	Glu	Arg	Ser	Arg	Leu	Arg	Ile	Glu	Glu	Glu	Ile	Arg	Val
1580						1585					1590			
Val	Arg	Leu	Gln	Leu	Glu	Ala	Thr	Glu	Arg	Gln	Arg	Gly	Gly	Ala
1595						1600					1605			
Glu	Gly	Glu	Leu	Gln	Ala	Leu	Arg	Ala	Arg	Ala	Glu	Glu	Ala	Glu

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1610	1615	1620
Ala Gln Lys Arg Gln Ala Gln	Glu Glu Ala Glu Arg	Leu Arg Arg
1625	1630	1635
Gln Val Gln Asp Glu Ser Gln	Arg Lys Arg Gln Ala	Glu Val Glu
1640	1645	1650
Leu Ala Ser Arg Val Lys Ala	Glu Ala Glu Ala Ala	Arg Glu Lys
1655	1660	1665
Gln Arg Ala Leu Gln Ala Leu	Glu Glu Leu Arg Leu	Gln Ala Glu
1670	1675	1680
Glu Ala Glu Arg Arg Leu Arg	Gln Ala Glu Val Glu	Arg Ala Arg
1685	1690	1695
Gln Val Gln Val Ala Leu Glu	Thr Ala Gln Arg Ser	Ala Glu Ala
1700	1705	1710
Glu Leu Gln Ser Lys Arg Ala	Ser Phe Ala Glu Lys	Thr Ala Gln
1715	1720	1725
Leu Glu Arg Ser Leu Gln Glu	Glu His Val Ala Val	Ala Gln Leu
1730	1735	1740
Arg Glu Glu Ala Glu Arg Arg	Ala Gln Gln Gln Ala	Glu Ala Glu
1745	1750	1755
Arg Ala Arg Glu Glu Ala Glu	Arg Glu Leu Glu Arg	Trp Gln Leu
1760	1765	1770
Lys Ala Asn Glu Ala Leu Arg	Leu Arg Leu Gln Ala	Glu Glu Val
1775	1780	1785
Ala Gln Gln Lys Ser Leu Ala	Gln Ala Glu Ala Glu	Lys Gln Lys
1790	1795	1800
Glu Glu Ala Glu Arg Glu Ala	Arg Arg Arg Gly Lys	Ala Glu Glu
1805	1810	1815
Gln Ala Val Arg Gln Arg Glu	Leu Ala Glu Gln Glu	Leu Glu Lys
1820	1825	1830
Gln Arg Gln Leu Ala Glu Gly	Thr Ala Gln Gln Arg	Leu Ala Ala
1835	1840	1845
Glu Gln Glu Leu Ile Arg Leu	Arg Ala Glu Thr Glu	Gln Gly Glu
1850	1855	1860
Gln Gln Arg Gln Leu Leu Glu	Glu Glu Leu Ala Arg	Leu Gln Arg
1865	1870	1875
Glu Ala Ala Ala Ala Thr Gln	Lys Arg Gln Glu Leu	Glu Ala Glu
1880	1885	1890
Leu Ala Lys Val Arg Ala Glu	Met Glu Val Leu Leu	Ala Ser Lys
1895	1900	1905
Ala Arg Ala Glu Glu Glu Ser	Arg Ser Thr Ser Glu	Lys Ser Lys
1910	1915	1920
Gln Arg Leu Glu Ala Glu Ala	Gly Arg Phe Arg Glu	Leu Ala Glu
1925	1930	1935
Glu Ala Ala Arg Leu Arg Ala	Leu Ala Glu Glu Ala	Lys Arg Gln
1940	1945	1950
Arg Gln Leu Ala Glu Glu Asp	Ala Ala Arg Gln Arg	Ala Glu Ala
1955	1960	1965
Glu Arg Val Leu Ala Glu Lys	Leu Ala Ala Ile Gly	Glu Ala Thr
1970	1975	1980
Arg Leu Lys Thr Glu Ala Glu	Ile Ala Leu Lys Glu	Lys Glu Ala
1985	1990	1995

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Glu 2000	Asn	Glu	Arg	Leu	Arg	Arg	Leu	Ala	Glu	Asp	Glu	Ala	Phe	Gln
						2005					2010			
Arg	Arg	Arg	Leu	Glu	Glu	Gln	Ala	Ala	Gln	His	Lys	Ala	Asp	Ile
2015						2020					2025			
Glu	Glu	Arg	Leu	Ala	Gln	Leu	Arg	Lys	Ala	Ser	Asp	Ser	Glu	Leu
2030						2035					2040			
Glu	Arg	Gln	Lys	Gly	Leu	Val	Glu	Asp	Thr	Leu	Arg	Gln	Arg	Arg
2045						2050					2055			
Gln	Val	Glu	Glu	Glu	Ile	Leu	Ala	Leu	Lys	Ala	Ser	Phe	Glu	Lys
2060						2065					2070			
Ala	Ala	Ala	Gly	Lys	Ala	Glu	Leu	Glu	Leu	Glu	Leu	Gly	Arg	Ile
2075						2080					2085			
Arg	Ser	Asn	Ala	Glu	Asp	Thr	Leu	Arg	Ser	Lys	Glu	Gln	Ala	Glu
2090						2095					2100			
Leu	Glu	Ala	Ala	Arg	Gln	Arg	Gln	Leu	Ala	Ala	Glu	Glu	Glu	Arg
2105						2110					2115			
Arg	Arg	Arg	Glu	Ala	Glu	Glu	Arg	Val	Gln	Lys	Ser	Leu	Ala	Ala
2120						2125					2130			
Glu	Glu	Glu	Ala	Ala	Arg	Gln	Arg	Lys	Ala	Ala	Leu	Glu	Glu	Val
2135						2140					2145			
Glu	Arg	Leu	Lys	Ala	Lys	Val	Glu	Glu	Ala	Arg	Arg	Leu	Arg	Glu
2150						2155					2160			
Arg	Ala	Glu	Gln	Glu	Ser	Ala	Arg	Gln	Leu	Gln	Leu	Ala	Gln	Glu
2165						2170					2175			
Ala	Ala	Gln	Lys	Arg	Leu	Gln	Ala	Glu	Glu	Lys	Ala	His	Ala	Phe
2180						2185					2190			
Ala	Val	Gln	Gln	Lys	Glu	Gln	Glu	Leu	Gln	Gln	Thr	Leu	Gln	Gln
2195						2200					2205			
Glu	Gln	Ser	Val	Leu	Asp	Gln	Leu	Arg	Gly	Glu	Ala	Glu	Ala	Ala
2210						2215					2220			
Arg	Arg	Ala	Ala	Glu	Glu	Ala	Glu	Glu	Ala	Arg	Val	Gln	Ala	Glu
2225						2230					2235			
Arg	Glu	Ala	Ala	Gln	Ser	Arg	Arg	Gln	Val	Glu	Glu	Ala	Glu	Arg
2240						2245					2250			
Leu	Lys	Gln	Ser	Ala	Glu	Glu	Gln	Ala	Gln	Ala	Arg	Ala	Gln	Ala
2255						2260					2265			
Gln	Ala	Ala	Ala	Glu	Lys	Leu	Arg	Lys	Glu	Ala	Glu	Gln	Glu	Ala
2270						2275					2280			
Ala	Arg	Arg	Ala	Gln	Ala	Glu	Gln	Ala	Ala	Leu	Arg	Gln	Lys	Gln
2285						2290					2295			
Ala	Ala	Asp	Ala	Glu	Met	Glu	Lys	His	Lys	Lys	Phe	Ala	Glu	Gln
2300						2305					2310			
Thr	Leu	Arg	Gln	Lys	Ala	Gln	Val	Glu	Gln	Glu	Leu	Thr	Thr	Leu
2315						2320					2325			
Arg	Leu	Gln	Leu	Glu	Glu	Thr	Asp	His	Gln	Lys	Asn	Leu	Leu	Asp
2330						2335					2340			
Glu	Glu	Leu	Gln	Arg	Leu	Lys	Ala	Glu	Ala	Thr	Glu	Ala	Ala	Arg
2345						2350					2355			
Gln	Arg	Ser	Gln	Val	Glu	Glu	Glu	Leu	Phe	Ser	Val	Arg	Val	Gln
2360						2365					2370			

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Met	Glu	Glu	Leu	Ser	Lys	Leu	Lys	Ala	Arg	Ile	Glu	Ala	Glu	Asn
2375						2380					2385			
Arg	Ala	Leu	Ile	Leu	Arg	Asp	Lys	Asp	Asn	Thr	Gln	Arg	Phe	Leu
2390						2395					2400			
Gln	Glu	Glu	Ala	Glu	Lys	Met	Lys	Gln	Val	Ala	Glu	Glu	Ala	Ala
2405						2410					2415			
Arg	Leu	Ser	Val	Ala	Ala	Gln	Glu	Ala	Ala	Arg	Leu	Arg	Gln	Leu
2420						2425					2430			
Ala	Glu	Glu	Asp	Leu	Ala	Gln	Gln	Arg	Ala	Leu	Ala	Glu	Lys	Met
2435						2440					2445			
Leu	Lys	Glu	Lys	Met	Gln	Ala	Val	Gln	Glu	Ala	Thr	Arg	Leu	Lys
2450						2455					2460			
Ala	Glu	Ala	Glu	Leu	Leu	Gln	Gln	Gln	Lys	Glu	Leu	Ala	Gln	Glu
2465						2470					2475			
Gln	Ala	Arg	Arg	Leu	Gln	Glu	Asp	Lys	Glu	Gln	Met	Ala	Gln	Gln
2480						2485					2490			
Leu	Ala	Glu	Glu	Thr	Gln	Gly	Phe	Gln	Arg	Thr	Leu	Glu	Ala	Glu
2495						2500					2505			
Arg	Gln	Arg	Gln	Leu	Glu	Met	Ser	Ala	Glu	Ala	Glu	Arg	Leu	Lys
2510						2515					2520			
Leu	Arg	Val	Ala	Glu	Met	Ser	Arg	Ala	Gln	Ala	Arg	Ala	Glu	Glu
2525						2530					2535			
Asp	Ala	Gln	Arg	Phe	Arg	Lys	Gln	Ala	Glu	Glu	Ile	Gly	Glu	Lys
2540						2545					2550			
Leu	His	Arg	Thr	Glu	Leu	Ala	Thr	Gln	Glu	Lys	Val	Thr	Leu	Val
2555						2560					2565			
Gln	Thr	Leu	Glu	Ile	Gln	Arg	Gln	Gln	Ser	Asp	His	Asp	Ala	Glu
2570						2575					2580			
Arg	Leu	Arg	Glu	Ala	Ile	Ala	Glu	Leu	Glu	Arg	Glu	Lys	Glu	Lys
2585						2590					2595			
Leu	Gln	Gln	Glu	Ala	Lys	Leu	Leu	Gln	Leu	Lys	Ser	Glu	Glu	Met
2600						2605					2610			
Gln	Thr	Val	Gln	Gln	Glu	Gln	Leu	Leu	Gln	Glu	Thr	Gln	Ala	Leu
2615						2620					2625			
Gln	Gln	Ser	Phe	Leu	Ser	Glu	Lys	Asp	Ser	Leu	Leu	Gln	Arg	Glu
2630						2635					2640			
Arg	Phe	Ile	Glu	Gln	Glu	Lys	Ala	Lys	Leu	Glu	Gln	Leu	Phe	Gln
2645						2650					2655			
Asp	Glu	Val	Ala	Lys	Ala	Gln	Gln	Leu	Arg	Glu	Glu	Gln	Gln	Arg
2660						2665					2670			
Gln	Gln	Gln	Gln	Met	Glu	Gln	Glu	Arg	Gln	Arg	Leu	Val	Ala	Ser
2675						2680					2685			
Met	Glu	Glu	Ala	Arg	Arg	Arg	Gln	His	Glu	Ala	Glu	Glu	Gly	Val
2690						2695					2700			
Arg	Arg	Lys	Gln	Glu	Glu	Leu	Gln	Gln	Leu	Glu	Gln	Gln	Arg	Arg
2705						2710					2715			
Gln	Gln	Glu	Glu	Leu	Leu	Ala	Glu	Glu	Asn	Gln	Arg	Leu	Arg	Glu
2720						2725					2730			
Gln	Leu	Gln	Leu	Leu	Glu	Glu	Gln	His	Arg	Ala	Ala	Leu	Ala	His
2735						2740					2745			
Ser	Glu	Glu	Val	Thr	Ala	Ser	Gln	Val	Ala	Ala	Thr	Lys	Thr	Leu

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2750	2755	2760
Pro Asn Gly Arg Asp Ala Leu	Asp Gly Pro Ala Ala	Glu Ala Glu
2765	2770	2775
Pro Glu His Ser Phe Asp Gly	Leu Arg Arg Lys Val	Ser Ala Gln
2780	2785	2790
Arg Leu Gln Glu Ala Gly Ile	Leu Ser Ala Glu Glu	Leu Gln Arg
2795	2800	2805
Leu Ala Gln Gly His Thr Thr	Val Asp Glu Leu Ala	Arg Arg Glu
2810	2815	2820
Asp Val Arg His Tyr Leu Gln	Gly Arg Ser Ser Ile	Ala Gly Leu
2825	2830	2835
Leu Leu Lys Ala Thr Asn Glu	Lys Leu Ser Val Tyr	Ala Ala Leu
2840	2845	2850
Gln Arg Gln Leu Leu Ser Pro	Gly Thr Ala Leu Ile	Leu Leu Glu
2855	2860	2865
Ala Gln Ala Ala Ser Gly Phe	Leu Leu Asp Pro Val	Arg Asn Arg
2870	2875	2880
Arg Leu Thr Val Asn Glu Ala	Val Lys Glu Gly Val	Val Gly Pro
2885	2890	2895
Glu Leu His His Lys Leu Leu	Ser Ala Glu Arg Ala	Val Thr Gly
2900	2905	2910
Tyr Lys Asp Pro Tyr Thr Gly	Gln Gln Ile Ser Leu	Phe Gln Ala
2915	2920	2925
Met Gln Lys Gly Leu Ile Val	Arg Glu His Gly Ile	Arg Leu Leu
2930	2935	2940
Glu Ala Gln Ile Ala Thr Gly	Gly Val Ile Asp Pro	Val His Ser
2945	2950	2955
His Arg Val Pro Val Asp Val	Ala Tyr Arg Arg Gly	Tyr Phe Asp
2960	2965	2970
Glu Glu Met Asn Arg Val Leu	Ala Asp Pro Ser Asp	Asp Thr Lys
2975	2980	2985
Gly Phe Phe Asp Pro Asn Thr	His Glu Asn Leu Thr	Tyr Leu Gln
2990	2995	3000
Leu Leu Glu Arg Cys Val Glu	Asp Pro Glu Thr Gly	Leu Cys Leu
3005	3010	3015
Leu Pro Leu Thr Asp Lys Ala	Ala Lys Gly Gly Glu	Leu Val Tyr
3020	3025	3030
Thr Asp Ser Glu Ala Arg Asp	Val Phe Glu Lys Ala	Thr Val Ser
3035	3040	3045
Ala Pro Phe Gly Lys Phe Gln	Gly Lys Thr Val Thr	Ile Trp Glu
3050	3055	3060
Ile Ile Asn Ser Glu Tyr Phe	Thr Ala Glu Gln Arg	Arg Asp Leu
3065	3070	3075
Leu Arg Gln Phe Arg Thr Gly	Arg Ile Thr Val Glu	Lys Ile Ile
3080	3085	3090
Lys Ile Ile Ile Thr Val Val	Glu Glu Gln Glu Gln	Lys Gly Arg
3095	3100	3105
Leu Cys Phe Glu Gly Leu Arg	Ser Leu Val Pro Ala	Ala Glu Leu
3110	3115	3120
Leu Glu Ser Arg Val Ile Asp	Arg Glu Leu Tyr Gln	Gln Leu Gln
3125	3130	3135

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Arg Gly	Glu Arg Ser Val	Arg Asp Val Ala Glu Val	Asp Thr Val
3140		3145	3150
Arg Arg	Ala Leu Arg Gly Ala	Asn Val Ile Ala Gly	Val Trp Leu
3155		3160	3165
Glu Glu	Ala Gly Gln Lys Leu	Ser Ile Tyr Asn Ala	Leu Lys Lys
3170		3175	3180
Asp Leu	Leu Pro Ser Asp Met	Ala Val Ala Leu Leu	Glu Ala Gln
3185		3190	3195
Ala Gly	Thr Gly His Ile Ile	Asp Pro Ala Thr Ser	Ala Arg Leu
3200		3205	3210
Thr Val	Asp Glu Ala Val Arg	Ala Gly Leu Val Gly	Pro Glu Phe
3215		3220	3225
His Glu	Lys Leu Leu Ser Ala	Glu Lys Ala Val Thr	Gly Tyr Arg
3230		3235	3240
Asp Pro	Tyr Thr Gly Gln Ser	Val Ser Leu Phe Gln	Ala Leu Lys
3245		3250	3255
Lys Gly	Leu Ile Pro Arg Glu	Gln Gly Leu Arg Leu	Leu Asp Ala
3260		3265	3270
Gln Leu	Ser Thr Gly Gly Ile	Val Asp Pro Ser Lys	Ser His Arg
3275		3280	3285
Val Pro	Leu Asp Val Ala Cys	Ala Arg Gly Cys Leu	Asp Glu Glu
3290		3295	3300
Thr Ser	Arg Ala Leu Ser Ala	Pro Arg Ala Asp Ala	Lys Ala Tyr
3305		3310	3315
Ser Asp	Pro Ser Thr Gly Glu	Pro Ala Thr Tyr Gly	Glu Leu Gln
3320		3325	3330
Gln Arg	Cys Arg Pro Asp Gln	Leu Thr Gly Leu Ser	Leu Leu Pro
3335		3340	3345
Leu Ser	Glu Lys Ala Ala Arg	Ala Arg Gln Glu Glu	Leu Tyr Ser
3350		3355	3360
Glu Leu	Gln Ala Arg Glu Thr	Phe Glu Lys Thr Pro	Val Glu Val
3365		3370	3375
Pro Val	Gly Gly Phe Lys Gly	Arg Thr Val Thr Val	Trp Glu Leu
3380		3385	3390
Ile Ser	Ser Glu Tyr Phe Thr	Ala Glu Gln Arg Gln	Glu Leu Leu
3395		3400	3405
Arg Gln	Phe Arg Thr Gly Lys	Val Thr Val Glu Lys	Val Ile Lys
3410		3415	3420
Ile Leu	Ile Thr Ile Val Glu	Glu Val Glu Thr Leu	Arg Gln Glu
3425		3430	3435
Arg Leu	Ser Phe Ser Gly Leu	Arg Ala Pro Val Pro	Ala Ser Glu
3440		3445	3450
Leu Leu	Ala Ser Gly Val Leu	Ser Arg Ala Gln Phe	Glu Gln Leu
3455		3460	3465
Lys Asp	Gly Lys Thr Thr Val	Lys Asp Leu Ser Glu	Leu Gly Ser
3470		3475	3480
Val Arg	Thr Leu Leu Gln Gly	Ser Gly Cys Leu Ala	Gly Ile Tyr
3485		3490	3495
Leu Glu	Asp Thr Lys Glu Lys	Val Ser Ile Tyr Glu	Ala Met Arg
3500		3505	3510

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Arg Gly	Leu Leu Arg Ala Thr	Thr Ala Ala Leu Leu	Leu Glu Ala
3515	3520	3525	
Gln Ala	Ala Thr Gly Phe Leu	Val Asp Pro Val Arg	Asn Gln Arg
3530	3535	3540	
Leu Tyr	Val His Glu Ala Val	Lys Ala Gly Val Val	Gly Pro Glu
3545	3550	3555	
Leu His	Glu Gln Leu Leu Ser	Ala Glu Lys Ala Val	Thr Gly Tyr
3560	3565	3570	
Arg Asp	Pro Tyr Ser Gly Ser	Thr Ile Ser Leu Phe	Gln Ala Met
3575	3580	3585	
Gln Lys	Gly Leu Val Leu Arg	Gln His Gly Ile Arg	Leu Leu Glu
3590	3595	3600	
Ala Gln	Ile Ala Thr Gly Gly	Ile Ile Asp Pro Val	His Ser His
3605	3610	3615	
Arg Val	Pro Val Asp Val Ala	Tyr Gln Arg Gly Tyr	Phe Ser Glu
3620	3625	3630	
Glu Met	Asn Arg Val Leu Ala	Asp Pro Ser Asp Asp	Thr Lys Gly
3635	3640	3645	
Phe Phe	Asp Pro Asn Thr His	Glu Asn Leu Thr Tyr	Arg Gln Leu
3650	3655	3660	
Leu Glu	Arg Cys Val Glu Asp	Pro Glu Thr Gly Leu	Arg Leu Leu
3665	3670	3675	
Pro Leu	Lys Gly Ala Glu Lys	Ala Glu Val Val Glu	Thr Thr Gln
3680	3685	3690	
Val Tyr	Thr Glu Glu Glu Thr	Arg Arg Ala Phe Glu	Glu Thr Gln
3695	3700	3705	
Ile Asp	Ile Pro Gly Gly Gly	Ser His Gly Gly Ser	Thr Met Ser
3710	3715	3720	
Leu Trp	Glu Val Met Gln Ser	Asp Leu Ile Pro Glu	Glu Gln Arg
3725	3730	3735	
Ala Gln	Leu Met Ala Asp Phe	Gln Ala Gly Arg Val	Thr Lys Glu
3740	3745	3750	
Arg Met	Ile Ile Ile Ile Ile	Glu Ile Ile Glu Lys	Thr Glu Ile
3755	3760	3765	
Ile Arg	Gln Gln Gly Leu Ala	Ser Tyr Asp Tyr Val	Arg Arg Arg
3770	3775	3780	
Leu Thr	Ala Glu Asp Leu Phe	Glu Ala Arg Ile Ile	Ser Leu Glu
3785	3790	3795	
Thr Tyr	Asn Leu Leu Arg Glu	Gly Thr Arg Ser Leu	Arg Glu Ala
3800	3805	3810	
Leu Glu	Ala Glu Ser Ala Trp	Cys Tyr Leu Tyr Gly	Thr Gly Ser
3815	3820	3825	
Val Ala	Gly Val Tyr Leu Pro	Gly Ser Arg Gln Thr	Leu Ser Ile
3830	3835	3840	
Tyr Gln	Ala Leu Lys Lys Gly	Leu Leu Ser Ala Glu	Val Ala Arg
3845	3850	3855	
Leu Leu	Leu Glu Ala Gln Ala	Ala Thr Gly Phe Leu	Leu Asp Pro
3860	3865	3870	
Val Lys	Gly Glu Arg Leu Thr	Val Asp Glu Ala Val	Arg Lys Gly
3875	3880	3885	
Leu Val	Gly Pro Glu Leu His	Asp Arg Leu Leu Ser	Ala Glu Arg

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Val Ile	Val Asp	Pro Glu	Thr	Gly Lys	Glu Met	Ser	Val Tyr	Glu
4280			4285			4290		
Ala Tyr	Arg Lys	Gly Leu	Ile	Asp His	Gln Thr	Tyr	Leu Glu	Leu
4295			4300			4305		
Ser Glu	Gln Glu	Cys Glu	Trp	Glu Glu	Ile Thr	Ile	Ser Ser	Ser
4310			4315			4320		
Asp Gly	Val Val	Lys Ser	Met	Ile Ile	Asp Arg	Arg	Ser Gly	Arg
4325			4330			4335		
Gln Tyr	Asp Ile	Asp Asp	Ala	Ile Ala	Lys Asn	Leu	Ile Asp	Arg
4340			4345			4350		
Ser Ala	Leu Asp	Gln Tyr	Arg	Ala Gly	Thr Leu	Ser	Ile Thr	Glu
4355			4360			4365		
Phe Ala	Asp Met	Leu Ser	Gly	Asn Ala	Gly Gly	Phe	Arg Ser	Arg
4370			4375			4380		
Ser Ser	Ser Val	Gly Ser	Ser	Ser Ser	Tyr Pro	Ile	Ser Pro	Ala
4385			4390			4395		
Val Ser	Arg Thr	Gln Leu	Ala	Ser Trp	Ser Asp	Pro	Thr Glu	Glu
4400			4405			4410		
Thr Gly	Pro Val	Ala Gly	Ile	Leu Asp	Thr Glu	Thr	Leu Glu	Lys
4415			4420			4425		
Val Ser	Ile Thr	Glu Ala	Met	His Arg	Asn Leu	Val	Asp Asn	Ile
4430			4435			4440		
Thr Gly	Gln Arg	Leu Leu	Glu	Ala Gln	Ala Cys	Thr	Gly Gly	Ile
4445			4450			4455		
Ile Asp	Pro Ser	Thr Gly	Glu	Arg Phe	Pro Val	Thr	Asp Ala	Val
4460			4465			4470		
Asn Lys	Gly Leu	Val Asp	Lys	Ile Met	Val Asp	Arg	Ile Asn	Leu
4475			4480			4485		
Ala Gln	Lys Ala	Phe Cys	Gly	Phe Glu	Asp Pro	Arg	Thr Lys	Thr
4490			4495			4500		
Lys Met	Ser Ala	Ala Gln	Ala	Leu Lys	Lys Gly	Trp	Leu Tyr	Tyr
4505			4510			4515		
Glu Ala	Gly Gln	Arg Phe	Leu	Glu Val	Gln Tyr	Leu	Thr Gly	Gly
4520			4525			4530		
Leu Ile	Glu Pro	Asp Thr	Pro	Gly Arg	Val Pro	Leu	Asp Glu	Ala
4535			4540			4545		
Leu Gln	Arg Gly	Thr Val	Asp	Ala Arg	Thr Ala	Gln	Lys Leu	Arg
4550			4555			4560		
Asp Val	Gly Ala	Tyr Ser	Lys	Tyr Leu	Thr Cys	Pro	Lys Thr	Lys
4565			4570			4575		
Leu Lys	Ile Ser	Tyr Lys	Asp	Ala Leu	Asp Arg	Ser	Met Val	Glu
4580			4585			4590		
Glu Gly	Thr Gly	Leu Arg	Leu	Leu Glu	Ala Ala	Ala	Gln Ser	Thr
4595			4600			4605		
Lys Gly	Tyr Tyr	Ser Pro	Tyr	Ser Val	Ser Gly	Ser	Gly Ser	Thr
4610			4615			4620		
Ala Gly	Ser Arg	Thr Gly	Ser	Arg Thr	Gly Ser	Arg	Ala Gly	Ser
4625			4630			4635		
Arg Arg	Gly Ser	Phe Asp	Ala	Thr Gly	Ser Gly	Phe	Ser Met	Thr
4640			4645			4650		

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Phe Ser Ser Ser Ser Tyr Ser Ser Ser Gly Tyr Gly Arg Arg Tyr
 4655 4660 4665
 Ala Ser Gly Ser Ser Ala Ser Leu Gly Gly Pro Glu Ser Ala Val
 4670 4675 4680

Ala

<210> SEQ ID NO 9
 <211> LENGTH: 2353
 <212> TYPE: PRT
 <213> ORGANISM: Xenopus laevis

<400> SEQUENCE: 9

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 1 5 10 15
 Met Gln Phe Glu Pro Ser Thr Met Ile Tyr Asp Ala Cys Arg Ile Ile
 20 25 30
 Arg Glu Lys Val Pro Glu Ala Gln Ile Gly Gln Pro Asn Asp Phe Gly
 35 40 45
 Leu Phe Leu Ser Asp Glu Asp Pro Lys Lys Gly Ile Trp Leu Glu Ala
 50 55 60
 Gly Lys Ala Leu Asp Tyr Tyr Met Leu Arg Asn Gly Asp Thr Leu Glu
 65 70 75 80
 Tyr Arg Lys Lys Gln Arg Pro Leu Glu Ile Arg Met Leu Asp Gly Thr
 85 90 95
 Val Lys Thr Val Met Val Asp Asp Ser Asn Thr Met Ser Asp Leu Leu
 100 105 110
 Met Thr Ile Cys Ala Arg Ile Gly Ile Thr Asn Tyr Asp Glu Tyr Ser
 115 120 125
 Leu Val Arg Glu Ile Met Glu Glu Lys Lys Glu Glu Val Thr Gly Thr
 130 135 140
 Leu Lys Arg Asp Lys Thr Leu Leu Arg Asp Asp Lys Lys Met Glu Lys
 145 150 155 160
 Leu Lys Gln Lys Leu His Thr Asp Asp Glu Leu Asn Trp Leu Asp Pro
 165 170 175
 Gly Arg Thr Leu Arg Glu Gln Gly Val Asp Glu Asn Glu Thr Leu Leu
 180 185 190
 Leu Arg Arg Lys Phe Phe Tyr Ser Asp Gln Asn Val Asp Ser Arg Asp
 195 200 205
 Pro Val Gln Leu Asn Leu Leu Tyr Val Gln Ala Arg Asp Asp Ile Leu
 210 215 220
 Asn Gly Ser His Pro Val Ser Phe Asp Lys Ala Cys Glu Phe Ala Gly
 225 230 235 240
 Tyr Gln Cys Gln Val Gln Phe Gly Pro His Asn Glu Val Lys His Lys
 245 250 255
 Pro Gly Phe Leu Glu Leu Lys Asp Phe Leu Pro Lys Glu Tyr Ile Lys
 260 265 270
 Gln Lys Gly Glu Arg Lys Ile Phe Leu Ala His Lys Gln Cys Gly Asn
 275 280 285
 Met Ser Glu Ile Glu Ala Lys Ala Arg Tyr Val Lys Leu Ala Arg Ser
 290 295 300
 Leu Lys Thr Tyr Gly Val Ser Phe Phe Leu Val Lys Glu Lys Met Lys
 305 310 315 320

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725					730					735					
Leu	Glu	Ala	Gly	Lys	Gln	Val	Ala	Lys	Ser	Val	Glu	Gly	Cys	Val	Glu
			740					745					750		
Ala	Ser	Glu	Ala	Ala	Val	Glu	Asp	Pro	Glu	Leu	Leu	Lys	Ser	Val	Gly
		755					760					765			
Val	Ala	Ala	Ser	Gly	Val	Thr	Gln	Ala	Leu	Asn	Asn	Leu	Leu	Gln	His
		770					775					780			
Ile	Lys	Lys	His	Ala	Ser	Gly	Gly	Pro	Ser	Thr	Gly	Arg	Tyr	Asp	Gln
				790								795			800
Ala	Thr	Asp	Thr	Ile	Leu	Asn	Val	Thr	Glu	Asn	Ile	Phe	Ser	Ser	Met
				805					810						815
Gly	Asp	Ala	Gly	Glu	Met	Val	Arg	Gln	Ala	Arg	Ile	Leu	Ala	Gln	Ala
			820					825					830		
Thr	Ser	Asp	Leu	Val	Gly	Ala	Ile	Lys	Ala	Asp	Ala	Glu	Arg	Glu	Ser
			835					840				845			
Asp	Leu	Glu	Asn	Ser	Arg	Lys	Leu	Leu	Cys	Ala	Ala	Lys	Leu	Leu	Ala
		850				855						860			
Asp	Ala	Thr	Ala	Arg	Met	Val	Glu	Ala	Ala	Lys	Gly	Ala	Ala	Ala	His
				870							875				880
Pro	Asp	Ser	Glu	Glu	Gln	Gln	Gln	Lys	Leu	Arg	Glu	Ala	Ala	Glu	Gly
				885					890						895
Leu	Arg	Met	Ala	Thr	Asn	Ala	Ala	Ala	Gln	Asn	Ala	Ile	Lys	Lys	Lys
			900						905					910	
Leu	Val	His	Lys	Leu	Glu	Gln	Ala	Ala	Lys	Gln	Ala	Ala	Ala	Ser	Ala
			915				920						925		
Thr	Gln	Thr	Ile	Ala	Ala	Ala	Gln	Asn	Ala	Ala	Ser	Ser	Asn	Lys	Asn
			930				935						940		
Pro	Ala	Ala	Gln	Gln	Gln	Leu	Val	Gln	Ser	Cys	Lys	Val	Val	Ala	Glu
				950							955				960
Gln	Ile	Pro	Met	Leu	Val	Gln	Gly	Val	Arg	Gly	Ser	Gln	Ser	Gln	Pro
				965					970					975	
Asp	Ser	Pro	Ser	Ala	Gln	Leu	Ser	Leu	Ile	Ser	Ala	Ser	Gln	Asn	Phe
				980				985						990	
Leu	Gln	Pro	Gly	Ala	Lys	Leu	Val	Thr	Ala	Gly	Lys	Ser	Ala	Val	Pro
			995				1000						1005		
Thr	Val	Ser	Asp	Pro	Ala	Ser	Ala	Met	Gln	Leu	Gly	Gln	Cys	Thr	
							1015					1020			
Lys	Asn	Leu	Ala	Ser	Ala	Leu	Ala	Glu	Leu	Arg	Thr	Ala	Ala	Gln	
							1030					1035			
Lys	Ala	His	Glu	Ala	Cys	Gly	Pro	Leu	Glu	Ile	Asp	Ser	Ala	Leu	
							1045					1050			
Asn	Val	Val	Arg	Ser	Leu	Glu	Gln	Asp	Leu	Gln	Glu	Ala	Arg	Ala	
							1060					1065			
Ala	Ala	Arg	Glu	Gly	Lys	Leu	Gln	Pro	Leu	Pro	Gly	Glu	Thr	Met	
							1075					1080			
Glu	Lys	Cys	Ala	Gln	Asp	Leu	Gly	Ser	Ser	Thr	Lys	Ala	Val	Ser	
							1090					1095			
Ser	Ser	Ile	Ala	Gln	Leu	Leu	Gly	Glu	Ile	Val	His	Gly	Asn	Glu	
							1105					1110			
Asn	Tyr	Thr	Gly	Arg	Ala	Ala	Arg	Asp	Val	Ala	Gln	Ala	Leu	Arg	
							1120					1125			

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Ser	Leu	Ala	Gln	Ala	Ser	Arg	Gly	Val	Ala	Ala	Asn	Ser	Thr	Asp
1130						1135					1140			
Pro	Ala	Val	Gln	Asn	Ala	Met	Leu	Glu	Cys	Ala	Glu	Asp	Val	Met
1145						1150					1155			
Asp	Lys	Ala	Gly	Asn	Leu	Ile	Glu	Glu	Ala	Lys	Arg	Ala	Val	Gly
1160						1165					1170			
Lys	Pro	Thr	Asp	Pro	Glu	Gly	Gln	Gln	Arg	Leu	Val	Gln	Val	Ala
1175						1180					1185			
Lys	Ala	Val	Ser	Gln	Ala	Leu	Ser	Arg	Cys	Val	Asn	Cys	Leu	Pro
1190						1195					1200			
Gly	Gln	Arg	Asp	Val	Asp	Ala	Ala	Ile	Lys	Ser	Ile	Gly	Glu	Ala
1205						1210					1215			
Ser	Lys	Ile	Leu	Leu	Ala	Ser	Ser	Phe	Pro	Ser	Gly	Thr	Lys	Asn
1220						1225					1230			
Phe	Gln	Glu	Ala	Gln	Ser	Gln	Leu	Asn	Gln	Ala	Ala	Ala	Gly	Leu
1235						1240					1245			
Asn	Gln	Ser	Ala	Asn	Glu	Leu	Val	Gln	Ala	Ser	Arg	Thr	Thr	Pro
1250						1255					1260			
Gln	Glu	Leu	Ala	Lys	Ala	Ser	Gly	Lys	Tyr	Ser	Gln	Asp	Phe	Asn
1265						1270					1275			
Glu	Phe	Leu	Gln	Ala	Gly	Val	Glu	Met	Ala	Gly	Gln	Ser	Gln	Asn
1280						1285					1290			
Lys	Glu	Asp	Gln	Ala	Gln	Val	Val	Ser	Asn	Leu	Lys	Ser	Ile	Ser
1295						1300					1305			
Leu	Ser	Ser	Ser	Lys	Leu	Leu	Leu	Ala	Ala	Lys	Ala	Leu	Ser	Ala
1310						1315					1320			
Asp	Pro	Ala	Ala	Pro	Asn	Leu	Lys	Asn	Gln	Leu	Ala	Ala	Ala	Ala
1325						1330					1335			
Arg	Ala	Val	Thr	Asp	Ser	Ile	Asn	Gln	Leu	Ile	Thr	Val	Cys	Thr
1340						1345					1350			
Gln	Gln	Ala	Pro	Gly	Gln	Lys	Glu	Cys	Asp	Asn	Ala	Leu	Arg	Glu
1355						1360					1365			
Leu	Glu	Thr	Val	Arg	Glu	Leu	Leu	Gln	Asn	Pro	Thr	Gln	Pro	Val
1370						1375					1380			
Asn	Asp	Gln	Ser	Tyr	Phe	His	Cys	Leu	Asp	Ser	Val	Met	Glu	Asn
1385						1390					1395			
Ser	Lys	Val	Leu	Gly	Glu	Ser	Met	Ala	Gly	Ile	Ser	Gln	Asn	Ala
1400						1405					1410			
Lys	Thr	Ser	Asn	Leu	Pro	Glu	Phe	Gly	Glu	Ser	Val	Gly	Ala	Ala
1415						1420					1425			
Ser	Lys	Ala	Leu	Cys	Gly	Leu	Thr	Glu	Ala	Ala	Ala	Gln	Ala	Ala
1430						1435					1440			
Tyr	Leu	Val	Gly	Val	Ser	Asp	Ala	Asn	Ser	His	Ala	Gly	Met	Gln
1445						1450					1455			
Gly	Leu	Val	Asp	Pro	Thr	Gln	Phe	Ala	Arg	Ala	Asn	Gln	Ala	Ile
1460						1465					1470			
Gln	Met	Ala	Cys	Gln	Asn	Leu	Gly	Asp	Pro	Ala	Cys	Thr	Gln	Ser
1475						1480					1485			
Gln	Val	Leu	Ser	Ala	Ala	Thr	Ile	Val	Ala	Lys	His	Thr	Ser	Ala
1490						1495					1500			

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Leu	Cys	Asn	Ala	Cys	Arg	Val	Ala	Ser	Thr	His	Thr	Ser	Asn	Pro
1505						1510					1515			
Val	Ala	Lys	Arg	Gln	Phe	Val	Gln	Ser	Ala	Lys	Glu	Val	Ala	Asn
1520						1525					1530			
Ser	Thr	Ala	Asn	Leu	Val	Lys	Thr	Ile	Lys	Ala	Leu	Asp	Gly	Thr
1535						1540					1545			
Phe	Asn	Asp	Glu	Asn	Arg	Val	Lys	Cys	Arg	Asn	Ala	Thr	Val	Pro
1550						1555					1560			
Leu	Ile	Gln	Ala	Val	Glu	Asn	Leu	Thr	Ala	Phe	Ala	Ser	Asn	Pro
1565						1570					1575			
Glu	Phe	Ala	Ser	Val	Pro	Ala	Gln	Ile	Ser	Pro	Glu	Gly	Leu	Arg
1580						1585					1590			
Ala	Met	Glu	Pro	Ile	Val	Thr	Ala	Ala	Lys	Leu	Met	Leu	Glu	Ser
1595						1600					1605			
Ser	Ser	Gly	Leu	Ile	Gln	Thr	Ala	Arg	Ser	Leu	Ala	Ala	Asn	Pro
1610						1615					1620			
Lys	Asp	Pro	Pro	Gln	Trp	Ser	Val	Leu	Ala	Gly	His	Ser	Arg	Asn
1625						1630					1635			
Val	Ser	Asp	Ser	Ile	Lys	Lys	Leu	Ile	Thr	Asn	Met	Arg	Asp	Lys
1640						1645					1650			
Ala	Pro	Gly	Gln	Arg	Glu	Cys	Asp	Gln	Ala	Ile	Glu	Leu	Leu	Asn
1655						1660					1665			
Gln	Ala	Val	Arg	Asp	Leu	Asp	Gln	Ala	Ser	Leu	Glu	Ala	Ile	Ser
1670						1675					1680			
Gln	Gln	Leu	Ala	Pro	Arg	Glu	Gly	Ile	Ser	Gln	Glu	Ala	Leu	His
1685						1690					1695			
Asn	Gln	Met	Gln	Thr	Ser	Val	Gln	Glu	Ile	Ser	Asn	Leu	Ile	Glu
1700						1705					1710			
Pro	Met	Ala	Ala	Ala	Ala	Arg	Ala	Asp	Ser	Ser	Gln	Leu	Gly	His
1715						1720					1725			
Lys	Val	Ser	Gln	Met	Ala	Gln	Tyr	Phe	Glu	Pro	Leu	Thr	His	Ala
1730						1735					1740			
Ser	Ile	Gly	Thr	Ala	Ser	Lys	Thr	Ile	Asn	His	Gln	Gln	Gln	Met
1745						1750					1755			
Asn	Leu	Leu	Asp	Gln	Thr	Lys	Thr	Leu	Ala	Glu	Ser	Ala	Leu	Gln
1760						1765					1770			
Met	Leu	Tyr	Thr	Ala	Lys	Glu	Ala	Gly	Gly	Asn	Pro	Lys	Val	Ala
1775						1780					1785			
Ala	Gln	Thr	Gln	Glu	Ala	Leu	Asp	Glu	Ala	Ala	Gln	Met	Met	His
1790						1795					1800			
Glu	Ala	Val	Gly	Asp	Leu	Thr	Val	Thr	Leu	Asn	Glu	Ala	Ala	Ser
1805						1810					1815			
Ala	Ala	Gly	Ala	Val	Gly	Gly	Met	Val	Asp	Ser	Ile	Thr	Gln	Ala
1820						1825					1830			
Ile	Asn	Lys	Leu	Asp	Glu	Glu	Pro	Thr	Gly	Glu	Pro	Glu	Gly	Ser
1835						1840					1845			
Phe	Val	Asp	Tyr	Gln	Thr	Thr	Met	Val	Lys	Thr	Ala	Lys	Ala	Ile
1850						1855					1860			
Ala	Val	Thr	Val	Gln	Glu	Met	Val	Thr	Lys	Ser	Thr	Thr	Asn	Pro
1865						1870					1875			
Asp	Glu	Leu	Gly	Thr	Leu	Ala	Asn	Gln	Leu	Thr	Asn	Glu	Tyr	Ser

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Ala Ala Tyr Ser Lys Gln Val Ala Gly Ser Val Thr Glu Leu Ile
 2270 2275 2280

Gln Ala Ala Glu Ala Met Lys Gly Thr Glu Trp Val Asp Pro Glu
 2285 2290 2295

Asp Pro Thr Val Ile Ala Glu Asn Glu Leu Leu Gly Ala Ala Ala
 2300 2305 2310

Ala Ile Glu Ala Ala Ala Lys Lys Leu Glu Gln Leu Lys Pro Arg
 2315 2320 2325

Ala Lys Pro Lys Gln Ala Asp Glu Ser Leu Asn Phe Glu Glu Gln
 2330 2335 2340

Ile Leu Glu Ala Ala Lys Ser Ile Ala Ala
 2345 2350

<210> SEQ ID NO 10

<211> LENGTH: 1006

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 10

Met Ala Ala Ala Tyr Leu Asp Pro Asn Leu Asn His Thr Pro Asn Ser
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Ser Thr Lys Thr His Leu Gly Thr Gly Met Glu Arg Ser Pro Gly Ala
 20 25 30

Met Glu Arg Val Leu Lys Val Phe His Tyr Phe Glu Ser Asn Ser Glu
 35 40 45

Pro Thr Thr Trp Ala Ser Ile Ile Arg His Gly Asp Ala Thr Asp Val
 50 55 60

Arg Gly Ile Ile Gln Lys Ile Val Asp Ser His Lys Val Lys His Val
 65 70 75 80

Ala Cys Tyr Gly Phe Arg Leu Ser His Leu Arg Ser Glu Glu Val His
 85 90 95

Trp Leu His Val Asp Met Gly Val Ser Ser Val Arg Glu Lys Tyr Glu
 100 105 110

Leu Ala His Pro Pro Glu Glu Trp Lys Tyr Glu Leu Arg Ile Arg Tyr
 115 120 125

Leu Pro Lys Gly Phe Leu Asn Gln Phe Thr Glu Asp Lys Pro Thr Leu
 130 135 140

Asn Phe Phe Tyr Gln Gln Val Lys Ser Asp Tyr Met Leu Glu Ile Ala
 145 150 155 160

Asp Gln Val Asp Gln Glu Ile Ala Leu Lys Leu Gly Cys Leu Glu Ile
 165 170 175

Arg Arg Ser Tyr Trp Glu Met Arg Gly Asn Ala Leu Glu Lys Lys Ser
 180 185 190

Asn Tyr Glu Val Leu Glu Lys Asp Val Gly Leu Lys Arg Phe Phe Pro
 195 200 205

Lys Ser Leu Leu Asp Ser Val Lys Ala Lys Thr Leu Arg Lys Leu Ile
 210 215 220

Gln Gln Thr Phe Arg Gln Phe Ala Asn Leu Asn Arg Glu Glu Ser Ile
 225 230 235 240

Leu Lys Phe Phe Glu Ile Leu Ser Pro Val Tyr Arg Phe Asp Lys Glu
 245 250 255

Cys Phe Lys Cys Ala Leu Gly Ser Ser Trp Ile Ile Ser Val Glu Leu

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260				265				270							
Ala	Ile	Gly	Pro	Glu	Glu	Gly	Ile	Ser	Tyr	Leu	Thr	Asp	Lys	Gly	Cys
		275					280					285			
Asn	Pro	Thr	His	Leu	Ala	Asp	Phe	Thr	Gln	Val	Gln	Thr	Ile	Gln	Tyr
	290					295					300				
Ser	Asn	Ser	Glu	Asp	Lys	Asp	Arg	Lys	Gly	Met	Leu	Gln	Leu	Lys	Ile
305					310					315					320
Ala	Gly	Ala	Pro	Glu	Pro	Leu	Thr	Val	Thr	Ala	Pro	Ser	Leu	Thr	Ile
			325							330					335
Ala	Glu	Asn	Met	Ala	Asp	Leu	Ile	Asp	Gly	Tyr	Cys	Arg	Leu	Val	Asn
			340						345				350		
Gly	Thr	Ser	Gln	Ser	Phe	Ile	Ile	Arg	Pro	Gln	Lys	Glu	Gly	Glu	Arg
		355					360						365		
Ala	Leu	Pro	Ser	Ile	Pro	Lys	Leu	Ala	Asn	Ser	Glu	Lys	Gln	Gly	Met
	370					375					380				
Arg	Thr	His	Ala	Val	Ser	Val	Ser	Glu	Thr	Asp	Asp	Tyr	Ala	Glu	Ile
385					390					395					400
Ile	Asp	Glu	Glu	Asp	Thr	Tyr	Thr	Met	Pro	Ser	Thr	Arg	Asp	Tyr	Glu
			405						410						415
Ile	Gln	Arg	Glu	Arg	Ile	Glu	Leu	Gly	Arg	Cys	Ile	Gly	Glu	Gly	Gln
			420						425						430
Phe	Gly	Asp	Val	His	Gln	Gly	Ile	Tyr	Met	Ser	Pro	Glu	Asn	Pro	Ala
		435					440						445		
Leu	Ala	Val	Ala	Ile	Lys	Thr	Cys	Lys	Asn	Cys	Thr	Ser	Asp	Ser	Val
	450					455					460				
Arg	Glu	Lys	Phe	Leu	Gln	Glu	Ala	Leu	Thr	Met	Arg	Gln	Phe	Asp	His
465					470					475					480
Pro	His	Ile	Val	Lys	Leu	Ile	Gly	Val	Ile	Thr	Glu	Asn	Pro	Val	Trp
			485							490					495
Ile	Ile	Met	Glu	Leu	Cys	Thr	Leu	Gly	Glu	Leu	Arg	Ser	Phe	Leu	Gln
		500							505				510		
Val	Arg	Lys	Tyr	Ser	Leu	Asp	Leu	Ala	Ser	Leu	Ile	Leu	Tyr	Ala	Tyr
		515				520							525		
Gln	Leu	Ser	Thr	Ala	Leu	Ala	Tyr	Leu	Glu	Ser	Lys	Arg	Phe	Val	His
	530					535					540				
Arg	Asp	Ile	Ala	Ala	Arg	Asn	Val	Leu	Val	Ser	Ser	Asn	Asp	Cys	Val
545					550					555					560
Lys	Leu	Gly	Asp	Phe	Gly	Leu	Ser	Arg	Tyr	Met	Glu	Asp	Ser	Thr	Tyr
			565							570					575
Tyr	Lys	Ala	Ser	Lys	Gly	Lys	Leu	Pro	Ile	Lys	Trp	Met	Ala	Pro	Glu
		580							585				590		
Ser	Ile	Asn	Phe	Arg	Arg	Phe	Thr	Ser	Ala	Ser	Asp	Val	Trp	Met	Phe
		595					600						605		
Gly	Val	Cys	Met	Trp	Glu	Ile	Leu	Met	His	Gly	Val	Lys	Pro	Phe	Gln
	610					615					620				
Gly	Val	Lys	Asn	Asn	Asp	Val	Ile	Gly	Arg	Ile	Glu	Asn	Gly	Glu	Arg
	625				630					635					640
Leu	Pro	Met	Pro	Pro	Asn	Cys	Pro	Pro	Thr	Leu	Tyr	Ser	Leu	Met	Thr
			645						650						655
Lys	Cys	Trp	Ala	Tyr	Asp	Pro	Ser	Arg	Arg	Pro	Arg	Phe	Thr	Glu	Leu
			660						665						670

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Lys Ala Gln Leu Ser Thr Ile Leu Glu Glu Glu Lys Ala Gln Gln Glu
 675 680 685

Glu Arg Met Arg Met Glu Ser Arg Arg Gln Ala Thr Val Ser Trp Asp
 690 695 700

Ser Gly Gly Ser Asp Glu Ala Pro Pro Lys Pro Ser Arg Pro Gly Tyr
 705 710 715 720

Pro Ser Pro Arg Ser Ser Glu Gly Phe Tyr Pro Ser Pro Gln His Met
 725 730 735

Val Gln Thr Asn His Tyr Gln Asp Ser Thr Val Leu Asp Leu Arg Gly
 740 745 750

Ile Gly Gln Val Leu Pro Thr His Leu Met Glu Glu Arg Leu Ile Arg
 755 760 765

Gln Gln Gln Glu Met Glu Glu Asp Gln Arg Trp Leu Glu Lys Glu Glu
 770 775 780

Arg Phe Leu Lys Pro Asp Val Arg Leu Ser Arg Gly Ser Ile Asp Arg
 785 790 795 800

Glu Asp Gly Ser Leu Gln Gly Pro Ile Gly Asn Gln His Ile Tyr Gln
 805 810 815

Pro Val Gly Lys Pro Asp Pro Ala Ala Pro Pro Lys Lys Pro Pro Arg
 820 825 830

Pro Gly Ala Pro Gly His Leu Gly Ser Leu Ala Ser Leu Ser Ser Pro
 835 840 845

Ala Asp Ser Tyr Asn Glu Gly Val Lys Leu Gln Pro Gln Glu Ile Ser
 850 855 860

Pro Pro Pro Thr Ala Asn Leu Asp Arg Ser Asn Asp Lys Val Tyr Glu
 865 870 875 880

Asn Val Thr Gly Leu Val Lys Ala Val Ile Glu Met Ser Ser Lys Ile
 885 890 895

Gln Pro Ala Pro Pro Glu Glu Tyr Val Pro Met Val Lys Glu Val Gly
 900 905 910

Leu Ala Leu Arg Thr Leu Leu Ala Thr Val Asp Glu Thr Ile Pro Leu
 915 920 925

Leu Pro Ala Ser Thr His Arg Glu Ile Glu Met Ala Gln Lys Leu Leu
 930 935 940

Asn Ser Asp Leu Gly Glu Leu Ile Asn Lys Met Lys Leu Ala Gln Gln
 945 950 955 960

Tyr Val Met Thr Ser Leu Gln Gln Glu Tyr Lys Lys Gln Met Leu Thr
 965 970 975

Ala Ala His Ala Leu Ala Val Asp Ala Lys Asn Leu Leu Asp Val Ile
 980 985 990

Asp Gln Ala Arg Leu Lys Met Leu Gly Gln Thr Arg Pro His
 995 1000 1005

<210> SEQ ID NO 11
 <211> LENGTH: 268
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 11

Met Ala Val Asn Val Tyr Ser Thr Ser Val Thr Ser Asp Asn Leu Ser
 1 5 10 15

Arg His Asp Met Leu Ala Trp Ile Asn Glu Ser Leu Gln Leu Asn Leu

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Pro	Leu	Lys	Gly	Ile	Phe	Thr	Arg	Pro	Ser	Lys	Leu	Thr	Arg	Lys	Val
	115						120					125			
Gln	Ala	Glu	Asp	Glu	Ala	Asn	Gly	Leu	Gln	Thr	Ala	His	Ala	Arg	Ala
	130					135					140				
Ala	Ser	Pro	Leu	Ser	Thr	Ala	Ala	Ala	Thr	Met	Val	Ser	Ser	Ser	Pro
145					150					155					160
Ala	Thr	Pro	Ser	Asn	Ile	Pro	Gln	Lys	Pro	Ser	Gln	Pro	Val	Ala	Lys
				165					170						175
Glu	Thr	Ser	Ala	Thr	Pro	Gln	Ile	Ser	Asn	Leu	Thr	Lys	Thr	Ala	Ser
		180						185					190		
Glu	Ser	Ile	Ser	Asn	Leu	Ser	Glu	Ala	Gly	Ser	Val	Lys	Lys	Gly	Glu
	195						200					205			
Arg	Glu	Leu	Lys	Ile	Gly	Asp	Arg	Val	Leu	Val	Gly	Gly	Thr	Lys	Ala
	210					215					220				
Gly	Val	Val	Arg	Phe	Leu	Gly	Glu	Thr	Asp	Phe	Ala	Lys	Gly	Glu	Trp
225					230					235					240
Cys	Gly	Val	Glu	Leu	Asp	Glu	Pro	Leu	Gly	Lys	Asn	Asp	Gly	Ala	Val
				245					250					255	
Ala	Gly	Thr	Arg	Tyr	Phe	Gln	Cys	Gln	Pro	Lys	Tyr	Gly	Leu	Phe	Ala
			260					265					270		
Pro	Val	His	Lys	Val	Thr	Lys	Ile	Gly	Phe	Pro	Ser	Thr	Thr	Pro	Ala
		275					280					285			
Lys	Ala	Lys	Ala	Ala	Ala	Val	Arg	Arg	Val	Met	Ala	Thr	Thr	Pro	Ala
	290					295					300				
Ser	Leu	Lys	Arg	Ser	Pro	Ser	Ala	Ser	Ser	Leu	Ser	Ser	Met	Ser	Ser
305					310					315					320
Val	Ala	Ser	Ser	Val	Ser	Ser	Lys	Pro	Ser	Arg	Thr	Gly	Leu	Leu	Thr
				325					330					335	
Glu	Thr	Ser	Ser	Arg	Tyr	Ala	Arg	Lys	Ile	Ser	Gly	Thr	Thr	Ala	Leu
				340				345					350		
Gln	Glu	Ala	Leu	Lys	Glu	Lys	Gln	Gln	His	Ile	Glu	Gln	Leu	Leu	Ala
		355					360					365			
Glu	Arg	Asp	Leu	Glu	Arg	Ala	Glu	Val	Ala	Lys	Ala	Thr	Ser	His	Val
	370					375						380			
Gly	Glu	Ile	Glu	Gln	Glu	Leu	Ala	Leu	Ala	Arg	Asp	Gly	His	Asp	Gln
385					390					395					400
His	Val	Leu	Glu	Leu	Glu	Ala	Lys	Met	Asp	Gln	Leu	Arg	Thr	Met	Val
			405						410					415	
Glu	Ala	Ala	Asp	Arg	Glu	Lys	Val	Glu	Leu	Leu	Asn	Gln	Leu	Glu	Glu
			420					425					430		
Glu	Lys	Arg	Lys	Val	Glu	Asp	Leu	Gln	Phe	Arg	Val	Glu	Glu	Glu	Ser
		435					440						445		
Ile	Thr	Lys	Gly	Asp	Leu	Glu	Thr	Gln	Thr	Lys	Leu	Glu	His	Ala	Arg
	450					455						460			
Ile	Lys	Glu	Leu	Glu	Gln	Ser	Leu	Leu	Phe	Glu	Lys	Thr	Lys	Ala	Asp
465					470					475					480
Lys	Leu	Gln	Arg	Glu	Leu	Glu	Asp	Thr	Arg	Val	Ala	Thr	Val	Ser	Glu
			485						490					495	
Lys	Ser	Arg	Ile	Met	Glu	Leu	Glu	Lys	Asp	Leu	Ala	Leu	Arg	Val	Gln
			500						505					510	
Glu	Val	Ala	Glu	Leu	Arg	Arg	Arg	Leu	Glu	Ser	Ser	Lys	Pro	Pro	Gly

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Asp Lys Leu Lys Ala Ala Gln Glu Ala Asn Arg Asp Leu Met Gln Asp
 930 935 940

Met Glu Glu Leu Lys Ser Gln Ala Asp Lys Ala Lys Ala Ala Gln Thr
 945 950 955 960

Ala Glu Asp Ala Met Gln Ile Met Glu Gln Met Thr Lys Glu Lys Thr
 965 970 975

Glu Thr Leu Ala Ser Leu Glu Asp Thr Lys Gln Thr Asn Ala Lys Leu
 980 985 990

Gln Ser Glu Leu Asp Thr Leu Lys Glu Asn Asn Leu Lys Thr Val Glu
 995 1000 1005

Glu Leu Asn Lys Ser Lys Glu Leu Leu Asn Glu Glu Asn Gln Lys
 1010 1015 1020

Met Glu Glu Phe Lys Lys Glu Ile Glu Thr Leu Lys Gln Ala Ala
 1025 1030 1035

Ala Gln Lys Ser Gln Gln Leu Ser Ala Leu Gln Glu Glu Asn Val
 1040 1045 1050

Lys Leu Ala Glu Glu Leu Gly Arg Thr Arg Asp Glu Val Thr Ser
 1055 1060 1065

His Gln Lys Leu Glu Glu Glu Arg Ser Val Leu Asn Asn Gln Leu
 1070 1075 1080

Leu Glu Met Lys Lys Ser Leu Pro Ser Asn Thr Leu Arg Glu Ser
 1085 1090 1095

Glu Tyr Arg Lys Asp Ala Asp Glu Glu Lys Ala Ser Leu Gln Lys
 1100 1105 1110

Ser Ile Ser Leu Thr Ser Ala Leu Leu Thr Glu Lys Asp Ala Glu
 1115 1120 1125

Leu Glu Lys Leu Arg Asn Glu Val Thr Val Leu Arg Gly Glu Asn
 1130 1135 1140

Ala Ser Ala Lys Ser Leu His Ser Val Val Gln Thr Leu Glu Ser
 1145 1150 1155

Asp Lys Val Lys Leu Glu Leu Lys Val Lys Asn Leu Glu Leu Gln
 1160 1165 1170

Leu Lys Glu Asn Lys Arg Gln Leu Ser Ser Ser Ser Gly Asn Thr
 1175 1180 1185

Asp Val Gln Thr Glu Glu Asp Glu Arg Ala Gln Glu Ser Gln Gln
 1190 1195 1200

Met Ile Asp Phe Leu Asn Ser Val Ile Val Asp Leu Gln Arg Lys
 1205 1210 1215

Asn Gln Asp Leu Lys Met Lys Val Glu Met Met Ser Glu Gly Ala
 1220 1225 1230

Leu Asn Gly Asn Gly Glu Asp Pro Asn Ser Tyr Asp Ser Asp Asp
 1235 1240 1245

Gln Glu Lys Gln Ser Lys Lys Lys Pro Arg Leu Phe Cys Asp Ile
 1250 1255 1260

Cys Asp Cys Phe Asp Leu His Asp Thr Glu Asp Cys Pro Thr Gln
 1265 1270 1275

Ala Gln Met Ser Glu Asp Pro Pro His Ser Thr His His Gly Ser
 1280 1285 1290

Arg Ser Glu Glu Arg Pro Tyr Cys Glu Ile Cys Glu Met Phe Gly
 1295 1300 1305

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His Trp Ala Thr Asn Cys Asn Asp Asp Glu Thr Phe
 1310 1315 1320

<210> SEQ ID NO 13
 <211> LENGTH: 1294
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 13

Met Ala Met Gly Asp Asp Lys Ser Phe Asp Asp Glu Glu Ser Val Asp
 1 5 10 15

Gly Asn Arg Pro Ser Ser Ala Ala Ser Ala Phe Lys Val Pro Ala Pro
 20 25 30

Lys Thr Ser Gly Asn Pro Ala Asn Ser Ala Arg Lys Pro Gly Ser Ala
 35 40 45

Gly Gly Pro Lys Val Gly Gly Ala Ser Lys Glu Gly Ala Gly Ala
 50 55 60

Val Asp Glu Asp Asp Phe Ile Lys Ala Phe Thr Asp Val Pro Ser Ile
 65 70 75 80

Gln Ile Tyr Ser Ser Arg Glu Leu Glu Glu Thr Leu Asn Lys Ile Arg
 85 90 95

Glu Ile Leu Ser Asp Asp Lys His Asp Trp Asp Gln Arg Ala Asn Ala
 100 105 110

Leu Lys Lys Ile Arg Ser Leu Leu Val Ala Gly Ala Ala Gln Tyr Asp
 115 120 125

Cys Phe Phe Gln His Leu Arg Leu Leu Asp Gly Ala Leu Lys Leu Ser
 130 135 140

Ala Lys Asp Leu Arg Ser Gln Val Val Arg Glu Ala Cys Ile Thr Val
 145 150 155 160

Ala His Leu Ser Thr Val Leu Gly Asn Lys Phe Asp His Gly Ala Glu
 165 170 175

Ala Ile Val Pro Thr Leu Phe Asn Leu Val Pro Asn Ser Ala Lys Val
 180 185 190

Met Ala Thr Ser Gly Cys Ala Ala Ile Arg Phe Ile Ile Arg His Thr
 195 200 205

His Val Pro Arg Leu Ile Pro Leu Ile Thr Ser Asn Cys Thr Ser Lys
 210 215 220

Ser Val Pro Val Arg Arg Ser Phe Glu Phe Leu Asp Leu Leu Leu
 225 230 235 240

Gln Glu Trp Gln Thr His Ser Leu Glu Arg His Ala Ala Val Leu Val
 245 250 255

Glu Thr Ile Lys Lys Gly Ile His Asp Ala Asp Ala Glu Ala Arg Val
 260 265 270

Glu Ala Arg Lys Thr Tyr Met Gly Leu Arg Asn His Phe Pro Gly Glu
 275 280 285

Ala Glu Thr Leu Tyr Asn Ser Leu Glu Pro Ser Tyr Gln Lys Ser Leu
 290 295 300

Gln Thr Tyr Leu Lys Ser Ser Gly Ser Val Ala Ser Leu Pro Gln Ser
 305 310 315 320

Asp Arg Ser Ser Ser Ser Ser Gln Glu Ser Leu Asn Arg Pro Phe Ser
 325 330 335

Ser Lys Trp Ser Thr Ala Asn Pro Ser Thr Val Ala Gly Arg Val Ser
 340 345 350

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Ala Gly Ser Ser Lys Ala Ser Ser Leu Pro Gly Ser Leu Gln Arg Ser
 355 360 365

Arg Ser Asp Ile Asp Val Asn Ala Ala Ala Gly Ala Lys Ala His His
 370 375 380

Ala Ala Gly Gln Ser Val Arg Ser Gly Arg Leu Gly Ala Gly Ala Leu
 385 390 395 400

Asn Ala Gly Ser Tyr Ala Ser Leu Glu Asp Thr Ser Asp Lys Leu Asp
 405 410 415

Gly Thr Ala Ser Glu Asp Gly Arg Val Arg Ala Lys Leu Ser Ala Pro
 420 425 430

Leu Ala Gly Met Gly Asn Ala Lys Ala Asp Ser Arg Gly Arg Ser Arg
 435 440 445

Thr Lys Met Val Ser Gln Ser Gln Pro Gly Ser Arg Ser Gly Ser Pro
 450 455 460

Gly Arg Val Leu Thr Thr Thr Ala Leu Ser Thr Val Ser Ser Gly Val
 465 470 475 480

Gln Arg Val Leu Val Asn Ser Ala Ser Ala Gln Lys Arg Ser Lys Ile
 485 490 495

Pro Arg Ser Gln Gly Cys Ser Arg Glu Ala Ser Pro Ser Arg Leu Ser
 500 505 510

Val Ala Arg Ser Ser Arg Ile Pro Arg Pro Ser Val Ser Gln Gly Cys
 515 520 525

Ser Arg Glu Ala Ser Arg Glu Ser Ser Arg Asp Thr Ser Pro Val Arg
 530 535 540

Ser Phe Gln Pro Leu Ala Ser Arg His His Ser Arg Ser Thr Gly Ala
 545 550 555 560

Leu Tyr Ala Pro Glu Val Tyr Gly Ala Ser Gly Pro Gly Tyr Gly Ile
 565 570 575

Ser Gln Ser Ser Arg Leu Ser Ser Ser Val Ser Ala Met Arg Val Leu
 580 585 590

Asn Thr Gly Ser Asp Val Glu Glu Ala Val Ala Asp Ala Leu Lys Lys
 595 600 605

Pro Ala Arg Arg Arg Tyr Glu Ser Tyr Gly Met His Ser Asp Asp Asp
 610 615 620

Ala Asn Ser Asp Ala Ser Ser Ala Cys Ser Glu Arg Ser Tyr Ser Ser
 625 630 635 640

Arg Asn Gly Ser Ile Pro Thr Tyr Met Arg Gln Thr Glu Asp Val Ala
 645 650 655

Glu Val Leu Asn Arg Cys Ala Ser Ser Asn Trp Ser Glu Arg Lys Glu
 660 665 670

Gly Leu Leu Gly Leu Gln Asn Leu Leu Lys Asn Gln Arg Thr Leu Ser
 675 680 685

Arg Val Glu Leu Lys Arg Leu Cys Glu Ile Phe Thr Arg Met Phe Ala
 690 695 700

Asp Pro His Gly Lys Arg Val Phe Ser Met Phe Leu Glu Thr Leu Val
 705 710 715 720

Asp Phe Ile Gln Val His Lys Asp Asp Leu Gln Asp Trp Leu Phe Val
 725 730 735

Leu Leu Thr Gln Leu Leu Lys Lys Met Gly Ala Asp Leu Leu Gly Ser
 740 745 750

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Val	Gln	Ala	Lys	Val	Gln	Lys	Ala	Leu	Asp	Val	Thr	Arg	Glu	Ser	Phe
	755						760					765			
Pro	Asn	Asp	Leu	Gln	Phe	Asn	Ile	Leu	Met	Arg	Phe	Thr	Val	Asp	Gln
	770					775					780				
Thr	Gln	Thr	Pro	Ser	Leu	Lys	Val	Lys	Val	Ala	Ile	Leu	Lys	Tyr	Ile
	785				790					795					800
Glu	Thr	Leu	Ala	Lys	Gln	Met	Asp	Pro	Gly	Asp	Phe	Ile	Asn	Ser	Ser
			805						810					815	
Glu	Thr	Arg	Leu	Ala	Val	Ser	Arg	Val	Ile	Thr	Trp	Thr	Thr	Glu	Pro
			820					825						830	
Lys	Ser	Ser	Asp	Val	Arg	Lys	Ala	Ala	Gln	Ser	Val	Leu	Ile	Ser	Leu
		835					840					845			
Phe	Glu	Leu	Asn	Thr	Pro	Glu	Phe	Thr	Met	Leu	Leu	Gly	Ala	Leu	Pro
	850					855						860			
Lys	Thr	Phe	Gln	Asp	Gly	Ala	Thr	Lys	Leu	Leu	His	Asn	His	Leu	Arg
	865				870					875					880
Asn	Thr	Gly	Asn	Gly	Thr	Gln	Ser	Ser	Met	Gly	Ser	Pro	Leu	Thr	Arg
			885						890					895	
Pro	Thr	Pro	Arg	Ser	Pro	Ala	Asn	Trp	Ser	Ser	Pro	Leu	Thr	Ser	Pro
			900					905					910		
Thr	Asn	Thr	Ser	Gln	Asn	Thr	Leu	Ser	Pro	Ser	Ala	Phe	Asp	Tyr	Asp
	915					920						925			
Thr	Glu	Asn	Met	Asn	Ser	Glu	Asp	Ile	Tyr	Ser	Ser	Leu	Arg	Gly	Val
	930					935						940			
Thr	Glu	Ala	Ile	Gln	Asn	Phe	Ser	Phe	Arg	Ser	Gln	Glu	Asp	Met	Asn
	945				950					955					960
Glu	Pro	Leu	Lys	Arg	Asp	Ser	Lys	Lys	Asp	Asp	Gly	Asp	Ser	Met	Cys
			965						970					975	
Gly	Gly	Pro	Gly	Met	Ser	Asp	Pro	Arg	Ala	Gly	Gly	Asp	Ala	Thr	Asp
		980						985					990		
Ser	Ser	Gln	Thr	Ala	Leu	Asp	Asn	Lys	Ala	Ser	Leu	Leu	His	Ser	Met
		995					1000						1005		
Pro	Thr	His	Ser	Ser	Pro	Arg	Ser	Arg	Asp	Tyr	Asn	Pro	Tyr	Asn	
	1010					1015					1020				
Tyr	Ser	Asp	Ser	Ile	Ser	Pro	Phe	Asn	Lys	Ser	Ala	Leu	Lys	Glu	
	1025					1030					1035				
Ala	Met	Phe	Asp	Asp	Asp	Ala	Asp	Gln	Phe	Pro	Asp	Asp	Leu	Ser	
	1040					1045					1050				
Leu	Asp	His	Ser	Asp	Leu	Val	Ala	Glu	Leu	Leu	Lys	Glu	Leu	Ser	
	1055					1060					1065				
Asn	His	Asn	Glu	Arg	Val	Glu	Glu	Arg	Lys	Ile	Ala	Leu	Tyr	Glu	
	1070					1075					1080				
Leu	Met	Lys	Leu	Thr	Gln	Glu	Glu	Ser	Phe	Ser	Val	Trp	Asp	Glu	
	1085					1090					1095				
His	Phe	Lys	Thr	Ile	Leu	Leu	Leu	Leu	Leu	Glu	Thr	Leu	Gly	Asp	
	1100					1105					1110				
Lys	Glu	Pro	Thr	Ile	Arg	Ala	Leu	Ala	Leu	Lys	Val	Leu	Arg	Glu	
	1115					1120					1125				
Ile	Leu	Arg	His	Gln	Pro	Ala	Arg	Phe	Lys	Asn	Tyr	Ala	Glu	Leu	
	1130					1135					1140				
Thr	Val	Met	Lys	Thr	Leu	Glu	Ala	His	Lys	Asp	Pro	His	Lys	Glu	

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1145          1150          1155
Val Val Arg Ser Ala Glu Glu Ala Ala Ser Val Leu Ala Thr Ser
1160          1165          1170

Ile Ser Pro Glu Gln Cys Ile Lys Val Leu Cys Pro Ile Ile Gln
1175          1180          1185

Thr Ala Asp Tyr Pro Ile Asn Leu Ala Ala Ile Lys Met Gln Thr
1190          1195          1200

Lys Val Ile Glu Arg Val Ser Lys Glu Thr Leu Asn Leu Leu Leu
1205          1210          1215

Pro Glu Ile Met Pro Gly Leu Ile Gln Gly Tyr Asp Asn Ser Glu
1220          1225          1230

Ser Ser Val Arg Lys Ala Cys Val Phe Cys Leu Val Ala Val His
1235          1240          1245

Ala Val Ile Gly Asp Glu Leu Lys Pro His Leu Ser Gln Leu Thr
1250          1255          1260

Gly Ser Lys Met Lys Leu Leu Asn Leu Tyr Ile Lys Arg Ala Gln
1265          1270          1275

Thr Gly Ser Gly Gly Ala Asp Pro Thr Thr Asp Val Ser Gly Gln
1280          1285          1290

Ser

<210> SEQ ID NO 14
<211> LENGTH: 2843
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 14
Met Ala Ala Ala Ser Tyr Asp Gln Leu Leu Lys Gln Val Glu Ala Leu
1          5          10          15

Lys Met Glu Asn Ser Asn Leu Arg Gln Glu Leu Glu Asp Asn Ser Asn
20          25          30

His Leu Thr Lys Leu Glu Thr Glu Ala Ser Asn Met Lys Glu Val Leu
35          40          45

Lys Gln Leu Gln Gly Ser Ile Glu Asp Glu Ala Met Ala Ser Ser Gly
50          55          60

Gln Ile Asp Leu Leu Glu Arg Leu Lys Glu Leu Asn Leu Asp Ser Ser
65          70          75          80

Asn Phe Pro Gly Val Lys Leu Arg Ser Lys Met Ser Leu Arg Ser Tyr
85          90          95

Gly Ser Arg Glu Gly Ser Val Ser Ser Arg Ser Gly Glu Cys Ser Pro
100         105         110

Val Pro Met Gly Ser Phe Pro Arg Arg Gly Phe Val Asn Gly Ser Arg
115         120         125

Glu Ser Thr Gly Tyr Leu Glu Glu Leu Glu Lys Glu Arg Ser Leu Leu
130         135         140

Leu Ala Asp Leu Asp Lys Glu Glu Lys Glu Lys Asp Trp Tyr Tyr Ala
145         150         155         160

Gln Leu Gln Asn Leu Thr Lys Arg Ile Asp Ser Leu Pro Leu Thr Glu
165         170         175

Asn Phe Ser Leu Gln Thr Asp Met Thr Arg Arg Gln Leu Glu Tyr Glu
180         185         190

Ala Arg Gln Ile Arg Val Ala Met Glu Glu Gln Leu Gly Thr Cys Gln

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195				200				205							
Asp	Met	Glu	Lys	Arg	Ala	Gln	Arg	Arg	Ile	Ala	Arg	Ile	Gln	Gln	Ile
210						215					220				
Glu	Lys	Asp	Ile	Leu	Arg	Ile	Arg	Gln	Leu	Leu	Gln	Ser	Gln	Ala	Thr
225					230					235					240
Glu	Ala	Glu	Arg	Ser	Ser	Gln	Asn	Lys	His	Glu	Thr	Gly	Ser	His	Asp
				245					250					255	
Ala	Glu	Arg	Gln	Asn	Glu	Gly	Gln	Gly	Val	Gly	Glu	Ile	Asn	Met	Ala
			260						265					270	
Thr	Ser	Gly	Asn	Gly	Gln	Gly	Ser	Thr	Thr	Arg	Met	Asp	His	Glu	Thr
		275					280					285			
Ala	Ser	Val	Leu	Ser	Ser	Ser	Ser	Thr	His	Ser	Ala	Pro	Arg	Arg	Leu
						295					300				
Thr	Ser	His	Leu	Gly	Thr	Lys	Val	Glu	Met	Val	Tyr	Ser	Leu	Leu	Ser
305					310					315					320
Met	Leu	Gly	Thr	His	Asp	Lys	Asp	Asp	Met	Ser	Arg	Thr	Leu	Leu	Ala
				325					330					335	
Met	Ser	Ser	Ser	Gln	Asp	Ser	Cys	Ile	Ser	Met	Arg	Gln	Ser	Gly	Cys
				340					345					350	
Leu	Pro	Leu	Leu	Ile	Gln	Leu	Leu	His	Gly	Asn	Asp	Lys	Asp	Ser	Val
		355					360					365			
Leu	Leu	Gly	Asn	Ser	Arg	Gly	Ser	Lys	Glu	Ala	Arg	Ala	Arg	Ala	Ser
		370				375					380				
Ala	Ala	Leu	His	Asn	Ile	Ile	His	Ser	Gln	Pro	Asp	Asp	Lys	Arg	Gly
385					390					395					400
Arg	Arg	Glu	Ile	Arg	Val	Leu	His	Leu	Leu	Glu	Gln	Ile	Arg	Ala	Tyr
				405					410					415	
Cys	Glu	Thr	Cys	Trp	Glu	Trp	Gln	Glu	Ala	His	Glu	Pro	Gly	Met	Asp
				420					425				430		
Gln	Asp	Lys	Asn	Pro	Met	Pro	Ala	Pro	Val	Glu	His	Gln	Ile	Cys	Pro
		435					440						445		
Ala	Val	Cys	Val	Leu	Met	Lys	Leu	Ser	Phe	Asp	Glu	Glu	His	Arg	His
				450		455				460					
Ala	Met	Asn	Glu	Leu	Gly	Gly	Leu	Gln	Ala	Ile	Ala	Glu	Leu	Leu	Gln
465					470					475					480
Val	Asp	Cys	Glu	Met	Tyr	Gly	Leu	Thr	Asn	Asp	His	Tyr	Ser	Ile	Thr
				485					490					495	
Leu	Arg	Arg	Tyr	Ala	Gly	Met	Ala	Leu	Thr	Asn	Leu	Thr	Phe	Gly	Asp
				500					505				510		
Val	Ala	Asn	Lys	Ala	Thr	Leu	Cys	Ser	Met	Lys	Gly	Cys	Met	Arg	Ala
		515					520					525			
Leu	Val	Ala	Gln	Leu	Lys	Ser	Glu	Ser	Glu	Asp	Leu	Gln	Gln	Val	Ile
				530			535					540			
Ala	Ser	Val	Leu	Arg	Asn	Leu	Ser	Trp	Arg	Ala	Asp	Val	Asn	Ser	Lys
545					550					555					560
Lys	Thr	Leu	Arg	Glu	Val	Gly	Ser	Val	Lys	Ala	Leu	Met	Glu	Cys	Ala
				565					570					575	
Leu	Glu	Val	Lys	Lys	Glu	Ser	Thr	Leu	Lys	Ser	Val	Leu	Ser	Ala	Leu
				580					585				590		
Trp	Asn	Leu	Ser	Ala	His	Cys	Thr	Glu	Asn	Lys	Ala	Asp	Ile	Cys	Ala
		595					600						605		

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Val Asp Gly Ala Leu Ala Phe Leu Val Gly Thr Leu Thr Tyr Arg Ser
 610 615 620
 Gln Thr Asn Thr Leu Ala Ile Ile Glu Ser Gly Gly Gly Ile Leu Arg
 625 630 635 640
 Asn Val Ser Ser Leu Ile Ala Thr Asn Glu Asp His Arg Gln Ile Leu
 645 650 655
 Arg Glu Asn Asn Cys Leu Gln Thr Leu Leu Gln His Leu Lys Ser His
 660 665 670
 Ser Leu Thr Ile Val Ser Asn Ala Cys Gly Thr Leu Trp Asn Leu Ser
 675 680 685
 Ala Arg Asn Pro Lys Asp Gln Glu Ala Leu Trp Asp Met Gly Ala Val
 690 695 700
 Ser Met Leu Lys Asn Leu Ile His Ser Lys His Lys Met Ile Ala Met
 705 710 715 720
 Gly Ser Ala Ala Ala Leu Arg Asn Leu Met Ala Asn Arg Pro Ala Lys
 725 730 735
 Tyr Lys Asp Ala Asn Ile Met Ser Pro Gly Ser Ser Leu Pro Ser Leu
 740 745 750
 His Val Arg Lys Gln Lys Ala Leu Glu Ala Glu Leu Asp Ala Gln His
 755 760 765
 Leu Ser Glu Thr Phe Asp Asn Ile Asp Asn Leu Ser Pro Lys Ala Ser
 770 775 780
 His Arg Ser Lys Gln Arg His Lys Gln Ser Leu Tyr Gly Asp Tyr Val
 785 790 795 800
 Phe Asp Thr Asn Arg His Asp Asp Asn Arg Ser Asp Asn Phe Asn Thr
 805 810 815
 Gly Asn Met Thr Val Leu Ser Pro Tyr Leu Asn Thr Thr Val Leu Pro
 820 825 830
 Ser Ser Ser Ser Ser Arg Gly Ser Leu Asp Ser Ser Arg Ser Glu Lys
 835 840 845
 Asp Arg Ser Leu Glu Arg Glu Arg Gly Ile Gly Leu Gly Asn Tyr His
 850 855 860
 Pro Ala Thr Glu Asn Pro Gly Thr Ser Ser Lys Arg Gly Leu Gln Ile
 865 870 875 880
 Ser Thr Thr Ala Ala Gln Ile Ala Lys Val Met Glu Glu Val Ser Ala
 885 890 895
 Ile His Thr Ser Gln Glu Asp Arg Ser Ser Gly Ser Thr Thr Glu Leu
 900 905 910
 His Cys Val Thr Asp Glu Arg Asn Ala Leu Arg Arg Ser Ser Ala Ala
 915 920 925
 His Thr His Ser Asn Thr Tyr Asn Phe Thr Lys Ser Glu Asn Ser Asn
 930 935 940
 Arg Thr Cys Ser Met Pro Tyr Ala Lys Leu Glu Tyr Lys Arg Ser Ser
 945 950 955 960
 Asn Asp Ser Leu Asn Ser Val Ser Ser Ser Asp Gly Tyr Gly Lys Arg
 965 970 975
 Gly Gln Met Lys Pro Ser Ile Glu Ser Tyr Ser Glu Asp Asp Glu Ser
 980 985 990
 Lys Phe Cys Ser Tyr Gly Gln Tyr Pro Ala Asp Leu Ala His Lys Ile
 995 1000 1005

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His	Ser	Ala	Asn	His	Met	Asp	Asp	Asn	Asp	Gly	Glu	Leu	Asp	Thr
1010						1015					1020			
Pro	Ile	Asn	Tyr	Ser	Leu	Lys	Tyr	Ser	Asp	Glu	Gln	Leu	Asn	Ser
1025						1030					1035			
Gly	Arg	Gln	Ser	Pro	Ser	Gln	Asn	Glu	Arg	Trp	Ala	Arg	Pro	Lys
1040						1045					1050			
His	Ile	Ile	Glu	Asp	Glu	Ile	Lys	Gln	Ser	Glu	Gln	Arg	Gln	Ser
1055						1060					1065			
Arg	Asn	Gln	Ser	Thr	Thr	Tyr	Pro	Val	Tyr	Thr	Glu	Ser	Thr	Asp
1070						1075					1080			
Asp	Lys	His	Leu	Lys	Phe	Gln	Pro	His	Phe	Gly	Gln	Gln	Glu	Cys
1085						1090					1095			
Val	Ser	Pro	Tyr	Arg	Ser	Arg	Gly	Ala	Asn	Gly	Ser	Glu	Thr	Asn
1100						1105					1110			
Arg	Val	Gly	Ser	Asn	His	Gly	Ile	Asn	Gln	Asn	Val	Ser	Gln	Ser
1115						1120					1125			
Leu	Cys	Gln	Glu	Asp	Asp	Tyr	Glu	Asp	Asp	Lys	Pro	Thr	Asn	Tyr
1130						1135					1140			
Ser	Glu	Arg	Tyr	Ser	Glu	Glu	Glu	Gln	His	Glu	Glu	Glu	Glu	Arg
1145						1150					1155			
Pro	Thr	Asn	Tyr	Ser	Ile	Lys	Tyr	Asn	Glu	Glu	Lys	Arg	His	Val
1160						1165					1170			
Asp	Gln	Pro	Ile	Asp	Tyr	Ser	Leu	Lys	Tyr	Ala	Thr	Asp	Ile	Pro
1175						1180					1185			
Ser	Ser	Gln	Lys	Gln	Ser	Phe	Ser	Phe	Ser	Lys	Ser	Ser	Ser	Gly
1190						1195					1200			
Gln	Ser	Ser	Lys	Thr	Glu	His	Met	Ser	Ser	Ser	Ser	Glu	Asn	Thr
1205						1210					1215			
Ser	Thr	Pro	Ser	Ser	Asn	Ala	Lys	Arg	Gln	Asn	Gln	Leu	His	Pro
1220						1225					1230			
Ser	Ser	Ala	Gln	Ser	Arg	Ser	Gly	Gln	Pro	Gln	Lys	Ala	Ala	Thr
1235						1240					1245			
Cys	Lys	Val	Ser	Ser	Ile	Asn	Gln	Glu	Thr	Ile	Gln	Thr	Tyr	Cys
1250						1255					1260			
Val	Glu	Asp	Thr	Pro	Ile	Cys	Phe	Ser	Arg	Cys	Ser	Ser	Leu	Ser
1265						1270					1275			
Ser	Leu	Ser	Ser	Ala	Glu	Asp	Glu	Ile	Gly	Cys	Asn	Gln	Thr	Thr
1280						1285					1290			
Gln	Glu	Ala	Asp	Ser	Ala	Asn	Thr	Leu	Gln	Ile	Ala	Glu	Ile	Lys
1295						1300					1305			
Glu	Lys	Ile	Gly	Thr	Arg	Ser	Ala	Glu	Asp	Pro	Val	Ser	Glu	Val
1310						1315					1320			
Pro	Ala	Val	Ser	Gln	His	Pro	Arg	Thr	Lys	Ser	Ser	Arg	Leu	Gln
1325						1330					1335			
Gly	Ser	Ser	Leu	Ser	Ser	Glu	Ser	Ala	Arg	His	Lys	Ala	Val	Glu
1340						1345					1350			
Phe	Ser	Ser	Gly	Ala	Lys	Ser	Pro	Ser	Lys	Ser	Gly	Ala	Gln	Thr
1355						1360					1365			
Pro	Lys	Ser	Pro	Pro	Glu	His	Tyr	Val	Gln	Glu	Thr	Pro	Leu	Met
1370						1375					1380			
Phe	Ser	Arg	Cys	Thr	Ser	Val	Ser	Ser	Leu	Asp	Ser	Phe	Glu	Ser

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Ser	Pro	Val	Lys	Pro	Ile	Pro	Gln	Asn	Thr	Glu	Tyr	Arg	Thr	Arg
1775						1780						1785		
Val	Arg	Lys	Asn	Ala	Asp	Ser	Lys	Asn	Asn	Leu	Asn	Ala	Glu	Arg
1790						1795						1800		
Val	Phe	Ser	Asp	Asn	Lys	Asp	Ser	Lys	Lys	Gln	Asn	Leu	Lys	Asn
1805						1810						1815		
Asn	Ser	Lys	Val	Phe	Asn	Asp	Lys	Leu	Pro	Asn	Asn	Glu	Asp	Arg
1820						1825						1830		
Val	Arg	Gly	Ser	Phe	Ala	Phe	Asp	Ser	Pro	His	His	Tyr	Thr	Pro
1835						1840						1845		
Ile	Glu	Gly	Thr	Pro	Tyr	Cys	Phe	Ser	Arg	Asn	Asp	Ser	Leu	Ser
1850						1855						1860		
Ser	Leu	Asp	Phe	Asp	Asp	Asp	Asp	Val	Asp	Leu	Ser	Arg	Glu	Lys
1865						1870						1875		
Ala	Glu	Leu	Arg	Lys	Ala	Lys	Glu	Asn	Lys	Glu	Ser	Glu	Ala	Lys
1880						1885						1890		
Val	Thr	Ser	His	Thr	Glu	Leu	Thr	Ser	Asn	Gln	Gln	Ser	Ala	Asn
1895						1900						1905		
Lys	Thr	Gln	Ala	Ile	Ala	Lys	Gln	Pro	Ile	Asn	Arg	Gly	Gln	Pro
1910						1915						1920		
Lys	Pro	Ile	Leu	Gln	Lys	Gln	Ser	Thr	Phe	Pro	Gln	Ser	Ser	Lys
1925						1930						1935		
Asp	Ile	Pro	Asp	Arg	Gly	Ala	Ala	Thr	Asp	Glu	Lys	Leu	Gln	Asn
1940						1945						1950		
Phe	Ala	Ile	Glu	Asn	Thr	Pro	Val	Cys	Phe	Ser	His	Asn	Ser	Ser
1955						1960						1965		
Leu	Ser	Ser	Leu	Ser	Asp	Ile	Asp	Gln	Glu	Asn	Asn	Asn	Lys	Glu
1970						1975						1980		
Asn	Glu	Pro	Ile	Lys	Glu	Thr	Glu	Pro	Pro	Asp	Ser	Gln	Gly	Glu
1985						1990						1995		
Pro	Ser	Lys	Pro	Gln	Ala	Ser	Gly	Tyr	Ala	Pro	Lys	Ser	Phe	His
2000						2005						2010		
Val	Glu	Asp	Thr	Pro	Val	Cys	Phe	Ser	Arg	Asn	Ser	Ser	Leu	Ser
2015						2020						2025		
Ser	Leu	Ser	Ile	Asp	Ser	Glu	Asp	Asp	Leu	Leu	Gln	Glu	Cys	Ile
2030						2035						2040		
Ser	Ser	Ala	Met	Pro	Lys	Lys	Lys	Lys	Pro	Ser	Arg	Leu	Lys	Gly
2045						2050						2055		
Asp	Asn	Glu	Lys	His	Ser	Pro	Arg	Asn	Met	Gly	Gly	Ile	Leu	Gly
2060						2065						2070		
Glu	Asp	Leu	Thr	Leu	Asp	Leu	Lys	Asp	Ile	Gln	Arg	Pro	Asp	Ser
2075						2080						2085		
Glu	His	Gly	Leu	Ser	Pro	Asp	Ser	Glu	Asn	Phe	Asp	Trp	Lys	Ala
2090						2095						2100		
Ile	Gln	Glu	Gly	Ala	Asn	Ser	Ile	Val	Ser	Ser	Leu	His	Gln	Ala
2105						2110						2115		
Ala	Ala	Ala	Ala	Cys	Leu	Ser	Arg	Gln	Ala	Ser	Ser	Asp	Ser	Asp
2120						2125						2130		
Ser	Ile	Leu	Ser	Leu	Lys	Ser	Gly	Ile	Ser	Leu	Gly	Ser	Pro	Phe
2135						2140						2145		

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His	Leu	Thr	Pro	Asp	Gln	Glu	Glu	Lys	Pro	Phe	Thr	Ser	Asn	Lys
	2150					2155					2160			
Gly	Pro	Arg	Ile	Leu	Lys	Pro	Gly	Glu	Lys	Ser	Thr	Leu	Glu	Thr
	2165					2170					2175			
Lys	Lys	Ile	Glu	Ser	Glu	Ser	Lys	Gly	Ile	Lys	Gly	Gly	Lys	Lys
	2180					2185					2190			
Val	Tyr	Lys	Ser	Leu	Ile	Thr	Gly	Lys	Val	Arg	Ser	Asn	Ser	Glu
	2195					2200					2205			
Ile	Ser	Gly	Gln	Met	Lys	Gln	Pro	Leu	Gln	Ala	Asn	Met	Pro	Ser
	2210					2215					2220			
Ile	Ser	Arg	Gly	Arg	Thr	Met	Ile	His	Ile	Pro	Gly	Val	Arg	Asn
	2225					2230					2235			
Ser	Ser	Ser	Ser	Thr	Ser	Pro	Val	Ser	Lys	Lys	Gly	Pro	Pro	Leu
	2240					2245					2250			
Lys	Thr	Pro	Ala	Ser	Lys	Ser	Pro	Ser	Glu	Gly	Gln	Thr	Ala	Thr
	2255					2260					2265			
Thr	Ser	Pro	Arg	Gly	Ala	Lys	Pro	Ser	Val	Lys	Ser	Glu	Leu	Ser
	2270					2275					2280			
Pro	Val	Ala	Arg	Gln	Thr	Ser	Gln	Ile	Gly	Gly	Ser	Ser	Lys	Ala
	2285					2290					2295			
Pro	Ser	Arg	Ser	Gly	Ser	Arg	Asp	Ser	Thr	Pro	Ser	Arg	Pro	Ala
	2300					2305					2310			
Gln	Gln	Pro	Leu	Ser	Arg	Pro	Ile	Gln	Ser	Pro	Gly	Arg	Asn	Ser
	2315					2320					2325			
Ile	Ser	Pro	Gly	Arg	Asn	Gly	Ile	Ser	Pro	Pro	Asn	Lys	Leu	Ser
	2330					2335					2340			
Gln	Leu	Pro	Arg	Thr	Ser	Ser	Pro	Ser	Thr	Ala	Ser	Thr	Lys	Ser
	2345					2350					2355			
Ser	Gly	Ser	Gly	Lys	Met	Ser	Tyr	Thr	Ser	Pro	Gly	Arg	Gln	Met
	2360					2365					2370			
Ser	Gln	Gln	Asn	Leu	Thr	Lys	Gln	Thr	Gly	Leu	Ser	Lys	Asn	Ala
	2375					2380					2385			
Ser	Ser	Ile	Pro	Arg	Ser	Glu	Ser	Ala	Ser	Lys	Gly	Leu	Asn	Gln
	2390					2395					2400			
Met	Asn	Asn	Gly	Asn	Gly	Ala	Asn	Lys	Lys	Val	Glu	Leu	Ser	Arg
	2405					2410					2415			
Met	Ser	Ser	Thr	Lys	Ser	Ser	Gly	Ser	Glu	Ser	Asp	Arg	Ser	Glu
	2420					2425					2430			
Arg	Pro	Val	Leu	Val	Arg	Gln	Ser	Thr	Phe	Ile	Lys	Glu	Ala	Pro
	2435					2440					2445			
Ser	Pro	Thr	Leu	Arg	Arg	Lys	Leu	Glu	Glu	Ser	Ala	Ser	Phe	Glu
	2450					2455					2460			
Ser	Leu	Ser	Pro	Ser	Ser	Arg	Pro	Ala	Ser	Pro	Thr	Arg	Ser	Gln
	2465					2470					2475			
Ala	Gln	Thr	Pro	Val	Leu	Ser	Pro	Ser	Leu	Pro	Asp	Met	Ser	Leu
	2480					2485					2490			
Ser	Thr	His	Ser	Ser	Val	Gln	Ala	Gly	Gly	Trp	Arg	Lys	Leu	Pro
	2495					2500					2505			
Pro	Asn	Leu	Ser	Pro	Thr	Ile	Glu	Tyr	Asn	Asp	Gly	Arg	Pro	Ala
	2510					2515					2520			
Lys	Arg	His	Asp	Ile	Ala	Arg	Ser	His	Ser	Glu	Ser	Pro	Ser	Arg

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His Ile Ser Lys Arg Pro Val Phe Leu Ser Glu Glu Thr Pro Tyr Ser
 20 25 30
 Tyr Pro Thr Gly Asn His Thr Tyr Gln Glu Ile Ala Val Pro Pro Pro
 35 40 45
 Val Pro Pro Pro Pro Ser Ser Glu Ala Leu Asn Gly Thr Ile Leu Asp
 50 55 60
 Pro Leu Asp Gln Trp Gln Pro Ser Gly Ser Arg Phe Ile His Gln Gln
 65 70 75 80
 Pro Gln Ser Ser Ser Pro Val Tyr Gly Ser Ser Ala Lys Thr Ser Ser
 85 90
 Val Ser Asn Pro Gln Asp Ser Val Gly Ser Pro Cys Ser Arg Val Gly
 100 105 110
 Glu Glu Glu His Val Tyr Ser Phe Pro Asn Lys Gln Lys Ser Ala Glu
 115 120 125
 Pro Ser Pro Thr Val Met Ser Thr Ser Leu Gly Ser Asn Leu Ser Glu
 130 135 140
 Leu Asp Arg Leu Leu Leu Glu Leu Asn Ala Val Gln His Asn Pro Pro
 145 150 155 160
 Gly Phe Pro Ala Asp Glu Ala Asn Ser Ser Pro Pro Leu Pro Gly Ala
 165 170 175
 Leu Ser Pro Leu Tyr Gly Val Pro Glu Thr Asn Ser Pro Leu Gly Gly
 180 185 190
 Lys Ala Gly Pro Leu Thr Lys Glu Lys Pro Lys Arg Asn Gly Gly Arg
 195 200 205
 Gly Leu Glu Asp Val Arg Pro Ser Val Glu Ser Leu Leu Asp Glu Leu
 210 215 220
 Glu Ser Ser Val Pro Ser Pro Val Pro Ala Ile Thr Val Asn Gln Gly
 225 230 235 240
 Glu Met Ser Ser Pro Gln Arg Val Thr Ser Thr Gln Gln Gln Thr Arg
 245 250 255
 Ile Ser Ala Ser Ser Ala Thr Arg Glu Leu Asp Glu Leu Met Ala Ser
 260 265 270
 Leu Ser Asp Phe Lys Phe Met Ala Gln Gly Lys Thr Gly Ser Ser Ser
 275 280 285
 Pro Pro Gly Gly Pro Pro Lys Pro Gly Ser Gln Leu Asp Ser Met Leu
 290 295 300
 Gly Ser Leu Gln Ser Asp Leu Asn Lys Leu Gly Val Ala Thr Val Ala
 305 310 315 320
 Lys Gly Val Cys Gly Ala Cys Lys Lys Pro Ile Ala Gly Gln Val Val
 325 330 335
 Thr Ala Met Gly Lys Thr Trp His Pro Glu His Phe Val Cys Thr His
 340 345 350
 Cys Gln Glu Glu Ile Gly Ser Arg Asn Phe Phe Glu Arg Asp Gly Gln
 355 360 365
 Pro Tyr Cys Glu Lys Asp Tyr His Asn Leu Phe Ser Pro Arg Cys Tyr
 370 375 380
 Tyr Cys Asn Gly Pro Ile Leu Asp Lys Val Val Thr Ala Leu Asp Arg
 385 390 395 400
 Thr Trp His Pro Glu His Phe Phe Cys Ala Gln Cys Gly Ala Phe Phe
 405 410 415
 Gly Pro Glu Gly Phe His Glu Lys Asp Gly Lys Ala Tyr Cys Arg Lys

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                420                425                430
Asp Tyr Phe Asp Met Phe Ala Pro Lys Cys Gly Gly Cys Ala Arg Ala
    435                440                445

Ile Leu Glu Asn Tyr Ile Ser Ala Leu Asn Thr Leu Trp His Pro Glu
    450                455                460

Cys Phe Val Cys Arg Glu Cys Phe Thr Pro Phe Val Asn Gly Ser Phe
    465                470                475                480

Phe Glu His Asp Gly Gln Pro Tyr Cys Glu Val His Tyr His Glu Arg
    485                490                495

Arg Gly Ser Leu Cys Ser Gly Cys Gln Lys Pro Ile Thr Gly Arg Cys
    500                505                510

Ile Thr Ala Met Ala Lys Lys Phe His Pro Glu His Phe Val Cys Ala
    515                520                525

Phe Cys Leu Lys Gln Leu Asn Lys Gly Thr Phe Lys Glu Gln Asn Asp
    530                535                540

Lys Pro Tyr Cys Gln Asn Cys Phe Leu Lys Leu Phe Cys
    545                550                555

<210> SEQ ID NO 16
<211> LENGTH: 533
<212> TYPE: PRT
<213> ORGANISM: Gallus gallus domesticus

<400> SEQUENCE: 16

Met Gly Ser Ser Lys Ser Lys Pro Lys Asp Pro Ser Gln Arg Arg Arg
 1                5                10                15

Ser Leu Glu Pro Pro Asp Ser Thr His His Gly Gly Phe Pro Ala Ser
 20                25                30

Gln Thr Pro Asn Lys Thr Ala Ala Pro Asp Thr His Arg Thr Pro Ser
 35                40                45

Arg Ser Phe Gly Thr Val Ala Thr Glu Pro Lys Leu Phe Gly Gly Phe
 50                55                60

Asn Thr Ser Asp Thr Val Thr Ser Pro Gln Arg Ala Gly Ala Leu Ala
 65                70                75                80

Gly Gly Val Thr Thr Phe Val Ala Leu Tyr Asp Tyr Glu Ser Arg Thr
 85                90                95

Glu Thr Asp Leu Ser Phe Lys Lys Gly Glu Arg Leu Gln Ile Val Asn
100                105                110

Asn Thr Glu Gly Asp Trp Trp Leu Ala His Ser Leu Thr Thr Gly Gln
115                120                125

Thr Gly Tyr Ile Pro Ser Asn Tyr Val Ala Pro Ser Asp Ser Ile Gln
130                135                140

Ala Glu Glu Trp Tyr Phe Gly Lys Ile Thr Arg Arg Glu Ser Glu Arg
145                150                155                160

Leu Leu Leu Asn Pro Glu Asn Pro Arg Gly Thr Phe Leu Val Arg Glu
165                170                175

Ser Glu Thr Thr Lys Gly Ala Tyr Cys Leu Ser Val Ser Asp Phe Asp
180                185                190

Asn Ala Lys Gly Leu Asn Val Lys His Tyr Lys Ile Arg Lys Leu Asp
195                200                205

Ser Gly Gly Phe Tyr Ile Thr Ser Arg Thr Gln Phe Ser Ser Leu Gln
210                215                220

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Gln Leu Val Ala Tyr Tyr Ser Lys His Ala Asp Gly Leu Cys His Arg
 225 230 235 240
 Leu Thr Asn Val Cys Pro Thr Ser Lys Pro Gln Thr Gln Gly Leu Ala
 245 250 255
 Lys Asp Ala Trp Glu Ile Pro Arg Glu Ser Leu Arg Leu Glu Val Lys
 260 265 270
 Leu Gly Gln Gly Cys Phe Gly Glu Val Trp Met Gly Thr Trp Asn Gly
 275 280 285
 Thr Thr Arg Val Ala Ile Lys Thr Leu Lys Pro Gly Thr Met Ser Pro
 290 295 300
 Glu Ala Phe Leu Gln Glu Ala Gln Val Met Lys Lys Leu Arg His Glu
 305 310 315 320
 Lys Leu Val Gln Leu Tyr Ala Val Val Ser Glu Glu Pro Ile Tyr Ile
 325 330 335
 Val Thr Glu Tyr Met Ser Lys Gly Ser Leu Leu Asp Phe Leu Lys Gly
 340 345 350
 Glu Met Gly Lys Tyr Leu Arg Leu Pro Gln Leu Val Asp Met Ala Ala
 355 360 365
 Gln Ile Ala Ser Gly Met Ala Tyr Val Glu Arg Met Asn Tyr Val His
 370 375 380
 Arg Asp Leu Arg Ala Ala Asn Ile Leu Val Gly Glu Asn Leu Val Cys
 385 390 395 400
 Lys Val Ala Asp Phe Gly Leu Ala Arg Leu Ile Glu Asp Asn Glu Tyr
 405 410 415
 Thr Ala Arg Gln Gly Ala Lys Phe Pro Ile Lys Trp Thr Ala Pro Glu
 420 425 430
 Ala Ala Leu Tyr Gly Arg Phe Thr Ile Lys Ser Asp Val Trp Ser Phe
 435 440 445
 Gly Ile Leu Leu Thr Glu Leu Thr Thr Lys Gly Arg Val Pro Tyr Pro
 450 455 460
 Gly Met Val Asn Arg Glu Val Leu Asp Gln Val Glu Arg Gly Tyr Arg
 465 470 475 480
 Met Pro Cys Pro Pro Glu Cys Pro Glu Ser Leu His Asp Leu Met Cys
 485 490 495
 Gln Cys Trp Arg Lys Asp Pro Glu Glu Arg Pro Thr Phe Glu Tyr Leu
 500 505 510
 Gln Ala Phe Leu Glu Asp Tyr Phe Thr Ser Thr Glu Pro Gln Tyr Gln
 515 520 525
 Pro Gly Glu Asn Leu
 530

<210> SEQ ID NO 17

<211> LENGTH: 1066

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 17

Met Pro Val Phe His Thr Arg Thr Ile Glu Ser Ile Leu Glu Pro Val
 1 5 10 15
 Ala Gln Gln Ile Ser His Leu Val Ile Met His Glu Glu Gly Glu Val
 20 25 30
 Asp Gly Lys Ala Ile Pro Asp Leu Thr Ala Pro Val Ala Ala Val Gln
 35 40 45

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Ala Ala Val Ser Asn Leu Val Arg Val Gly Lys Glu Thr Val Gln Thr
 50 55 60
 Thr Glu Asp Gln Ile Leu Lys Arg Asp Met Pro Pro Ala Phe Ile Lys
 65 70 75 80
 Val Glu Asn Ala Cys Thr Lys Leu Val Gln Ala Ala Gln Met Leu Gln
 85 90 95
 Ser Asp Pro Tyr Ser Val Pro Ala Arg Asp Tyr Leu Ile Asp Gly Ser
 100 105 110
 Arg Gly Ile Leu Ser Gly Thr Ser Asp Leu Leu Leu Thr Phe Asp Glu
 115 120 125
 Ala Glu Val Arg Lys Ile Ile Arg Val Cys Lys Gly Ile Leu Glu Tyr
 130 135 140
 Leu Thr Val Ala Glu Val Val Glu Thr Met Glu Asp Leu Val Thr Tyr
 145 150 155 160
 Thr Lys Asn Leu Gly Pro Gly Met Thr Lys Met Ala Lys Met Ile Asp
 165 170 175
 Glu Arg Gln Gln Glu Leu Thr His Gln Glu His Arg Val Met Leu Val
 180 185 190
 Asn Ser Met Asn Thr Val Lys Glu Leu Leu Pro Val Leu Ile Ser Ala
 195 200 205
 Met Lys Ile Phe Val Thr Thr Lys Asn Ser Lys Asn Gln Gly Ile Glu
 210 215 220
 Glu Ala Leu Lys Asn Arg Asn Phe Thr Leu Glu Lys Met Ser Ala Glu
 225 230 235 240
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What is claimed is:

1. An observation method of a sample containing a target substance, the observation method comprising:

an imaging step in which a step of obtaining a speckle image including, as a speckle, light emitted from a luminescent substance under a prescribed condition in a state in which a medium is brought into contact with the sample is performed a plurality of times at different times respectively so as to obtain a plurality of speckle images, the medium containing a probe that contains the luminescent substance emitting light under the prescribed condition and that repeatedly binds to and dissociates from the target substance directly and specifically; and

an observation image generation step of generating an observation image of the target substance in the sample from the plurality of speckle images, wherein

a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.

2. The method according to claim 1, wherein the observation image generation step is a step in which information of a position of a speckle included in each

of the plurality of speckle images is obtained for each of the plurality of speckle images and the observation image is generated on the basis of the information from the plurality of speckle images.

3. The method according to claim 1, wherein the sample includes two or more target substances, the imaging step is sequentially performed on the sample by using the probe that is specific to each of the target substances, and

the observation image generation step is a step in which observation images of the respective target substances in the sample are respectively generated from the plurality of speckle images obtained from the respective imaging steps.

4. The method according to claim 3, further comprising a multiple-observation image generation step in which observation images of the respective target substances in the sample generated in the observation image generation step are superposed so as to generate a multiple-observation image, which is an observation image of the two or more target substances in the sample.

5. The method according to claim 1, wherein the luminescent substance is a fluorescent substance, and the prescribed condition is irradiation with excitation light.

6. The method according to claim 1, wherein

a combination between the probe and the target substance is selected from a group of:

a combination wherein the probe is (a1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 19, (a2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (a1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (a3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (a1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is an actin polymer;

a combination wherein the probe is (b1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 12, that at least partially contains an amino acid sequence of 3-309 and that has 407 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2536-2843 and that has 408 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2781-2819 and that has 138 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 1-908 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 659-908 and that has 394 or fewer amino acids, an amino acid sequence of sequence number 5, or an amino acid sequence of sequence number 6, (b2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (b1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (b3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (b1) and for which

a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is a microtubule;

a combination wherein the probe is (c1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4684 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4364 and that has 688 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4313 and that has 637 or fewer amino acids, or an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 4022-4364 and that has 443 or fewer amino acids, (c2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (c1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (c3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (c1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is an intermediate filament; and

a combination wherein the probe is (d1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 15, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-557 and that has 556 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-498 and that has 545 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 167-557 and that has 491 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino acid sequence of 1-251 and that has 351 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino

acid sequence of 3-251 and that has 349 or fewer amino acids or an amino acid sequence of sequence number 18, (d2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (d1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (d3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (d1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, and the target substance is a focal adhesion.

7. The method according to claim 1, wherein the probe contains an antibody or a fragment of an antibody, the antibody or the fragment being to the target substance and the antibody or the fragment being linked to the luminescent substance.
8. The method according to claim 7, wherein the fragment of the antibody is a Fab fragment.
9. A probe used for labeling a target substance, wherein the probe contains a luminescent substance that emits light under a prescribed condition, the probe can repeatedly bind to and dissociate from the target substance directly and specifically, and a half-life of a probe-target complex formed by binding to the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.
10. The probe according to claim 9, wherein the target substance is an actin polymer and the probe is (a1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence of sequence number 19, (a2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (a1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (a3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (a1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, the target substance is a microtubule and the probe is (b1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 12, that at least partially contains an amino acid sequence of 3-309 and that has 407 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2536-2843 and that has 408 or fewer amino acids, an amino acid sequence that is a

partial amino acid sequence of an amino acid sequence of sequence number 14, that at least partially contains an amino acid sequence of 2781-2819 and that has 138 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 1-908 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 4, that at least partially contains an amino acid sequence of 659-908 and that has 394 or fewer amino acids, an amino acid sequence of sequence number 5 or an amino acid sequence of sequence number 6, (b2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (b1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (b3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (b1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds,

the target substance is an intermediate filament and the probe is (c1) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4684 and that has 1008 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4364 and that has 688 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 3777-4313 and that has 637 or fewer amino acids or an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 8, that at least partially contains an amino acid sequence of 4022-4364 and that has 443 or fewer amino acids, (c2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (c1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (c3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (c1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or

the target substance is a focal adhesion and the probe is (d1) a polypeptide, linked to the luminescent substance,

which consists of an amino acid sequence of sequence number 15, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-557 and that has 556 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 54-498 and that has 545 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 15, that at least partially contains an amino acid sequence of 167-557 and that has 491 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino acid sequence of 1-251 and that has 351 or fewer amino acids, an amino acid sequence that is a partial amino acid sequence of an amino acid sequence of sequence number 16, that at least partially contains an amino acid sequence of 3-251 and that has 349 or fewer amino acids, or an amino acid sequence of sequence number 18, (d2) a polypeptide, linked to the luminescent substance, which consists of the amino acid sequence described in (d1) where one or a plurality of amino acids have been substituted, deleted, inserted or added and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds, or (d3) a polypeptide, linked to the luminescent substance, which consists of an amino acid sequence having at least a 70% identity with the amino acid sequence described in (d1) and for which a half-life of a probe-target complex formed by binding between the probe and the target substance is equal to or more than 10 milliseconds and equal to or less than 3 seconds.

11. The probe according to claim 9, wherein the probe contains an antibody or a fragment of an antibody, the antibody or the fragment being to the target substance and the antibody or the fragment being linked to the luminescent substance.
12. The probe according to claim 11, wherein the fragment of the antibody is a Fab fragment.
13. A reagent kit for labeling a target substance, wherein the reagent kit at least includes the probe according to claim 9.
14. A screening method of a site which identifies a target substance in the probe according to claim 9, the screening method comprising:
- an immobilization step in which a candidate substance of the site or a substance partially containing the candidate substance is fixed to a solid support;
 - an observation step in which a target substance linked to a luminescent substance and a solid support obtained in the immobilization step are observed in a medium while the target substance linked to a luminescent substance and the solid support obtained in the immobilization step are kept in contact, in a condition that allows observation, in units of 1 molecule, of light emission from the luminescent substance in a probe-target complex formed by binding between the target substance and the candidate substance, and
 - a screening step in which the candidate substance resulting in a half-life of the probe-target complex that is equal to or more than 10 milliseconds and equal to or less than 3 seconds is selected as the site on the basis of observation in the observation step.
15. The method according to claim 14 wherein the candidate substance is an antibody or a fragment of an antibody from a library of hybridoma that produces an antibody to the target substance, and the antibody is fixed to a solid support in the immobilization step.

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